



Nutraceutical Potential of *Parkia speciosa* (Stink Bean): A Current Review

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Abstract

Background: Inadequate fruits and vegetables intake contributes to the prevalence of major diseases such as cardiovascular diseases and cancers. *Parkia speciosa* [stink bean] is a common vegetable consumed in the Southeast Asia. Although it contains various phytochemicals that can help prevent disease development, the effort to develop a specific treatment or food products from *Parkia speciosa* remains a challenge. Here, we explore research works of the medicinal benefits of *P. speciosa* that can be used as a guide to develop future clinical studies.

Method: We conducted a database search on PubMed, Google Scholar, and Science Direct using the keywords “nutraceutical potential”, “*Parkia speciosa*” “antioxidant”, “hypoglycemic”, “antitumor”, “antimicrobial” and “cardiovascular effects”. We included clinical trial, *in vitro* and *in vivo* studies that were written in English or Malay; and excluded review articles with no time limitations.

Result: We reviewed a total of 28 research articles. No clinical trial was found. The articles were grouped into antioxidative, hypoglycemic, antitumor, antimicrobial and cardiovascular effects. Six articles had combination of the medicinal properties. Seeds and empty pods are the most common plants parts used. Each bioactivities differed depending on the plant parts, extracts, methods, cultivar and plantation site.

Conclusion: *P. speciosa* demonstrated antioxidative, hypoglycemic, antitumor, antimicrobial and cardiovascular effects that were contributed by its phytochemical compounds. This finding could be used as a database for future clinical studies. We recommended researchers to use the information from the articles reviewed for drug development and clinical trial.

Keywords: Nutraceutical potential; *Parkia Speciosa*; Antioxidant, Hypoglycemic; Antitumor; Antimicrobial; Cardiovascular Effects.

Background

P. speciosa or stink bean is a classic Malaysian favourite and is commonly grown and cultivated in Southeast Asia e.g. Indonesia, Malaysia, and some parts of North-Eastern India [1]. *P. speciosa* is also known as ‘petai’ in Malaysia, Singapore, and Indonesia [2], ‘sator’ or ‘sataw’ in Thailand [3], ‘u’pang’ in Philippines [2], and ‘yongchak’ in India [4]. *P. speciosa* earned its nickname ‘stink bean’ from its strong and pungent odour. The plant belongs to the pea or bean family Fabaceae and placed in *Leguminosae* and *Mimosaceae* [4]. *P. speciosa* contained several phytochemical compounds such as polyphenols and flavonoids [5,6], terpenoids [5,7,8], Alkaloids [5,7-10], saponins [7-10], steroids [7,8,10], tannins [7,9] and phytosterol [11,12]. Globally, the World Health Organization [WHO] reported that about 80% of the world population relies on traditional med

icine to cure ailments [13]. In Malaysia, *P. speciosa* has been used traditionally to treat various diseases and ailments such as hypertension [4] and kidney disorders [14]. There is a limited data of medicinal benefits of *P. speciosa* especially on the cardiovascular and antioxidant effects. From the previous review articles, only two [15,16] research experiments were included for cardiovascular effects; and only seven [15] and nine [16] research experiments were included for antioxidant effects. The findings of the current review article could be used as a database for future clinical studies. This review aimed to explore the medicinal benefits of *P. speciosa* as to provide the foundational knowledge on the topic; so that the pre-clinical and clinical studies can be conducted for future drug development.



Methods

We conducted a literature search for medicinal benefits of *P. speciosa* using PubMed, Google Scholar, and Science Direct using the keywords “medicinal benefits”, “*Parkia speciosa*” “antioxidant”, “hypoglycemic”, “antitumor”, “antimicrobial” and “cardiovascular effects”. We included clinical trials, *in vitro* and *in vivo* studies; and exclude review articles. The literature search was limited to articles published in Malay and English without time limitations.

Results and Discussion

We found a total of 28 eligible articles: 23 *in vitro* and 5 *in vivo* studies. No clinical trial was found on this topic. Of the articles reviewed, 14 articles reported on antioxidant activity, 5 articles on hypoglycemic activity, 5 articles on antitumor activity, 6 articles on antimicrobial activity and 4 articles on cardiovascular effects. There were combinations of bioactivities studied in six articles reviewed: one article documented on the antioxidant, hypoglycemic and antimicrobial activities [6], one article performed research on antioxidant and antitumor effects [17], one article reported on antioxidant and antimicrobial effects [18], two studies described both the antioxidant and antimicrobial activities [5,18] and one study combined antioxidant and cardiovascular effects [19].

Antioxidant activity

A total of 13 *in vitro* and 1 *in vivo* study on the antioxidative property of *P. speciosa* were reviewed. In most of the studies cited, the seeds of *P. speciosa* were used [5,19-22] (Table 1). Other studies analysed empty pods [6,17,23,24], pods [9,18] and seed coat and pericarp of the *P. speciosa* bean [25]. The commonly used tests for antioxidative activity are 1,1-diphenyl-2-picrylhydrazyl free radical [DPPH] scavenging and reducing ferric ion antioxidant potential [FRAP] assay. Other assays used are anti-lipid peroxidation, superoxide radical scavenging activity, 2,2'-azino-bis [3-ethylbenzothiazoline-6-sulfonic acid] [ABTS] radical scavenging activity and metal chelating activity. The relationship between total phenolic and flavonoid contents with antioxidant level was evaluated in four studies [5,20,22,26]. Two studies examined the antioxidant level indirectly [21,25], whereby the antioxidant level was related to hydrogen sulphide [H₂S] in one study, and the other to Heinz body inhibition [aggregation of denatured hemoglobin in the red blood cell resulting from oxidative process]. Four studies compared the antioxidant level between *P. speciosa* and other plants [20,22,26]. Generally, the antioxidant level varies depending on the part of the plant, the extraction variables [solvent-extract ratio, time, temperature and solvent type] and the plantation location.

In vitro studies

In a study by Balaji and co-workers [9], the flavonoids and phenolic content, as well as the antioxidant activity were evaluated. The flavonoids content estimation was carried out using the calorimetric method, while the phenolic content was carried out using the Folin-Ciocalteu reagent method. Reducing power assay and DPPH

were used to examine the antioxidant activity. The study found that *P. speciosa* pod powder extract had 14.16±0.02 mg gallic acid equivalents per gram [GAE/g] dry weight total phenolic content and 5.28±0.03 mg rutin equivalents per gram [RE/g] dry weight total flavonoid content. Generally, the phenolic and flavonoid compounds are related to antioxidant activity of the plant. The extract also showed potent antioxidant activity as demonstrated by IC₅₀ values of the extract in DPPH, 74.37µg/ml compared to the standard butylatedhydroxy toluene [BHT] 35.40 µg/ml. IC₅₀ indicates the concentration of test extracts or positive controls that inhibit or scavenge the radical formation by 50% [23]. Although the DPPH radical scavenging activity was lower than BHT, it was evident that the extract contained a substance/s with proton-donating ability and was capable of inhibiting free radicals.

Ko and colleague [23] evaluated the antioxidant activities of aqueous and ethanolic extracts of *P. speciosa* empty pods using several assays namely: anti-lipid peroxidation, superoxide radical scavenging activity, DPPH radical scavenging activity, ABTS radical scavenging, metal chelating and reducing power. It showed that the ethanol extracts possessed stronger antioxidant activity via all the assays except for superoxide radical scavenging activity. This finding is influenced by a higher level of polyphenols [phenols and flavonoids] in the ethanolic extract. The anti-lipid peroxidation IC₅₀ was 5.02±1.06 µg/ml, DPPH radical scavenging activity was 64.2±3.46 µg/ml, ABTS radical scavenging 19.6±0.44 µg/ml, metal chelating activity 319 ± 26.3µg/ml and reducing power activity 274±16.1µg/ml. The extracts contained several polyphenolic constituents, the most abundant of which were gallic acid, catechin, ellagic acid and quercetin.

Two types of *P. speciosa* in Thailand ['Sataw-Khao' and 'Sataw-Dan'] were studied by Wonghirundecha and co-workers [18] for their total phenolic content, antioxidant and antimicrobial activities. It was found that the extraction yield, total phenolic and total flavonoid contents of 'Sataw-Dan' pod extracts were higher than that of 'Sataw-Khao' pod extracts. In contrast, 'Sataw-Khao' pod extracts showed higher DPPH [1218.07± 8.72 µmol Trolox equivalent/g dry weight [TE/g] vs 920.32±6.15 µmol TE/g dry weight], ABTS radical scavenging activity [1610.67±11.88µmol TE/g dry weight vs 1261.14±17.44µmol TE/g dry weight] and metal ion chelating activity [9.76±0.03 Ethylenediaminetetraacetic acid equivalent/g dry weight [EDTAE/g] vs. 5.86±0.02 EDTAE/g dry weight] compared to Sataw-Dan pod extracts. Thus, the authors concluded that there was no relationship between total phenolic content and antioxidant activity in the PS extract, suggesting that other phytochemicals apart from polyphenols may contribute to the antioxidant activity. In addition, both extracts showed antimicrobial effect in the form of inhibition zone formation through the agar well diffusion assay.

In a study by Aisha and co-workers [17], eight empty *P. speciosa* pod extracts were examined for phenolic content and anti-

oxidant, cytotoxic and antiangiogenic activities. The results of the study showed that the methanolic sub-extract had the highest total phenolic content, 25.55 ± 1.57 GAE/100 mg. In the DPPH scavenging assay, antioxidant activity was also highest in the methanolic sub-extract, [IC₅₀ 26 ± 3.0 µg/ml]. It can be visualized from all the extracts used that; the antioxidant level corresponded to the total phenols content.

Sonia and co-workers [6] further explored the antioxidant, anti-inflammatory, anti-diabetic and anti-microbial activity of *P. speciosa*. The FRAP value of the hydromethanolic extract was higher than of the ascorbic acid controls (1.9mM Ferrous sulfate (FESO₄)). Its strong antioxidant activity was supported by DPPH study, as it significantly decreased *in vitro* DPPH radical concentration (64.52 ± 2.4 % inhibition, IC₅₀ 315.75 µg/ml), and hydrogen peroxide (H₂O₂) assay [78.06 ± 5.7 % inhibition, IC₅₀ 166.3 µg/ml]. The extract exhibited anti-inflammatory properties via inhibition of lipooxygenase activity (38.6 ± 10.2 % inhibition, IC₅₀ 493.34 µg/ml; control 56.9 ± 11.4 % inhibition, IC₅₀ 280.71 µg/ml), proteinase inhibitory activity (22.78 ± 3.6 % inhibition, IC₅₀ 1142.3 µg/ml; control 29.9 ± 5.9 % inhibition, IC₅₀ 53.75 µg/ml) and RBC membrane stabilization activity (99.21 ± 12.6 % inhibition, IC₅₀ 67.01 µg/ml; control 99.36 ± 6.7 % inhibition, IC₅₀ 53.75 µg/ml).

There is increasing demand for new ingredients from natural sources in the food industry. Moreover, there are suggestions to use agrowaste materials [in this case, *P. speciosa* empty pods] as functional food ingredients. Gan and Aishah [24] studied the physicochemical properties' characterization, looking at the antioxidant property of *P. speciosa* as potential functional flour. Between two methods of drying [freeze dried and oven dried], different functional properties can be seen. Higher antioxidative properties were shown in freeze dried *P. speciosa* pod (FDPSF) which consist total phenolic content (TPC) of 110.0mg GAE/g sample and total flavonoid content (TFC) of 8.5 mg pyrocatechol equivalents/g sample. These extracts [pre-diluted 50x] gave %DPPHsc, %ABTSsc and FRAP values of 65.3%, 77.4% and 1.9mM FESO₄, respectively. Therefore, *P. speciosa* powder can be used as a substitute for commercial flour.

Five Malay raw salads namely: leaves of *Cosmos caudatus* ('Ulam Raja'), *Oenanthe javanica* ('Selom'), *Murraya koenigii* [curry leaf], *Centella asiatica* ('Pegaga') and the seeds of *P. speciosa* ('Petai') were studied by Reihani and Azhar [20] for their total phenolic content and antioxidant activities. The total phenolic content [mg GAE/g of plant on dry basis] were highest in *Murraya koenigii* (33.18) and lowest in *P. speciosa* (6.45). Pertaining to antioxidant property, *Cosmos caudatus* had the highest DPPH free radical scavenging activity (212.8) and *Centella asiatica* the lowest (32.4). *Oenanthe javanica* and *Cosmos caudatus* demonstrated the highest ferric reducing activities: 199.96 µmol TE/g and 183.11 µmol TE/g, respectively; while *P. speciosa* showed the lowest (44.67 µmol TE/g). It was found in this study that there was no significant correlation between antioxidant activity and total phenolic content.

This may be due to steric hindrance and presence of other reducing agents in the extracts studied.

Siow and Gan [19] examined the antioxidative bioactive peptides from *P. speciosa* seeds using alcalase. Prior to extraction, the *P. speciosa* seeds protein hydrolysate was analysed for amino acid composition. Glutamine and aspartic acid were found in the highest amounts, followed by cysteine (154, 132, 84.8 per 1000 amino acid residuals, respectively). Glutamine and aspartic acid are strong antioxidants as they act as electron donors [27]. In this study, the effects of temperature, substrate-to-enzyme ratio and incubation time were taken into consideration. The highest DPPH free radical scavenging activity and FRAP activity (2.9 mg GAE/g and 11.7mM FeSO₄, respectively) were demonstrated in a condition of 50°C, substrate/enzyme (S/E) ratio of 50 and 2 hours incubation time. The high temperature causes unfolding of the protein molecules thus making the protein active site more accessible to the enzyme and exposes the protein donating residues. Partial hydrolysis of the proteins is essential to give higher DPPH value, compared to extensive protein hydrolysis caused by higher enzyme concentration. The DPPH values increased along with the incubation time. Upon fractionation of the protein hydrolysates that had the highest DPPH free radical scavenging activity and FRAP activity, the peptide fraction of <10k Da showed the strongest bioactivities. Using advanced mass spectrometry, 29 peptide sequences were identified to be responsible for the potent bioactivities, that could be developed into novel nutraceuticals.

The production of phytochemicals in plants depends on the variety or species and external variables such as environmental conditions, agricultural practices and post-harvest handling [5]. A study by Ghasemzadeh and co-workers [5] compared the phytochemical constituents and biological activities of *P. speciosa* collected from 3 regions of Malaysia [Perak, Negeri Sembilan and Johor]. The result showed that the seeds (ethanol extract) collected from Perak contained highest phytochemical content at concentration of 100 µg/mL, DPPH (66.29%) and FRAP (522.1 µM of Fe (II)/g) scavenging activity, followed by Negeri Sembilan and Johor. The authors analyzed the correlation between the parameters studied and found that the antioxidative activity was significantly correlated with flavonoid content.

In another study, Maisuthisakul and co-workers [26] analyzed the correlation between parameters [yield, radical scavenging activity 1/EC₅₀, phenolic compounds, flavonoids, moisture, ash, protein, fat, carbohydrate, dietary fibre, calcium, iron and vitamin C] of 28 Thai plants. EC₅₀ is the concentration of extract necessary to decrease DPPH radical scavenging by 50%. Generally, the seeds of the plants studied had high energy, and *P. speciosa* seeds were among the plants with the highest energy content (441.5 Kcal/100g edible portion, db). It showed antiradical activity of 1.5 (1/EC₅₀), total phenolics of 51.9 mg GAE/g db and total phenolics of 20.3mg RE/g db. The yield of extractable compounds ranged from 0.6%

to 4.5%. The yields from berries, fruit and seeds were higher than from other parts of the plants. In terms of correlation analysis, plants with high yield had high carbohydrate content and low fibre content. A high yield did not correspond to high content of phenolic compounds and high antioxidant activity. The study also revealed that plants with strong antioxidant activity had high total phenolic and flavonoid contents.

Interestingly, Liang and co-workers [21] investigated the hydrogen sulphide [H_2S] releasing capacity of ten organosulphur rich fruits and vegetables [garlic, red onion, yellow onion, scallion, shallot, leek, spring onion, Chinese chives, durian and stinky beans]. H_2S is a gaseous signaling molecule that has several effects on human health including antioxidant effect [28]. To demonstrate the importance of this gas, the authors quoted previous studies to investigate how garlic has been used as an herbal remedy for thousands of years and having cardioprotective effect. Benavides and co-workers [29] from their work in rats proposed that the two major components of garlic extract, namely diallyl trisulphide [DATS] and diallyl disulphide [DADS] are converted to H_2S resulting in relaxation of the rat aorta ring. Similarly, Chuah and co-workers [30] showed that through H_2S -dependent mechanism, S-allyl cysteine, the major compound of aged garlic was able to protect against myocardial infarction. In the study by Liang and colleagues [21], the H_2S releasing capacity of plant essential oils was evaluated by fluorescent method using BCu [H_2S selective and sensitive turn-on fluorescent probe] as probe and human breast cancer MCF-7 cells as the medium. MCF-7 cells incubated with BCu for 3 hours were treated with H_2S donors to produce H_2S . Fluorescence produced by H_2S captured by the probe was measured by a microplate reader. The results showed that *P. speciosa* had the highest DATS equivalent (DATS-E) value (158 mmol DATS/kg of raw material), followed by garlic (18.5 mmol DATS/kg of raw material) and yellow onion (4.59 mmol DATS/kg of raw material). DATS-E value is a concept introduced in which each plate was divided into two zones; one zone loaded with different concentrations of samples, and the other zone loaded with different concentration of DATS. This method was performed to enable data comparison among different groups. Among the reasons *P. speciosa* had the highest DATS-E value was because of its high sulphur content, and presumably its cyclic sulphur compounds could be converted to H_2S more efficiently than linear compounds. This study indirectly relates the presence of H_2S in *P. speciosa* to antioxidant bioactivities.

Twenty five edible tropical plants were examined by Wong and co-workers [22] for their antioxidant properties. The total polyphenol contents, free radical scavenging, ferric ion reducing (expressed as TEAC-Trolox equivalent antioxidant capacity) and cupric ion chelating capabilities (CCA) were determined. The result showed that *P. speciosa* leaves extract had the highest TPC but showed low TEAC_{DPPH} and TEAC_{FRAP}. The non-correspondence between TPC and antioxidant activities is due to the fact that *P. speciosa* may contain other compounds that can oxidize the Folin-Ciocalteu, other than

polyphenols. In addition, *P. speciosa* leaf extract chelated the cupric ions the most. This finding indicated that the extract had potential secondary compounds. TPC showed satisfactory correlation with TEAC_{DPPH} and TEAC_{FRAP}, indicating that polyphenols in the extracts were partly responsible for the antioxidant activities. Conversely, TPC correlated poorly with CCA, indicating that polyphenols might not be the main cupric ion chelators.

Indirect relationship between inhibitory activity of Heinz body induction and antioxidant activity was studied by Tunsaringkarn and co-workers [25]. Hemoglobin chains undergo denaturation process through oxidative damage by reactive oxygen species and produced Heinz body, an aggregation of denatured and precipitated hemoglobin within red blood cells [31]. Hence it can be a biomarker for oxidative damage in the body. In this study, *P. speciosa* seed coat extracts showed the highest activity of Heinz body inhibition (39.98%, IC25 2.68 mg/ml), followed by *X. xylocarpa* bark (42.75%, IC25 15.71 mg/ml), *P. speciosa* Hassk. pericarp (44.89%, IC25 28.14 mg/ml) and *E. rheedii* Spreng. seed coat (55.12%, IC25 33.20 mg/ml) *X. xylocarpa* bark, *E. rheedii* seed coat, Hassk seed coat, and *X. xylocarpa* stem contained high tannin concentration. It has been shown that the percentage of Heinz body inhibition was correlated to tannin concentration. The previous study suggested that the antioxidant activity of tannin was mainly due to iron chelation rather than hydroxide radical scavenging [32]. This finding also supported the results of the study by Wong and co-workers [22] discussed before.

In vivo study

Al Batran and co-workers [1] studied antioxidant and antiulcer activity of *P. speciosa* ethanolic leaf extract in ethanol-induced gastric ulcer in rats. Sprague Dawley rats were divided into 7 groups: Groups 1 and 2 received 0.5% carboxymethylcellulose (CMC) as vehicle; Group 3 received 20mg/kg omeprazole and groups 4-7 received ethanolic leaves extract of *P. speciosa* at doses of 50, 100, 200 or 400 mg/kg. After 1 hour, CMC or absolute ethanol was administered to groups 2-7. The rats were euthanized after 1 hour and the gastric mucosa was examined for gastric juice acidity, gastric wall mucus, macroscopic gastric lesion evaluation, antioxidant activity, histological examination and immunohistochemical staining. Regarding the antioxidant activity, gastric tissue homogenate prepared from the groups that were treated with plant extract displayed significant antioxidant activity, with decreased levels of malondialdehyde (MDA) and elevated levels of total glutathione (GSH) and superoxide dismutase (SOD), in response to oxidative stress due to ethanol treatment. MDA is the final product of lipid peroxidation which causes a loss of membrane fluidity, impaired ion transport and membrane integrity resulting in loss of cellular function [33]. It was found that the groups treated with the plant extract reduced this process. GSH plays a role in determining ulcer severity and acts as tissue protective agent [34,35]. SOD converts superoxide to hydrogen peroxide (H_2O_2), which is transformed into

water by catalase in the lysosomes or by glutathione peroxidase in the mitochondria [36].

Hypoglycemic Activity

There were 5 articles that demonstrated the hypoglycemic property of *P. speciosa*, 2 *in vitro* and another 3 *in vivo* studies. One of the *in vitro* hypoglycemic studies has been discussed earlier by Sonia and co-workers [6] as part of their study. Two studies examined empty pods and seeds separately [37,38], one study examined seeds and pericarp separately [39] and two studies examined empty pod [6,11]. Among the assays used were α -glucosidase inhibition activity, alpha amylase inhibition activity, and porcine pancreatic lipase (PPL) inhibition assay and glucose oxidase method. All five studies exhibited a strong hypoglycemic activity of the plant. Two studies explored the compounds responsible for the hypoglycemic activity [11,38]. There was some inconsistency on parts of the plant that gives greater hypoglycemic effect. Tunsaringkarn and colleagues [39] found that the *P. speciosa* pericarp had a higher hypoglycemic activity than the seeds, but Jamaluddin and colleagues [37,38] had opposite findings. This inconsistency could be due to the cultivar, harvesting time and method, types of extract and assays performed.

In vitro study

A study by Tunsaringkarn and colleagues [39] evaluated twenty species of Thai plants of Mimosaceae family. α -glucosidase is the enzyme responsible for digestion of polysaccharide and oligosaccharide to monosaccharides [40]. Using α -glucosidase inhibition assay, *P. speciosa* pericarp was one of the plants that showed high inhibition activity; IC₅₀ 0.0581 mg/ml (89.46%). As a comparison, other plants had α -glucosidase inhibition of: *Entada rheedii* seed coat IC₅₀ 0.0043, *Archidendron jiringa* seed coat IC₅₀ 0.0054, *Albizia lebbek* branch bark IC₅₀ 0.0397 and *Albizia lebbekoides* bark IC₅₀ 0.0702mg/ml. *P. speciosa* seed showed lower percentage of α -glucosidase inhibition activity (45.72%).

Sonia and co-workers [6] studied the anti-diabetic activity of *P. speciosa* empty pods via two assays: alpha amylase inhibition and porcine pancreatic lipase (PPL) inhibitory assay. The alpha amylase acts to hydrolyze dietary starch [maltose] to glucose. In this study, the researchers explored how the *P. speciosa* pod extract can inhibit alpha amylase resulting in reduction of post prandial hyperglycemia. It was found that *P. speciosa* showed a maximum inhibition of 79.2% at 500 μ g/ml. The extract exhibited anti-diabetic property as the IC₅₀ of *P. speciosa* were found to be 199.29 μ g/ml compared to standard acarbose (324.18 μ g/ml). From another aspect, pancreatic lipase functions to digest dietary fat, specifically hydrolyzing triacylglycerol to 2-monoacylglycerol and fatty acids. Through PPL inhibitory assay, the maximum inhibitory activity of *P. speciosa* extract at 500 μ g/ml was 89.5%; IC₅₀ 196.61 μ g/ml compared to Orlistat, 76.3%; IC₅₀ 227.27 μ g/ml.

In vivo studies

Jamaludin & Mohamed [37] studied the hypoglycemic effect of *P. speciosa* extracts using glucose oxidase method. In this study, healthy Sprague Dawley rats were induced to be diabetic via intravenous injection of 60mg/kg alloxan. The two groups of normal and alloxan-induced diabetic rats were given the *P. speciosa* extract in the range of 25-500mg extract/ kg body weight (BW), together with 1g glucose/kg BW of rat. The result showed that only the chloroform extracts from both the empty pods and seeds had a strong hypoglycemic activity on diabetic rats. The seed had a higher hypoglycemic activity than the pod. The reduction of blood glucose in alloxan diabetic rats given 0.4g/ kg *P. speciosa* pod and seed were 36% and 57%, respectively. The hypoglycemic action took place as early as within an hour, best after 2 hours and lasted for at least 24 hours. The study also revealed the dose-response relationship of *P. speciosa* seed on blood glucose level. Maximum reduction of blood glucose was seen with 500mg/ kg BW [77% decrement]. Optimum dosage seen was 200mg/kg BW. There was insignificant raise of blood glucose levels of normal rats fed with 0.4 g seed or pod extracts.

As an extension of the above study, Jamaludin and co-workers [38] explored further on the actual compound contributing to the hypoglycemic effect of *P. speciosa* seeds. The hypoglycemic fraction S-9.4 was identified through chromatographic separation. The fraction showed 83% reduction of blood glucose at 100mg/kg BW, compared to 111% reduction of blood glucose at 5mg glibenclamide/kg BW. The minimum effective dose was 25mg/ kg BW. The fraction was found to contain mixture of β -sitosterol (66%) and stigmasterol (34%). Interestingly, the two sterols exhibited synergistic effect. No hypoglycemic effect produced when the sterols were tested individually.

The similar author, Jamaludin and co-workers [11] identified component P-7.1 (stigmast-4-en-3-one) as the compound responsible in producing hypoglycemic activity of pod extract. Compared to 111% blood glucose reduction by 5mg/ kg BW glibenclamide, 100mg/ kg BW of P-7.1 reduced the glucose levels by 84%. The minimum effective dose was 50mg empty pods/ kg. The extract at 400mg/ kg BW did not significantly change the blood glucose levels in normally fed rats.

Antitumor/Antimutagenicity

We found five articles highlighting the antitumor activity of the plant. Several terms including antimutagenicity, antiproliferation and antiangiogenic were combined as to describe antitumor activity. All of the articles were *in vitro* studies. One of the studies by Aisha and colleagues [17] has been mentioned earlier in antioxidant activity. Three articles used *P. speciosa* seed [41-43], one article used empty pods [17] and one article used pericarp and seed coat [44]. Various techniques have been used to demonstrate the antitumor activity such as Epstein-Barr virus (EBV) inhibitory as-

say [41], Ames preincubation method against Trp-P-1 in Salmonella typhimurium TA98 [42], thioproline determination [43], Peripheral Blood Mononuclear Cell (PBMC) culture and 2,3-bis-(2-methoxy-4-nitro-5-sulfophenyl)-2H-tetrazolium-5-carboxanilide (XTT) cell proliferation test [44]; and using rat aortic rings, matrigel matrix with Human Umbilical Vein Endothelial Cells [HUVEC] and Vascular Endothelial Growth Factor (VEGF) level [17]. Effect of thermal processing to the antitumor activity was assessed in two studies [42,43]. All articles demonstrated positive antitumor activity of *P. speciosa*.

In vitro studies

Murakami and colleagues [41] studied the antitumor activities of 114 methanol extracts of edible Malaysia plants for their antitumor activity towards tumor promoter HPA-induced Epstein Barr Virus (EBV) activation in Raji cells. EBV activation was evaluated by detecting early antigen (EA) which was stained with high titre EA-positive sera from nasopharyngeal carcinoma patients. The results were ranked into 4 categories, based on inhibitory rate (IR) toward EBV activation and cell viability (CV): +++ strongly active (IR \geq 70% and /or 50% >CV); ++ moderately active (70% > IR \geq 50% and CV \geq 50%); + weakly active (50% > IR \geq 30% and CV \geq 50%) and - inactive (30% > IR and CV \geq 50%) *P. speciosa* showed weakly active inhibitory rate towards tumor promoter HPA-induced Epstein Barr Virus (EBV) activation, with 45% IR and >90% CV. Of the samples studied, 32% [37 samples] showed strongly active, 25% (28 samples) moderately active and 17% (19 samples) weakly active inhibitory action.

Research study done by Tangkanakul and co-workers [42] examined 10 foods in central and southern Thailand containing local Thai vegetables. Methanol extracts of both the raw ingredients and homogenized foods were evaluated for antioxidant activity using DPPH scavenging assay and antimutagenic assay using Ames preincubation method (against Trp-P-1 in Salmonella typhimurium TA98). Total phenolic content was determined using the Folin-Ciocalteu reagent. Raw *P. speciosa* showed antioxidant activity of 0.04 g vitamin C equivalent / 100g [VCE/100 g], total phenolic content of 0.13 g GAE/100 g and 31% inhibition against Trp-P-1. A strong antimutagenicity was demonstrated by galangal, lemongrass, wild betel, turmeric and kaffir lime leaf (87-97%). Both antioxidant activity and antimutagenicity increased after the plant was cooked. The antioxidant activity increased from 0.04 g VCE/100g to 0.14gVCE/100 g, and the the antimutagenic activity increased from 31%-69%. This result showed benefit of cooking to enhance both bioactivities.

Thioproline or thiazolidine-4-carboxylic acid (TCA) is a sulphur-containing amino acid, produced from condensation of formaldehyde and cysteine. It was documented as an effective nitrite-trapping agent in human body, thus inhibiting the endogenous formation of carcinogenic N-nitroso compounds [45]. In this experiment, Suvachittanont and colleague [43] evaluated the formaldehyde, thiol and TCA level in various edible leguminous seeds in

Thailand. The uncooked *P. speciosa* was found to have the highest level of formaldehyde [0.77 ± 0.07 mmol/100 g], which decreased upon boiling [0.25 ± 0.18 mmol/100 g]. This observation could be due to volatilization of formaldehyde and formation of TCA upon boiling. Consistently, the TCA content of uncooked *P. speciosa* was <0.001 mmol/100 g dry beans and it increased to 0.14 ± 0.02 mmol/100 g dry beans after boiling. The highest level of TCA was found in Archidendron clypearia ['Niang Nok'] both in uncooked and cooked status.

In subsequent study, Tunsaringkarn and colleague [44] evaluated the antiproliferation activity of human white blood cells, Peripheral Blood Mononuclear Cell (PBMC) with 21 Thai Mimosaceae plant extracts. *P. speciosa* pericarp and seed coat were among plants with high inhibitory cell proliferation (17.17% and 12.16%, respectively). The researchers correlated the high inhibitory effect with tannin level, which was also consistently high, 250mg/g in *Parkia speciosa* pericarp and 350mg/g in *Parkia speciosa* seed coat. This finding suggested tannin as potential cancer therapeutic agent.

In the assessments of antiangiogenic activity, the methanolic extract and all its sub-extracts showed more than 50% inhibition of rat aortic microvessel outgrowth [17]. *P. speciosa* extracts also inhibited tube formation on matrigel matrix involving HUVECs (Human Umbilical Vein Endothelial cells). Under light microscopy, the HUVECs treated with *P. speciosa* extracts showed formation of cytoplasmic vacuoles, which are markers of autophagy as a result of nutritional deprivation which is essential to maintain cell viability [46]. The vascular endothelial growth factor (VEGF) concentration of treated HUVECs was also reduced, (36 ± 2.2 pg/ml) compared to 51 ± 1.6 pg/ml in untreated cells. The extracts did not show acute toxicity.

Antimicrobial activity

We found six articles on this bioactivity, and all were *in vitro* studies. Three studies have also experimented on the antioxidant studies described earlier [5,6,18]. Main methods used in the experiments were agar-well diffusion assay and disc diffusion method.

In vitro studies

Uyub and co-workers [47] reported that the extracts of *P. speciosa* seeds in petroleum ether, chloroform, and methanol demonstrated antibacterial activity against Helicobacter pylori but none was found in the water extract. The activity was found highest in the chloroform extract followed by methanol and petroleum ether. In comparison, the chloroform extract of *P. speciosa* showed a moderate inhibition zone diameter to mg extract ratio (25.0), compared to other plant extracts where the ratios were in the range of 1.5-117.5.

In the study of Sakunpak and Panichayupakaranant [48], amongst the forty-four extracts of twenty-two Thai edible plants that were investigated for antibacterial activity using the disc diffusion method, the methanolic *P. speciosa* seed extract was found able to inhibit Helicobacter pylori growth, while the ethyl acetate

extract was effective against *Escherichia coli*. These extracts, however, had no inhibitory effect on *Salmonella typhimurium*, *Salmonella typhi*, and *Shigella sonnei* growth. According to Gmelin and co-worker [49], two cyclic polysulfide compounds, hexathionine and trithiolane in the *P. speciosa* seeds are contributing to its antibacterial property.

In Thailand, the 'Sataw' or *P. speciosa* pods extract could be potentially used as a natural preservative [18]. Both 'Sataw-Khao' and 'Sataw-Dan' pod extracts under investigated by the same group of researchers showed antimicrobial activity against food-borne pathogenic bacteria (*Bacillus cereus*, *Listeria monocytogenes*, *Staphylococcus aureus*, *Escherichia coli*, *Salmonella typhimurium*, *Vibrio cholerae* non O1/ non O139) and food spoilage bacteria (*Aeromonas hydrophila*, *Pseudomonas aeruginosa*, *Serratia marcescens*). Although there were no significant differences between 'Sataw-Khao' and 'Sataw-Dan' pod extracts against all the tested bacteria, but for Gram negative bacteria, the extracts exhibited a lower range of inhibition zone than Gram positive bacteria. However, the result is not in favour for *Vibrio cholerae* which was the most susceptible strain in comparison with other tested Gram-negative bacteria. These results are in agreement with the findings of Musa and co-workers [50] who observed that *P. speciosa* extract could be effective against all Gram-positive bacteria [*Streptococcus agalataiae*, *S. aeruginosa* and *S. aureus*] and some Gram-negative bacteria (*A. hydrophila* and *V. parahaemolyticus*). However, the Gram-negative bacteria including *Citrobacter freundii*, *Edwardsiella tarda*, *E. coli* and *V. alginolyticus* were resisted to the extract.

Similar findings were reported by Sonia and co-workers [6], whereby the methanolic extract of *P. speciosa* pod showed the inhibition against common pathogens [*Staphylococcus aureus*, *Escherichia coli*, *Pseudomonas aeruginosa* and *Klebsiella pneumoniae*]. The highest zone of inhibition was demonstrated for the gram-positive *Staphylococcus aureus* at 10 mm. The bioactive compounds are responsible to effectively inhibit and/or stop microbial growth via disruption of the synthesis of microbial nucleic acids, proteins and cell walls [51]. According to the previous studies, *P. speciosa* pod extract could inhibit all tested pathogenic and spoilage bacteria compared to rambutan peel, mangosteen peel, palmyra peel and coconut husk [52], rambutan peel and seed [53], pomelo peel [54] and banana peel [55] which could only inhibit certain bacteria and their inhibition zones were reported to be inferior to the *P. speciosa* extract.

Fatimah [56] studied the utilization of *P. speciosa* pod extract as reducing agent in silver nanoparticles (Ag NPs) synthesis and its potential as antimicrobial agent. She reported that the microwave-assisted synthesis of Ag NPs using *P. speciosa* pod extract demonstrated antibacterial activity against *Escherichia coli*, *Staphylococcus aureus*, and *Pseudomonas aeruginosa*. The specific compounds that act as reducing agents and support the antimicrobial activity of Ag NPs are generally flavonoids and polyphenol compounds.

From the comparison study done by Ghasemzadeh and co-workers [5], *P. speciosa* extract from Perak had a strong inhibitory effect towards both Gram positive and Gram-negative bacteria. The bacterial strains that were most susceptible to *P. speciosa* extract were *Staphylococcus aureus* [7.2±0.346 mm inhibition zone, minimal inhibitory concentration of 40.0 µg/ml] and *Bacillus subtilis* [8.4±0.320mm inhibition zone, minimal inhibitory concentration of 40.0 µg/ml]. The Gram-negative bacteria were less sensitive to the extracts compared to Gram positive, due to the presence of a cell wall that prevents permeation of the extract into the cell. The antibacterial activity was correlated to the effect of gallic acid.

Effects on Cardiovascular system

Four articles were reviewed for the effects of *P. speciosa* on cardiovascular system. Three research studies done *in vitro* involving human umbilical vein endothelial cells [HUVECs], cardiomyocytes (cells of the heart) and Angiotensin-converting enzyme (ACE) [19,57,58]. The other study was done *in vivo* [59]. One study also experimented antioxidant activity [described in the antioxidant section] [19]. Three of the articles used empty pod extracts [57-59] and one article used seed [19]. The chemical agents used apart from *P. speciosa* extract were tumor necrosis factor- α (TNF- α), quercetin, nicardipine, N(G)-nitro-L-arginine methyl ester (L-NAME) and ACE solution. Among the outcomes observed are the inflammatory protein expressions: nuclear factor kappa B cell (NF κ B) p65, p38 mitogen-activated protein kinase [p38 MAPK], inducible nitric oxide synthase (iNOS), cyclooxygenase-2 (COX-2) and vascular cell adhesion molecule-1 (VCAM-1) [57,58]; blood pressure level, plasma nitric oxide level, cardiac angiotensin converting enzyme (ACE) inhibitory activity, NADPH oxidase activity and cardiac lipid peroxidation content [59]. The outcome proved cardioprotective effect of the *P. speciosa* extract.

In vitro studies

As a basis of understanding, TNF- α is a proinflammatory cytokine that has been used in many *in vitro* studies to induce inflammation [57]. It stimulates inflammatory markers, such as iNOS, nitric oxide [NO], COX-2 and VCAM-1, through [NF κ B] pathway activation [60]. NF κ B induces cardiomyocyte hypertrophy [61]. In addition, p38 MAPK was also described to be involved in the inflammatory process by triggering the synthesis of inflammatory regulators such as TNF- α and COX-2 [62]. Elevated p38 MAPK activity resulted in augmented inflammatory, hypertrophic and fibrotic processes in patients with end-stage heart failure and ischemic heart disease [63].

Mustafa and colleague [57] studied the anti-inflammatory activity of *P. speciosa* empty pod extract in human umbilical vein endothelial cells [HUVECs]. HUVECs were divided into four groups: HUVECs exposed to TNF- α [10 ng/ml] in the presence [25µg/ml] or absence of *P. speciosa* extract. Quercetin act as positive control while HUVECs without TNF- α served as negative control. The concentration of the *P. speciosa* extract was chosen at 25µg/ml because

of its highest cell viability in HUVECs co-incubated with TNF- α . Quercetin was used as the positive control as it was present in the extract. All the inflammatory protein expressions were reduced in HUVECs with *P. speciosa* extract, namely NF κ B p65, iNOS, COX-2 and VCAM-1, as determined with Western blot analysis. The nitric oxide [NO] and reactive oxygen species [ROS] level were also decreased [10.24 μ M and 120%, respectively]. These effects were comparable to that of quercetin. This observation was demonstrated as *P. speciosa* extract attenuates TNF- α -induced inflammatory responses by blocking the activation of NF κ B p65 and thus reduces the iNOS, COX-2 and VCAM-1 expressions as well as ROS and NO production.

A relatively similar study design was performed by Gui and colleague [58], in which inflammation-induced cardiomyocytes were used to determine the anti-inflammatory property of *P. speciosa* extract. The researchers proposed that the anti-inflammatory effect of *P. speciosa* were due to modulation of NF κ B and MAPK pathways. The cardiomyocytes were divided into four groups: negative control, cardiomyocytes exposed to TNF- α , cardiomyocytes exposed to *P. speciosa* extract and TNF- α and cardiomyocytes exposed to quercetin and TNF- α . The *P. speciosa* extract [500 μ g/ml] and quercetin [1000 μ M] used were different in quantity compared to the previous study. The NF κ B p65 and p38 MAPK expression were reduced in cardiomyocytes pre-treated with *P. speciosa* extract or quercetin. Similarly, the iNOS, COX-2 and VCAM-1 expression as well as NO and ROS levels were also reduced. This effect confirmed the postulation and could be attributable to the polyphenol content of *P. speciosa*, specifically quercetin.

Using alcalase, Siow and Gan [19] examined the antihypertensive bioactive peptides from *P. speciosa* seeds. Amino acids such as isoleucine, valine, phenylalanine and tyrosine contributed to the angiotensin converting enzyme [ACE]-inhibitory activity. A comparison of the ACE-inhibitory activity was made between different temperature, substrate-to-enzyme [S/E] ratio and incubation time. The highest percentage of ACE-inhibitory activities [80.2%] were demonstrated in a condition of 50 $^{\circ}$ C, S/E ratio of 50 and 2 hours incubation time.

In vivo studies

Kamisah and colleague [59] experimented the effects of *P. speciosa* empty pod extract in rats given N(G)-nitro-L-arginine methyl ester (L-NAME). L-NAME is an inhibitor of nitric oxide synthase. It reduces plasma nitric oxide and increases systolic blood pressure [due to relaxation of blood vessel], ACE and NADPH oxidase activities, as well as lipid peroxidation in the heart [59]. Twenty-four

male Sprague Dawley rats were divided into 4 groups: Group 1 was given L-NAME (25mg/kg, intraperitoneally), Group 2 was given L-NAME and *P. speciosa* empty pods methanolic extract (800mg/kg, orally), Group 3 was given L-NAME and nicardipine (3mg/kg, orally) and Group 4 served as negative control. The plasma nitric oxide reduction was inhibited in Group treated with *P. speciosa* (12.33%), but not with nicardipine (-29.29%). The blood pressure increase was prevented in groups given *P. speciosa* and nicardipine. However, no difference was seen between the two groups. Consistently, the cardiac ACE, NADPH oxidase activity, as well as cardiac lipid peroxidation content were reduced in groups given *P. speciosa* and nicardipine. These effects were the result of high polyphenol content in *P. speciosa*. As being reported in previous studies, quercetin exerted hypertensive effect by enhancing nitric oxide production via induction of endothelial nitric oxide synthase [eNOS] phosphorylation [64]. In addition, quercetin also was shown to reduce ACE protein level in endothelial cells [65] and inhibit myocardial NADPH oxidase-dependent superoxide anion generation in hypertensive rats [64].

Conclusion

P. speciosa or known as stinky bean, is a plant found abundantly in Southeast Asia. Despite of its odorous smell, it has been used both as culinary ingredients and indirectly as a medicine to treat various diseases such as hypertension and urinary tract infection. Various parts of the plant were used, including seed, pod and seed coat. The health benefits of *P. speciosa* are mainly contributed by the phenolic and flavonoid content. The antioxidant property is due to the presence of phenols, flavonoids, hydrogen sulphide and tannin, although the levels differ based on the types of extracts, parts of the plant, plantation and post-harvest handling. *P. speciosa* also has hypoglycemic effect by inhibiting α -glucosidase, α -amylase and pancreatic lipase. The effect was due to the synergistic effect of β -sitosterol and stigmasterol, and by a compound known as stigmast-4-en-3-one. The antitumor activity was exhibited more in pericarp, seed coat and empty pods, compared to the seed of the plant. *P. speciosa* demonstrated antimicrobial activity against both Gram positive and negative pathogen, although the effect was less potent in the latter. *P. speciosa* inhibit inflammatory markers by blocking the NF κ B and MAPK pathways, prevent plasma nitric oxide loss, as well as inhibiting heart angiotensin-converting enzyme. We found that limited animal study and absent of clinical trial was done in evaluating the medicinal effects of *Parkia speciosa*. Thus, future animal and human studies are needed to strengthen evidence of its medicinal effect.

Table 1: List of Abbreviations.

DPPH	1,1-DIPHENYL-2-PICRYLHYDRAZYL FREE RADICAL
FRAP	REDUCING FERRIC ION ANTIOXIDANT POTENTIAL
ABTS	2,2'-AZINO-BIS [3-ETHYLBENTHIAZOLINE-6-SULFONIC ACID]
H2S	HYDROGEN SULPHIDE
GAE/G	GALLIC ACID EQUIVALENTS PER GRAM

RE/G	RUTIN EQUIVALENTS PER GRAM
IC50	HALF-MAXIMAL INHIBITORY CONCENTRATION
BHT	BUTYLATEDHYDROXY TOLUENE
TE/G	TROLOX EQUIVALENTS PER GRAM
EDTAE/G	ETHYLENEDIAMINETETRAACETIC ACID EQUIVALENT/G DRY WEIGHT
FESO4	FERROUS SULFATE
H ₂ O ₂	HYDROGEN PEROXIDE
FDPSP	FREEZE DRIED P. SPECIOSA POD
TPC	TOTAL PHENOLIC CONTENT
TFC	TOTAL FLAVONOID CONTENT
%DPPHSC	PERCENTAGE DPPH WHICH WAS SCAVENGED
%ABTSSC	PERCENTAGE ABTS WHICH WAS SCAVENGED
EC50	HALF MAXIMAL EFFECTIVE CONCENTRATION
DADS	DIALLYL DISULPHIDE
DATS	DIALLYL TRISULPHIDE
BCU	
MCF- 7	MICHIGAN CANCER FOUNDATION-7
DATS- E	DATS EQUIVALENT
CCA	CUPRIC ION CHELATING CAPABILITIES
TEACDPPH	TROLOX EQUIVALENT ANTIOXIDANT CAPACITY DPPH
TEACFRAP	TROLOX EQUIVALENT ANTIOXIDANT CAPACITY FRAP
CMC	CARBOXYMETHYLCELLULOSE
MDA	MALONDIALDEHYDE
GSH	TOTAL GLUTATHIONE
SOD	SUPEROXIDE DISMUTASE
PPL	PORCINE PANCREATIC LIPASE
BW	BODY WEIGHT
EBV	EPSTEIN-BARR VIRUS
PBMC	PERIPHERAL BLOOD MONONUCLEAR CELL
XTT	2,3-BIS-(2-METHOXY-4-NITRO-5-SULFOPHENYL)-2H-TETRAZOLIUM-5-CARBOXANILIDE
HUVEC	HUMAN UMBILICAL VEIN ENDOTHELIAL CELLS
VEGF	VASCULAR ENDOTHELIAL GROWTH FACTOR
EA	EARLY ANTIGEN EA
IR	INHIBITORY RATE
CV	CELL VIABILITY
VCE/ 100 G	VITAMIN C EQUIVALENT / 100 G
TCA	THIOPROLINE OR THIAZOLIDINE-4-CARBOXYLIC ACID
AG NPS	SILVER NANOPARTICLES
ACE	ANGIOTENSIN-CONVERTING ENZYME
TNF-A	TUMOR NECROSIS FACTOR-A
L-NAME	N(G)-NITRO-L-ARGININE METHYL ESTER
NFKB) P65	NUCLEAR FACTOR KAPPA B CELL
P38 MAPK	P38 MITOGEN-ACTIVATED PROTEIN KINASE
INOS	INDUCIBLE NITRIC OXIDE SYNTHASE
COX-2	CYCLOOXYGENASE-2
VCAM-1	VASCULAR CELL ADHESION MOLECULE-1
NADPH	NICOTINAMIDE ADENINE DINUCLEOTIDE PHOSPHATE HYDROGEN
ENOS	ENDOTHELIAL NITRIC OXIDE SYNTHASE

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