



Influence of Endometrial Microbiota on Reproductive Outcomes in IVF Programs

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Abstract

Introduction: Implantation failures are one of the causes of the inefficiency of IVF programs. Recent studies have shown that in humans, some biotopes such as the uterus and placenta, previously considered sterile, are colonized by their unique microbiota. Disturbance of the uterine microbiota can be one of the reasons for the decline in the effectiveness of assisted reproductive technology programs.

Materials and methods: A total of 80 patients with infertility were enrolled in the study: group I included the patients with fresh embryo transfer and group II included the patients with frozen-thawed embryo transfer. After routine embryo transfer, the most distal 5 mm portion of the embryo transfer catheter was placed in the sterile container. A study was conducted using culture-based methods with extended nutrient media.

Results: In patients with fresh embryo transfer in ovarian hyperstimulation cycle, 28 species of microorganisms were detected. Pregnancy in this group occurred in 20 women (40 %). The microflora was dominated by *Lactobacillus* spp. (90% of women). In patients with frozen-thawed embryo transfer, 20 species of microorganisms were detected. Pregnancy in this group occurred in 13 women (43.3 %). The microflora was dominated by *Lactobacillus* spp. (69.2 % of women). *Bifidobacterium* spp. were significantly more frequently detected in patients of group II than in those of group I (23.3% and 2%, respectively).

Conclusion: The evidence of this study suggests that the uterus is not sterile, the uterine microbiota may differ from the cervical canal microbiota, the uterine microbiota is one of the predictors of the success of embryo implantation.

Keywords: Infertility; Endometrium; Uterine microbiota; Embryo transfer

Introduction

Implantation failures are one of the causes of the inefficiency of IVF programs. Recent studies have shown that in humans, some biotopes such as the uterus and placenta, previously considered sterile, are colonized by their unique microbiota. Disturbance of the uterine microbiota can be one of the reasons for the decline in the effectiveness of assisted reproductive technology programs [1-4]. It is known that microorganisms affect the immune system. Accordingly, the effect of microorganisms on the immune environment before pregnancy is associated with the endometrial receptive potential [5,6]. Successful embryo implantation depends on endometrial condition at the moment of the "implantation window". Endometrial susceptibility deterioration during the menstrual cycle leads to impaired placentation and infertility [7]. At the same time, the commensal microbiota can provide the protection against pathogenic microorganisms and contribute to the maintenance of

homeostasis in the uterine cavity [7,8]. The purpose of our research is to evaluate the effect of uterine microbiota on the success of embryo implantation in women in assisted reproductive technology programs.

Materials and Methods

A total of 80 patients with infertility were enrolled in the study: group I included the patients with fresh embryo transfer and group II included the patients with frozen-thawed embryo transfer. The average age of women was ± 31.4 years. After routine embryo transfer, the most distal 5 mm portion of the embryo transfer catheter was placed in the sterile container. A study was conducted using culture-based methods with extended nutrient media. In patients with implantation failures in history, 16S RNA sequencing is planned. In order to exclude other negative factors and to make

more accurate assessment of the role of microbiota in reproductive outcomes, we included women who were transferred embryos of good and excellent quality both in fresh embryo transfer and frozen-thawed embryo transfer.

Results

It was established that the uterine cavity in women of groups I and II was sterile in 12.0% and 13.3% of cases, respectively. The selected microflora in the comparison groups was characterized by the presence of monocultures (28% of women in group I and 30% in group II) and associations of microorganisms (52.9% and 56.6%, respectively). The most represented species in the microbiota were Lactobacillus, their isolation frequency was 80% и 66.6 %. Moreover, these microorganisms were present in the monoculture in 26% of patients in group I and in 20% in group II; they were detected in association with each other in 14% and 13.3%, and in association with opportunistic microorganisms in 38% and 20% of cases, respectively. The opportunistic microorganisms were the second most frequent component of the microbiota in 48% of women in group I and in 40% of women in group II. In the monoculture, opportunistic microorganisms were isolated in 2% and 3.3% of women, associations of only opportunistic

microorganisms were revealed in 8% and 6.6% of women, associations of opportunistic microorganisms with Lactobacillus spp. and Bifidobacterium spp. were detected in 38 % and 29.9 % of women, respectively. Bifidobacterium spp. were significantly more frequently detected in patients of group II than in those of group I (23.3% and 2%, respectively).

In patients with fresh embryo transfer in ovarian hyperstimulation cycle, 28 species of microorganisms were detected (Table 1). Pregnancy in this group occurred in 20 women (40 %). The microflora was dominated by Lactobacillus spp. (90% of women): *L. jensenii* (40%), *L. crispatus* (35%). The most common opportunistic microorganisms (50 %) were *Gardnerella vaginalis* (*G. vaginalis*) and *Enterococcus faecalis* (*E. faecalis*) (20%), *Streptococcus anginosus* (*S. anginosus*) (15%). In non-pregnant women (60 %) the frequency of isolation of Lactobacillus spp. was lower than in pregnant women (73.3%), but *L. jensenii* (53.3%) and *L. crispatus* (26.6%) were dominating. Among opportunistic microorganisms, *G. vaginalis* (10%) and optional anaerobes *S. anginosus* (10%) and *Staphylococcus haemolyticus* (6.6%) were most frequently detected. It should be noted that *Haemophilus influenzae* (3.3%) and *Actinomyces* sp. (6.6%) which may be seldom found in the reproductive organs were also revealed.

Table 1: Detected species of microorganisms.

Patients with fresh embryo transfer, n=50	Patients with frozen-thawed embryo transfer, n=30
Lactobacillus spp.:	Lactobacillus spp.:
1. <i>Lactobacillus jensenii</i>	1. <i>Lactobacillus jensenii</i>
2. <i>L. crispatus</i>	2. <i>L. crispatus</i>
3. <i>L. gasseri</i>	3. <i>L. gasseri</i>
4. <i>L. oris</i>	4. <i>L. iners</i>
5. <i>L. vaginalis</i>	5. <i>L. vaginalis</i>
6. <i>L. johnsonii</i>	6. <i>L. fermentum</i>
7. <i>L. reuteri</i>	7. <i>L. reuteri</i>
	8. <i>L. paracasei</i>
Opportunistic microorganisms:	Opportunistic microorganisms:
8. <i>Escherichia coli</i>	9. <i>Escherichia coli</i>
9. <i>Proteus mirabilis</i>	10. <i>Enterococcus faecalis</i>
10. <i>Staphylococcus epidermidis</i>	11. <i>Staphylococcus epidermidis</i>
11. <i>S. hominis</i>	12. <i>S. warneri</i>
12. <i>S. capitis</i>	13. <i>Streptococcus agalactiae</i>
13. <i>S. haemolyticus</i>	14. <i>S. anginosus</i>
14. <i>Enterococcus faecalis</i>	15. <i>Actinomyces neuii</i>
15. <i>Streptococcus agalactiae</i>	16. <i>Micrococcus luteus</i>
16. <i>S. salivarius</i>	17. <i>Propionibacterium avidum</i>
17. <i>S. anginosus</i>	
18. <i>Micrococcus luteus</i>	
19. <i>Haemophilus influenzae</i>	
20. <i>Atopobium vaginae</i>	
21. <i>Veillonella atypica</i>	
22. <i>Actinomyces radingae</i>	

23. <i>Actinomyces neuii</i>	
24. <i>Dialister microaerophilus</i>	
25. <i>Propionibacterium avidum</i>	
26. <i>P. acnes</i>	
27. <i>Gardnerella vaginalis</i>	
Bifidobacterium spp.	Bifidobacterium spp.
28. <i>Bifidobacterium bifidum</i>	18. <i>Bifidobacterium bifidum</i>
	19. <i>B. dentium</i>
	20. <i>B. longum</i>

In patients with frozen-thawed embryo transfer, 20 species of microorganisms were detected (Table 1). Pregnancy in this group occurred in 13 women (43.3%). The microflora was dominated by *Lactobacillus* spp. (69.2 % of women): *L. jensenii* (46.2%), *L. crispatus* (23%). The most common opportunistic microorganisms (38.5 %) were optional anaerobes *Escherichia coli* (15.4%) and *Staphylococcus epidermidis* (*S. epidermidis*) (15.4%). In non-pregnant women (56.7 %) the frequency of isolation of *Lactobacillus* spp. was lower than in pregnant women (64.7%); *L. jensenii* (41.2%) and *L. crispatus* (23.5%) were dominating. Among opportunistic microorganisms, optional anaerobes *S. epidermidis* (17.6%) and *E. faecalis* (11.8%) were most frequently detected. However, *Bifidobacterium* spp. were detected in 35.3% of women, and *Actinomyces neuii* were found in 5.9% of women.

Discussion

The results of the current study are consistent with those of M. Mitchell and E. Cicinelli who suggested that the uterine cavity is not sterile [9,10]. Moreno I. et al. performed the analysis of the microbiota in paired samples of endometrial fluid and vaginal aspirates obtained simultaneously from 13 fertile woman in pre-receptive and receptive phases within the same menstrual cycle (a total of 52 samples were analyzed). To investigate the hormonal regulation of the endometrial microbiota during the acquisition of endometrial receptivity, endometrial fluid was collected at pre-receptive and receptive phases within the same cycle from 22 fertile women (a total of 44 samples were analyzed). Finally, the reproductive impact of an altered endometrial microbiota in endometrial fluid was assessed by implantation, ongoing pregnancy, and live birth rates in 35 infertile patients undergoing IVF (a total of 41 samples were analyzed); receptive endometrium was assessed using the endometrial receptivity array. When bacterial communities from paired endometrial fluid and vaginal aspirate samples within the same subjects were interrogated, different bacterial communities were detected in the uterine cavity and the vagina in some subjects.

Based on its composition, the microbiota in the endometrial fluid, comprising up to 191 operational taxonomic units, was defined as *Lactobacillus*-dominated microbiota (>90% of *Lactobacillus* spp.) or a non-*Lactobacillus*-dominated microbiota (<90% *Lactobacillus* spp. with >10% of other bacteria). Although the endometrial microbiota was not hormonally regulated during the acquisition of endometrial receptivity, the presence of a non-*Lactobacillus*-dominated microbiota in a receptive endometrium was associated with a significant decrease in implantation (60.7%

vs 23.1%), pregnancy (70.6% vs 33.3%), ongoing pregnancy (58.8% vs 13.3%), and live birth (58.8% vs 6.7%) rates [11]. Endometrial microbiome in infertile patients was assessed in the study of [12] The endometrial microbiome during the embryo transfer was characterized by analyzing catheter tips with Illumina V4 metagenomics in 70 patients who underwent IVF cycles. *Lactobacillus* spp. were detected in all 70 samples. Among 70 samples, 33 samples contained over 90% of *Lactobacillus* abundance and 50 samples contained over 70% of *Lactobacillus* abundance; this finding was consistent with the dominance of *Lactobacillus* spp. in the lower reproductive tract.

Other vaginal bacteria were also identified from the transfer catheter tips. *Corynebacterium* spp. were detected in 40 out of 70 patients, *Bifidobacterium* spp. were detected in 15 out of 70 patients and *Staphylococcus* spp. were detected in 38 out of 70 patients. Other lactic acid-producing bacteria from the genera *Streptococcus* spp. were present in 38 out of 70 patients. Thus, our findings suggest the existence of a functional system "microbiota - endometrium", which is characterized by the dominance of *Lactobacillus* spp. As a result of various adverse factors, uterine cavity homeostasis may be impaired, which subsequently affects the reproductive outcomes [12].

Conclusion

The evidence of this study suggests that the uterus is not sterile, the uterine microbiota may differ from the cervical canal microbiota, the uterine microbiota is one of the predictors of the success of embryo implantation.

The study showed that among 33 women who became pregnant, the best results of implantation were observed during the colonization of the uterus only with *Lactobacillus* spp. both in fresh and frozen-thawed embryo transfers. The overall pregnancy rate was 36.4% after the colonization of the uterine cavity with association of opportunistic microorganisms with *Lactobacillus* spp. Provided the sterile uterine cavity, pregnancy occurred in 9.1% of cases (n = 3). Therefore, the presence of *Lactobacillus* spp. in the uterus increases the likelihood of pregnancy. At the same time, the high frequency of isolation of *Bifidobacterium* spp. in non-pregnant women requires clarification of the possible negative effect of these microorganisms on implantation in a larger population of women.

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