



Review Article

Copy Right@ Rui Han

Cytokines Involved in the Progression of Liver Fibrosis by Influencing DNA Synthesis

Shixin Li², Jiayin Li³, Hongxing Yang³, Shutang Wang³, Zhuangzhong Chen³, Jinghui Wang³, Sisi Wang³, Lingeng Lu¹ and Rui Han^{1,3*}

¹Department of Chronic Disease Epidemiology, Yale University, USA

²Department of Oncology, Guang anmen Hospital of China Academy of Chinese Medical Sciences, China

³Department of Oncology, The First Affiliated Hospital of Guangzhou University of Chinese Medicine, China

*Corresponding author: Rui Han, Department of Chronic Disease Epidemiology, Yale School of Public Health, School of Medicine, Yale University, New Haven, CT 06520-8034, USA.

To Cite This Article: Shixin Li, Jiayin Li, Hongxing Yang, Shutang Wang, Rui Han, et al., *Cytokines Involved in the Progression of Liver Fibrosis by Influencing DNA Synthesis*. *Am J Biomed Sci & Res.* 2021 - 13(1). *AJBSR.MS.ID.001825*. DOI: [10.34297/AJBSR.2021.13.001825](https://doi.org/10.34297/AJBSR.2021.13.001825).

Received: 📅 December 04, 2020; **Published:** 📅 May 27, 2021

Abstract

Liver fibrosis is a pathological change in which the connective tissue in the liver proliferates abnormally, resulting in abnormal liver function or structure. Long-term liver fibrosis can develop cirrhosis or liver cancer, which poses a great threat to human health. This article reviews the cytokines closely related to liver fibrosis in recent years. These cytokines affect the development of liver fibrosis mainly through processes that affect DNA synthesis. Platelet-derived growth factor promotes the division and proliferation of hepatic stellate cells by stimulating cellular DNA synthesis. Insulin-like growth factor, on the other hand, shows the proliferation of myofibroblastic cells in a manner that promotes the cell cycle to enter the DNA synthesis phase. Interleukin-6 promotes the progression of DNA synthesis in a manner that activates cyclins. We aim to find new approaches for the treatment of liver fibrosis by means of regulating DNA synthesis as the main approach by summarizing existing studies.

Keywords: Liver fibrosis; Cytokines; DNA synthesis

Introduction

Liver fibrosis is a pathological change in which the connective tissue in the liver is abnormally proliferated by chemical or viral infection, causes the abnormality of liver structure or function. It is characterized by massive deposition of fibrous connective tissue (such as collagen), ECM compounds and secretion of chemokines and fibrotic factors caused by acute or chronic injury. Liver fibrosis is a typical scar response to almost all forms of chronic liver injury [1]. Liver fibrosis has historically been considered a passive and irreversible process due to rupture of the liver parenchyma and replacement by collagen-rich tissue [2,3]. Current studies have shown that liver fibrosis could be reversible in the early stages. However, persistent liver damage and scar formation can still lead to advanced liver cirrhosis and even hepatocellular carcinoma (HCC) [4,5]. The characteristic of liver disease is the gradual progression from chronic hepatitis to liver fibrosis. Persistent liver fibrosis can lead to impaired liver function and destruction of liver

tissue structure, which can lead to liver cirrhosis or hepatocellular carcinoma. Liver disease is a disease with a high mortality rate as the disease advances [6]. Chronic liver disease and cirrhosis are the 11th leading cause of death in the United States [7], with more than one million death each year worldwide [8].

The process of DNA synthesis is embodied in the process of cell proliferation. The cell cycle is the basic process of cell proliferation. It refers to the whole process that a cell goes through from the completion of one division to the end of the next division. It has been divided into two stages: interphase and division. Furthermore, the interval is divided into three phases which are early DNA synthesis period (G1 period), the DNA synthesis period (S period) and the late DNA synthesis period (G2 period). The division period is the DNA division period (M period) [9,10]. The G1 phase of the cell cycle prepares for DNA replication, the S phase replicates chromosomes, the G2 phase prepares for mitosis before chromosome separation,



and the final M phase undergoes mitosis. G0/G1 is an important checkpoint, which is the restriction point occurs in the middle of G1, after this R point, the cell is committed to cell division. All those processes could be related to liver fibrosis. Here we summarized Cytokines involved in the progression of liver fibrosis which might help us to understand the mechanisms and look for the potential therapeutic targets or approaches for liver fibrosis.

Pathogenesis of Liver Fibrosis

Liver fibrosis is the result of the liver's wound healing response after repeated damage to liver cells [11]. After acute liver injury, parenchymal cells regenerate and replace necrotic or apoptotic cells. If liver injury keeps existing, liver regeneration might eventually fail, liver cells were then replaced by abundant ECM and fibrous collagen, and normal cells such as metabolic activity could be disappeared, resulting in liver fibrosis [1].

The damaged liver cells release ROS and fibrosis mediators and induce inflammatory cells to recruit white blood cells and activate Kupffer cells. Studies have also suggested that the activation of hepatic stellate cells (HSC) plays an important role in the formation of liver fibrosis [12]; in addition to HSC, portal myofibroblasts and bone marrow-derived cells also have the potential for fibrogenesis [13]. A variety of cells such as lymphocytes, polymorphonuclear cells and other inflammatory cells, damaged hepatocytes, bile duct cells and Kupffer cells [14] can secrete inflammatory cytokines to activate HSC; activated HSC could also secrete inflammation which maintains its activated state Sex cytokines. Therefore, vicious cycle of mutual stimulation of inflammatory cells and HSC could keep existing, which can also aggravate the occurrence of liver fibrosis, if the factors that cause liver cell damage remain existed [15]. This inflammatory environment will stimulate the transdifferentiation of native HSC into activated myofibroblasts (MFB), and the apoptosis of damaged liver cells can stimulate MFB to undergo fibrosis [16]. Then promoting HSC apoptosis can reduce the production of MFB cells, thereby interfering with the process of liver fibrosis [17]. Hepatocyte apoptosis directly triggers HSC activation through the phagocytosis of apoptotic bodies, or indirectly triggers HSC activation through injury-related molecular patterns that induce HSC migration and activation [18]. If liver damage persists, the accumulation of activated HSC and portal fibroblast MFB could be activated, a large amount of extracellular matrix protein (ECM) could also be synthesized [19]. Moreover, ECM protein has been proven to directly stimulate liver fibrosis. Studies have shown that, for delaying or reversing the progression of liver fibrosis, the cause of liver damage should be eliminated, or ECM degradation should be achieved through intervention (ECM degradation is inhibited by cytokines such as TIMPs, TGF- β).

Cytokines that Influence the Inflammatory Caused by Liver Injury

Damaged hepatocytes and bile duct cells can release inflammatory cytokines and soluble factors, which activate kupffer cells and stimulate the recruitment of activated T cells. This inflammatory environment stimulates the activation of resident HSCs into fibroblasts. HSC is the main fibroblast cell that is considered to play a role in liver fibrosis [20]. The HSC activation process is driven by cytokines released by damaged liver cells, bile duct cells, and T cells such as transforming growth factor (TGF- β) [21], platelet-derived growth factor (PDGF) [22], Interleukin-6 (IL-6) [23]. In the process of liver fibrosis, the transformation of static HSCs into activated fibroblasts is the central step of development. Therefore, inhibition of activated HSC is considered an important strategy for anti-fibrosis therapy [24].

TGF- β

TGF- β is a typical inflammatory cytokine secreted by a variety of hepatocytes including human hepatic stellate cells HSC, MFB [25], kupffer cells [26] and hepatocytes [27] with a typical profibrotic effect. It has obvious effects in HSCs activation The profibrotic effect [28] is the main fibrotic cytokine that activates HSCs [29,30]. This effect may be related to the stimulation of TGF- β that also activates other growth factors involved in HSC activation, including PDGF, CTGF, VEGFA and Fibroblast growth factor-2; and the release of TGF- β from necrotic hepatocytes is likely to be one of the primary signals for the activation of adjacent HSCs and subsequent transdifferentiation into MFB [31]. Therefore, TGF- β plays a key role in the HSC activation process, and epigenetic modification may be involved in this process [32]. Some studies have explored the epigenetic regulation of TGF- β signaling in other biological environments, and have shown that DNA methylation [33,34], MicroRNA (miRNA) and histone modifications regulate the gene expression of key effector factors of this pathway [35].

TGF- β activates HSC through the classical Smad signaling pathway and JNK signaling pathway. In the TGF- β /SMAD signaling pathway, TGF- β binds to the type I TGF- β receptor to initiate the pathway, activates and phosphorylates the downstream mediators SMAD2 and SMAD3, then forms a complex with SMAD4 and translocates to the nucleus, through binding to DNA or combined with DNA binding protein to regulate gene transcription [30,36], this process is negatively regulated by inhibitory SMAD7 [37]. SMAD7 is an effective inhibitor of TGF- β signaling. The overexpression of SMAD7 not only inhibits the activation of SMAD2/3, but also blocks NF- κ B signaling during fibrosis and inflammation. In terms of liver fibrosis, SMAD3 is a cause sick, and SMAD7 is protective [38]. It

has been proposed that SMAD7 is a potential therapeutic target for liver fibrosis.

PDGF

PDGF is produced by many types of cells. When hepatocytes are damaged, it is mainly secreted by non-parenchymal Kupffer cells and sinusoidal endothelial cells [39]. It is one of the most effective mitogens for activating HSC. PDGF has significant mitogenic and motor-promoting effects [40], which can promote cell division and control maturity. PDGF can activate HSC through autocrine and paracrine signals [41] and plays an important role in stimulating the proliferation of HSC and changing its cytoskeleton distribution. PDGF can also promote the differentiation of HSC into myofibroblasts (MFBs) and regulate the process of liver fibrosis [42]. Studies [43] have proved that PDGF promotes the secretion and release of IGF and its binding protein in rat HSC and promotes the division and proliferation of HSC by stimulating cell DNA synthesis.

IGF

Insulin-like growth factor (IGF) is a polypeptide mitogen similar to proinsulin. Therefore, it plays an important role in regulating cell proliferation, differentiation and apoptosis, and mediates cell growth, immune regulation and anabolism [44]. After acute liver injury, hepatocytes secrete IGF-I in the pre-inflammatory stage. During the inflammation process of liver fibrosis, kupffer cells and activated HSC secrete IGF-I [45]. There are two main pathways for IGF-I intracellular signal transduction, the PI3-K pathway and the extracellular signal-regulated kinase (ERK) pathway [46]. These pathways can mediate the interaction between IGF-I mitogenic response and anti-apoptotic response. In many cell types, IGF-I regulates proliferation by affecting the cell cycle [47]. In skeletal muscle, IGF-I enhances the G1/S phase process in a PI3 kinase-dependent manner [48] and can play a mitogenic effect. Studies [44] indicate that for HSCs in either the stationary or active phases, the effect of IGF-I on DNA synthesis is through the ERK1/2 pathway.

Cytokines Involved in the Hepatic Fibrogenesis

Chronic liver injury of various etiologies can lead to liver cell apoptosis, the repair mechanism is triggered, and an inflammatory response occurs [49]. Inflammatory cells such as lymphocytes or polymorphonuclear cells [50] can activate HSCs by releasing inflammatory cytokines. Therefore, makes those static, non-proliferative cells enriched with vitamin A transform into an activated contractile myofibroblast phenotype which is characterized by increased activation of DNA synthesis, proliferation, α -smooth muscle Actin (α -SMA) expression and synthesis of various ECM components [51]. Then, the production of ECM protein could be triggered. Accumulation of ECM protein could thereby replace the liver parenchymal area and lead to fibers into a state of liver cirrhosis [52]. MFB is the main source of ECM

protein accumulation and the main mediator of fibrosis. Increased expression of fibrotic ECM plays an important role in liver fibrosis [53-55].

TGF- β

TGF- β is a powerful fibrotic factor responsible for the synthesis of extracellular matrix [56], which forms liver fibrosis by inducing MFB to express α -SMA. TGF- β 1 activates intracellular mediators through TGF- β type I and type II receptors, such as Smad protein, p38 mitogen-activated protein kinase (MAPK) and extracellular signal-regulated kinase pathway [57]. Phosphatidylinositol 3-kinase (PI3K) and MAPK signaling pathways must both be activated to induce mitogen-mediated DNA synthesis, while Janus kinase (JAK) triggers DNA synthesis by activating PI3K and MAPK [58].

TGF- β 1 stimulates the secretion of activated HSC to induce the up-regulation of the two indicators of fibrosis, COL1A2 and α -SMA, and leads to excessive deposition of ECM protein. Studies have shown that the binding sites of transcription factors Sp1, AP-1 and Smad in the promoter region of Col1A2 are related to the transcription of fibroblasts induced by TGF- β 1 [59]. The site is related to the transcription of fibroblasts induced by TGF- β 1. Subsequent studies have shown that both Physalin D and Maleic acid derivative anti-oxidant can inhibit liver fibrosis by reducing the activity of the Col1A2 promoter [60]. Studies have shown that calcineurin is necessary for TGF- β -mediated ECM protein deposition. The regulator of calcineurin 1 (RCAN1) is an endogenous inhibitor of CaN [61]. Studies have shown that RCAN1.4 can reduce TGF- β -mediated liver fibrosis by inhibiting CaN / NFAT3 signaling [62]. On the other hand, overexpression of RCAN1.4 may induce cell cycle arrest in G0/G1 phase of activated HSCs and reduce the expression of cell cycle-related proteins. Therefore, RCAN1.4 can be used as a therapeutic target for the treatment of liver fibrosis.

IGF

In MFB, the ERK1/2 signaling pathway is necessary to promote proliferation [63]. In the process of MFB proliferation, IGF-I also promotes DNA synthesis. Research [44] displayed that IGF-I can initiate the process of DNA synthesis phase in the cell cycle, increases the number of cells in S phase, and also exerts an anti-apoptotic effect on the bcl system which locates in the upstream of ERK1/2. Therefore, increased proliferation of MFB cells promoted by IGF-I could be observed that instead of cell apoptosis.

Cytokines Involved in Fibrosis Resolution

Once the liver had been injured, a regeneration process could be initiated. During this process, the quiescent liver cells exit the quiescent phase (G0) and re-enter the cell cycle to proliferate and compensate for the lost liver tissue [57]. A variety of cytokines including hepatocyte growth factor (EGF), epidermal growth factor

(EGF), tumor necrosis factor- α (TNF- α), IL-6, insulin and TGF- β Coordinating cell cycle progression. They integrate with each other in the mitogen-sensitive G1 phase of the cell cycle until the restriction point (R), turning the cell into the S phase dedicated to proliferation, and starting the process of DNA synthesis [64]. HGF and EGF are powerful hepatocyte mitogens [65]. Although IL-6 is much smaller than the former, it can induce DNA synthesis. All three are important cytokines that promote hepatocyte proliferation [66]. Compared with IL-6, HGF and EGF, TGF- β 1 is one of the most well-known hepatocyte anti-proliferation factors which can regulate the termination of liver regeneration and keep liver cells remaining under normal conditions [67].

A large amount of ECM proteins are deposited during liver fibrosis, resulting in changes in liver tissue and distortion of vascular structure. Therefore, digestion of ECM proteins is also one of the pathways to eliminate hepatic fibrosis [68]. Protein degradation mainly regulated by two pathways, one is mediated by autophagy-lysosomal pathway (ALP), which mainly degrades long-lived proteins and insoluble protein aggregates. The other one is dependent on ATP, non-lysosomal pathways mediated by the ubiquitin-protease pathway. It participates in the regulation of biological processes such as cell cycle, cell apoptosis, DNA repair, and intracellular signal transduction.

IL-6

IL-6 is a key factor in the mitotic response in the liver. IL-6 plays a protective role by arresting the cell cycle, allowing excision of bases, and repairing oxidized DNA. Studies [69] have demonstrated that IL-6 is important for latent Signal transducers and activators of transcription 3 (STAT3) activation, expression of cell cycle-related immediate early genes (IEGs), and initiation of cell cycle progression after partial hepatectomy. STAT3 protein plays a role in the progression from G1 phase to S phase of the cell cycle, and its activation depends on the release of IL-6 [70]. Studies [71] have shown that after the IL-6/STAT3 signaling pathway is initiated, the cell cycle enters the DNA synthesis phase. It leads to extensive cell regeneration.

TGF- β

TGF- β inhibits the proliferation of hepatocytes and keeps hepatocytes quiescent in the normal liver [7]. In addition, at the stage of liver regeneration, TGF- β plays an important role in the process of reorganization of liver regenerative tissue and establishment of extracellular matrix [72]. Cyclin D1, CDK4 and P21 are factors that regulate proliferation and cell cycle [73]. Studies have shown that [10] TGF- β 1 is involved in the expression of p21 in LX-2 cells, which affects the synthesis of DNA.

IGF

Although IGF-I can induce DNA synthesis in HSC, it still has

pro-apoptotic and anti-proliferative effects. IGF can induce HSC proliferation and also can promote HSC apoptosis in a dose-dependent manner [46]. One study [44] showed that low doses of IGF-I had an anti-fibrotic effect in a model of CCl4-induced liver fibrosis. And IGF-I can promote the healing response of liver wounds by regulating the migration of HSCs through the PI3K signaling pathway [74]. The experiments of HSCs in rats showed that NFkB and ERK1/2 were involved in the regulation of IGF apoptosis [44]. IGF-I regulates apoptosis by regulating Bcl-2 upstream of ERK. The Bcl-2 gene family is a major modulator of the mitochondrial apoptotic pathway. These results indicate that IGF-I is inducing HSC apoptosis through the mitochondrial pathway [75]. From the perspective of DNA synthesis, while IGF-I promotes the initiation of cell cycle progression in HSCs, it is possible that IGF-I induces apoptotic processes earlier than affecting the cell cycle. Therefore, IGF-I showed the performance to promote HSC DNA synthesis, but hsc reduce the number of cells, increased apoptosis.

HGF

HGF is one of the first cytokines to be found to promote liver regeneration [76,77]. HGF binds and activates the tyrosine receptor kinase MET, a multifunctional receptor involved in many cellular processes such as proliferation, growth, survival, and metabolism [78]. The mitogenic activity of HGF-initiated signal requires the function of transcription factor C/EBP β [79]. Hepatocyte growth factor/scatter factor (HGF/SF) stimulates hepatocytes to initiate DNA synthesis during the cell cycle. It protects hepatocytes from apoptosis, promotes liver regeneration in vivo, and reduces fibrosis [80]. However inhibition of PI3K or MAPK pathways can prevent HGF-induced DNA synthesis during hepatocyte proliferation [62].

Discussion

Cirrhosis caused by liver fibrosis is the leading cause of death from chronic liver disease, and chronic liver disease and cirrhosis are the 12th leading causes of death in the United States in 2016, rising to the 11th place in 2017 [7]. Liver cancer is the sixth most common cancer worldwide and the fourth leading cause of cancer death worldwide. In 2018, there were 841,080 new cases of liver cancer and 781,631 deaths worldwide, including 392,868 new cases of liver cancer and 368,960 deaths in China [81]. More importantly, more than 80% of HCCs develop from liver fibrosis [82].

HSCs and kupffer cells can activate each other to form a persistent inflammatory environment with massive monocyte infiltration, which includes lymphocytes, macrophages, plasma cells and eosinophils [83]. Lymphocytes are activated upon contact with antigens, which produce lymphokines that activate macrophages. Activated macrophages produce cytokines and chemokines that can activate lymphocytes [84]. This inflammatory environment stimulates the activation of resident HSCs into myofibroblasts. TGF- β and PDGF are the main cytokines involved

in the process. The TGF- β /Smad signaling pathway binds to DNA or DNA-binding proteins mainly by producing protein complexes. This pathway is involved in the regulation and expression of genes to activate HSCs [36]. Elevated expression of Smad7 protein was found in this pathway as a form of therapeutic fibrosis. In addition, PDGF released from platelets is a serum mitogen that plays a role in various processes of cell biology, enhancing cell division and controlling maturation [38]. PDGF activates HSCs through autocrine and paracrine signaling and stimulates cellular DNA synthesis in a dose-dependent manner to promote HSC division and proliferation, which can alter their cytoskeleton and thus promote HSC transdifferentiation into MFBs [41]. Moreover, studies have shown that amygdalin can reduce the transcription of PDGF and IGF mRNA and the expression of PDGF protein. Amygdalin reduces the development of liver fibrosis by reducing the synthesis and release of PDGF and IGF, thus attenuating their effects on HSCs [38]. The anti-apoptotic effect of IGF-I was postulated *in vitro* by Issa [85]. However, IGF-I was found to induce apoptosis in quiescent and activated HSCs in another experiment of HSC in rats, proposing that although IGF initiates the progression of HSC cell cycle, it may simultaneously arrest HSCs in G2 phase, therefore induce HSC apoptosis [44].

Cytokines secreted by damaged hepatocytes, inflammatory cells, and kupffer cells transdifferentiates hscs into MFB-expressing α -SMA, thereby acquiring fibrogenic properties that produce ECM proteins. Suffering a continuous inflammatory environment of liver injury, HSCs will continuously express MFB. MFB will then continuously proliferate to produce ECM, which ultimately leads to massive deposition of ECM and thus irreversible fibrosis. Therefore, in this stage, inhibiting MFB proliferation is an idea to eliminate fibrosis. Experimental studies showed IGF-I has a great effect on the cell cycle and proliferation as well as apoptotic signaling of MFB in rats. IGF signals intracellularly regulated by two pathways, PI3-K and ERK, and during MFB proliferation, IGF-I initiates the progression of DNA synthesis cell cycle and increases S-phase cells, but also exerts anti-apoptotic effects by acting on the bcl system upstream of ERK1/2 [44]. Therefore, inhibiting IGF synthesis or EPK signaling may be the approaches for the treatment of fibrosis.

Fibrosis can be digested by three aspects before ECM proteins deposited to form irreversible fibrosis, mainly including HSC apoptosis, ECM protein degradation, and regeneration of hepatocytes. In this paper, we mainly introduce three cytokines, IL-6, HGF, and EGF, which induce DNA synthesis and make hepatocytes proliferate by promoting the rapid entry of the cell cycle into the S phase. IGF induces HSC apoptosis by regulating the Bcl system upstream of the ERK signaling pathway [41]. As the important cytokine in the process of liver fibrosis [30], TGF- β is not only involved in the process of activating HSCs to promote the synthesis

of matrix proteins by cells, but also involved in preventing ECM degradation. During liver fibrosis, TGF- β may affect cell proliferation, exhibiting its cell cycle arrest in G1 phase or frequently responding by apoptotic responses [72].

In the process of hepatocarcinogenesis, TGF- β plays a dual role. In early cancer, TGF- β plays a role in inhibiting tumor activity and inducing cell cycle arrest and apoptosis. However, in advanced cancers, once cells acquire resistance to their inhibitory effects, TGF- β action is switched to pro-carcinogenesis, conferring cell survival, inducing cell migration and invasion, and microenvironmental alterations [51,86].

Therefore, preventing, delaying or revising liver fibrosis have absolute positive effect on liver cancer or liver cirrhosis prevention and also treatment. That makes explaining the basic mechanisms involved in liver fibrosis extremely important. By illuminating the effects of liver fibrosis related cytokines, the potential therapeutic targets could be located, then those bioactive protein could be intervened specifically for treatment goal. Beside treatment, effective prevention approach for liver cirrhosis and primary liver cancer are also deeply needed. As the basic pathological process in early stage before liver cancer or liver cirrhosis, fibrosis is the main disease that needs to be intervened for preventing liver cell from denaturation or cancerization. Effective prevention methods could save patients from subsequent and harmful treatment, such as liver transplantation, radiotherapy, chemotherapy and target therapy, etc. that is also the reason we summarized those liver cytokines here. Certainly, further related *in vivo*, *in vitro* and clinical studies are indispensable.

Conclusion

The process of liver fibrosis involves a variety of cells and cytokines and affects its development through multiple signaling pathways. We summarized the cytokines that affect liver fibrosis by regulating DNA synthesis. Among them, the literature on the regulation of liver fibrosis by TGF- β through various signaling pathways is well-established, but there are few studies on affecting the process of DNA synthesis. In the future, more extensive and comprehensive experiments can be done on the effect of TGF- β on the process of DNA synthesis, its new target for the treatment of liver fibrosis was discovered.

Authors' Contributions

Shixin Li and Jiayin Li contributed equally to this work as co-first authors.

References

1. Bataller R, Brenner DA (2005) Liver fibrosis. *J Clin Invest* 115(2): 209-218.
2. Popper H, Uenfriend S (1970) Hepatic fibrosis. Correlation of biochemical and morphologic investigations. *Am J Med* 49: 707-721.

3. Suriawinata AA, Thung SN (2006) Acute and chronic hepatitis. *Semin Diagn Pathol* 23(3-4): 132-148.
4. Soyer MT, Ceballos R, Aldrete JS (1976) Reversibility of severe hepatic damage caused by jejunoileal bypass after re-establishment of normal intestinal continuity. *Surgery* 79(5): 601-604.
5. Mokdad AA, Lopez AD, Shahrzaz S, Lozano R, Mokdad AH, et al. (2014) Liver cirrhosis mortality in 187 countries between 1980 and 2010: a systematic analysis. *BMC Med* 12: 145.
6. Matsumoto K, Wu Y, Fujita S, Seto K, Hatakeyama Y, et al. (2021) Cost of illness of liver diseases in Japan. *Ann Hepatol*.
7. Kochanek KD, Murphy SL, Xu J, Arias E (2019) Deaths: Final Data for 2017. *Natl Vital Stat Rep* 68(9): 1-77.
8. Koyama Y, Brenner DA (2017) Liver inflammation and fibrosis. *J Clin Invest* 127(1): 55-64.
9. Hopkins M, Tyson JJ, Novák B (2017) Cell-cycle transitions: a common role for stoichiometric inhibitors. *Mol Biol Cell* 28(23): 3437-3446.
10. Xu Z, Li T, Li M, Yang L, Xiao R, et al. (2018) eRF3b-37 inhibits the TGF- β -induced activation of hepatic stellate cells by regulating cell proliferation, G0/G1 arrest, apoptosis and migration. *Int J Mol Med* 42(6): 3602-3612.
11. Friedman SL (2003) Liver fibrosis -- from bench to bedside. *J Hepatol* 1: S38-S53.
12. Zheng H, Wang X, Zhang Y, Chen L, Hua L, et al. (2019) Pien-Tze-Huang ameliorates hepatic fibrosis via suppressing NF- κ B pathway and promoting HSC apoptosis. *J Ethnopharmacol* 244: 111856.
13. Ramadori G, Saile B (2004) Portal tract fibrogenesis in the liver. *Lab Invest* 84(2): 153-159.
14. Naito M, Hasegawa G, Ebe Y, Yamamoto T (2004) Differentiation and function of Kupffer cells. *Med Electron Microsc* 37(1): 16-28.
15. Maher JJ (2001) Interactions between hepatic stellate cells and the immune system. *Semin Liver Dis* 21(3): 417-426.
16. Canbay A, Friedman S, Gores GJ (2004) Apoptosis: the nexus of liver injury and fibrosis. *Hepatology* 39(2): 273-278.
17. Tao YY, Yan XC, Zhou T, Shen L, Liu ZL, et al. (2014) Fuzheng Huayu recipe alleviates hepatic fibrosis via inhibiting TNF- α induced hepatocyte apoptosis. *BMC Complement Altern Med* 14: 449.
18. Jiang JX, Mikami K, Venugopal S, Li Y, Török NJ (2009) Apoptotic body engulfment by hepatic stellate cells promotes their survival by the JAK/STAT and Akt/NF- κ B-dependent pathways. *J Hepatol* 51(1): 139-148.
19. Han R, Chen XY (2019) Apoptotic protease activating factor-1 negatively regulates Wnt signaling in hepatocellular carcinoma. *Kaohsiung J Med Sci* 35(8): 459-466.
20. Viñas O, Bataller R, Sancho-Bru P, Ginès P, Berenguer C, et al. (2003) Human hepatic stellate cells show features of antigen-presenting cells and stimulate lymphocyte proliferation. *Hepatology* 38(4): 919-929.
21. Gressner AM, Weiskirchen R (2006) Modern pathogenetic concepts of liver fibrosis suggest stellate cells and TGF-beta as major players and therapeutic targets. *J Cell Mol Med* 10(1): 76-99.
22. Alatas FS, Matsuura T, Pudjiadi AH, Wijaya S, Taguchi T (2020) Peroxisome Proliferator-Activated Receptor Gamma Agonist Attenuates Liver Fibrosis by Several Fibrogenic Pathways in an Animal Model of Cholestatic Fibrosis. *Pediatr Gastroenterol Hepatol Nutr* 23(4): 346-355.
23. Farrell GC, Larter CZ (2006) Nonalcoholic fatty liver disease: from steatosis to cirrhosis. *Hepatology* 43(2 Suppl 1): S99-S112.
24. Kisseleva T, Brenner DA (2006) Hepatic stellate cells in fibrogenesis and the reversal of fibrosis. *J Gastroenterol Hepatol Suppl* 3: S84-S87.
25. Mangasser-Stephan K, Gartung C, Lahme B, Gressner AM (2001) Expression of isoforms and splice variants of the latent transforming growth factor beta binding protein (LTBP) in cultured human liver myofibroblasts. *Liver* 21(2): 105-113.
26. Roth S, Gong W, Gressner AM (1998) Expression of different isoforms of TGF-beta and the latent TGF-beta binding protein (LTBP) by rat Kupffer cells. *J Hepatol* 29(6): 915-922.
27. Roth Eichhorn S, Kühl K, Gressner AM (1998) Subcellular localization of (latent) transforming growth factor beta and the latent TGF-beta binding protein in rat hepatocytes and hepatic stellate cells. *Hepatology* 28(6): 1588-1596.
28. Friedman SL (2008) Hepatic stellate cells: protean, multifunctional, and enigmatic cells of the liver. *Physiol Rev* 88(1): 125-172.
29. Eissa LA, Kenawy HI, El Karef A, Elsherbiny NM, El Mihi KA (2018) Antioxidant and anti-inflammatory activities of berberine attenuate hepatic fibrosis induced by thioacetamide injection in rats. *Chem Biol Interact* 294: 91-100.
30. Friedman SL (1999) Cytokines and fibrogenesis. *Semin Liver Dis* 19(2): 129-140.
31. Gressner AM, Weiskirchen R, Breitkopf K, Dooley S (2002) Roles of TGF-beta in hepatic fibrosis. *Front Biosci* 7: d793-807.
32. Martin Mateos R, De Assuncao TM, Arab JP, Jalan Sakrikan N, Yaqoob U, et al. (2019) Enhancer of Zeste Homologue 2 Inhibition Attenuates TGF- β Dependent Hepatic Stellate Cell Activation and Liver Fibrosis. *Cell Mol Gastroenterol Hepatol* 7(1): 197-209.
33. Watson CJ, Collier P, Tea I, Neary R, Watson JA, et al. (2014) Hypoxia-induced epigenetic modifications are associated with cardiac tissue fibrosis and the development of a myofibroblast-like phenotype. *Hum Mol Genet* 23(8): 2176-2188.
34. Matsumura N, Huang Z, Mori S, Baba T, Fujii S, et al. (2011) Epigenetic suppression of the TGF-beta pathway revealed by transcriptome profiling in ovarian cancer. *Genome Res* 21(1): 74-82.
35. Fan Z, Hao C, Li M, Dai X, Qin H, et al. (2015) MKL1 is an epigenetic modulator of TGF- β induced fibrogenesis. *Biochim Biophys Acta* 1849(9): 1219-1228.
36. Piek E, Helden CH, Ten Dijke P (1999) Specificity, diversity, and regulation in TGF-beta superfamily signaling. *FASEB J* 13(15): 2105-2124.
37. Lan HY, Chung ACK (2011) Transforming growth factor- β and Smads. *Contrib Nephrol* 170: 75-82.
38. Xu F, Liu C, Zhou D, Zhang L (2016) TGF- β /SMAD Pathway and Its Regulation in Hepatic Fibrosis. *J Histochem Cytochem* 64(3): 157-167.
39. Luo H, Zhao F, Zhang F, Liu N (2018) Influence of amygdalin on PDG, IGF and PDGFR expression in HSC-T6 cells. *Exp Ther Med* 15(4): 3693-3698.
40. Kikuchi A, Pradhan Sundd T, Singh S, Nagarajan S, Loizos N, et al. (2017) Platelet-Derived Growth Factor Receptor α Contributes to Human Hepatic Stellate Cell Proliferation and Migration. *Am J Pathol* 187(10): 2273-2287.
41. Di Sario A, Bendia E, Svegliati-Baroni G, Marziani M, Ridolfi F, et al. (2002) Rearrangement of the cytoskeletal network induced by platelet-derived growth factor in rat hepatic stellate cells: role of different intracellular signalling pathways. *J Hepatol* 36(2): 179-190.
42. Cao S, Yaqoob U, Das A, Shergill U, Jagavelu K, et al. (2010) Neuropilin-1 promotes cirrhosis of the rodent and human liver by enhancing PDGF/TGF-beta signaling in hepatic stellate cells. *J Clin Invest* 120(7): 2379-2394.
43. Pinzani M, Marra F (2001) Cytokine receptors and signaling in hepatic stellate cells. *Semin Liver Dis* 21(3): 397-416.
44. Jiang G, Wang W, Cao Q, Gu J, Mi X, et al. (2015) Insulin growth factor-1 (IGF-1) enhances hippocampal excitatory and seizure activity through

- IGF-1 receptor-mediated mechanisms in the epileptic brain. *Clin Sci (Lond)* 129(12): 1047-1060.
45. Saile B, DiRocco P, Dudas J, El-Armouche H, Sebb H, et al. (2004) IGF-I induces DNA synthesis and apoptosis in rat liver hepatic stellate cells (HSC) but DNA synthesis and proliferation in rat liver myofibroblasts (rMF). *Lab Invest* 84(8): 1037-1049.
 46. Gallaher BW, Hille R, Raile K, Kiess W (2001) Apoptosis: live or die--hard work either way! *Horm Metab Res* 33(9): 511-519.
 47. Kuemmerle JF, Zhou H, Bowers JG (2004) IGF-I stimulates human intestinal smooth muscle cell growth by regulation of G1 phase cell cycle proteins. *Am J Physiol Gastrointest Liver Physiol* 286(3): G412-G419.
 48. Chakravarthy MV, Abraha TW, Schwartz RJ, Fiorotto ML, Booth FW (2000) Insulin-like growth factor-I extends in vitro replicative life span of skeletal muscle satellite cells by enhancing G1/S cell cycle progression via the activation of phosphatidylinositol 3'-kinase/Akt signaling pathway. *J Biol Chem* 275(46): 35942-35952.
 49. Pellicoro A, Ramachandran P, Iredale JP, Fallowfield JA (2014) Liver fibrosis and repair: immune regulation of wound healing in a solid organ. *Nat Rev Immunol* 14(3): 181-194.
 50. Casini A, Ceni E, Salzano R, Biondi P, Parola M, et al. (1997) Neutrophil-derived superoxide anion induces lipid peroxidation and stimulates collagen synthesis in human hepatic stellate cells: role of nitric oxide. *Hepatology* 25(2): 361-367.
 51. Deng ZY, Li J, Jin Y, Chen XL, Lü XW (2009) Effect of oxymatrine on the p38 mitogen-activated protein kinases signalling pathway in rats with CCl4 induced hepatic fibrosis. *Chin Med J (Engl)* 122(12): 1449-1454.
 52. Fabregat I, Caballero-Díaz D (2018) Transforming Growth Factor- β -Induced Cell Plasticity in Liver Fibrosis and Hepatocarcinogenesis. *Front Oncol* 8: 357.
 53. Canbay A, Friedman S, Gores GJ (2004) Apoptosis: the nexus of liver injury and fibrosis. *Hepatology* 39(2): 273-278.
 54. Troeger JS, Mederacke I, Gwak GY, Dapito DH, Mu X, et al. (2012) Deactivation of hepatic stellate cells during liver fibrosis resolution in mice. *Gastroenterology* 143(4): 1073-1083.
 55. Cheng CF, Pan TM (2016) Ankaflavin and Monascin Induce Apoptosis in Activated Hepatic Stellate Cells through Suppression of the Akt/NF- κ B/p38 Signaling Pathway. *J Agric Food Chem* 64(49): 9326-9334.
 56. Gressner OA, Gressner AM (2008) Connective tissue growth factor: a fibrogenic master switch in fibrotic liver diseases. *Liver Int* 28(8): 1065-1079.
 57. Varga J, Greten FR (2017) Cell plasticity in epithelial homeostasis and tumorigenesis. *Nat Cell Biol* 19(10): 1133-1141.
 58. Huard J, Mueller S, Gilles ED, Klingmüller U, Klamt S (2012) An integrative model links multiple inputs and signaling pathways to the onset of DNA synthesis in hepatocytes. *FEBS J* 279(18): 3290-3313.
 59. Kubota K, Okazaki J, Louie O, Kent KC, Liu B (2003) TGF- β stimulates collagen (I) in vascular smooth muscle cells via a short element in the proximal collagen promoter. *J Surg Res* 109(1): 43-50.
 60. Yang KL, Chang WT, Hong MY, Hung KC, Chuang CC (2017) Prevention of TGF- β -induced early liver fibrosis by a maleic acid derivative anti-oxidant through suppression of ROS, inflammation and hepatic stellate cells activation. *PLoS One* 12(4): e0174008.
 61. Gooch JL, Gorin Y, Zhang BX, Abboud HE (2004) Involvement of calcineurin in transforming growth factor-beta-mediated regulation of extracellular matrix accumulation. *J Biol Chem* 279(15): 15561-15570.
 62. Pan XY, You HM, Wang L, Bi YH, Yang Y, et al. (2019) Methylation of RCAN1.4 mediated by DNMT1 and DNMT3b enhances hepatic stellate cell activation and liver fibrogenesis through Calcineurin/NFAT3 signaling. *Theranostics* 9(15): 4308-4323.
 63. Ramadori G, Saile B (20002) Mesenchymal cells in the liver--one cell type or two? *Liver* 22(4): 283-294.
 64. Debonera F, Aldeguer X, Shen X, Gelman AE, Gao F, et al. (20010) Activation of interleukin-6/STAT3 and liver regeneration following transplantation. *J Surg Res* 96(2): 289-295.
 65. Huh CG, Factor VM, Sánchez A, Uchida K, Conner EA, Thorgeirsson SS (2004) Hepatocyte growth factor/c-met signaling pathway is required for efficient liver regeneration and repair. *Proc Natl Acad Sci U S A* 101(13): 4477-4482.
 66. Fausto N, Riehle KJ (2005) Mechanisms of liver regeneration and their clinical implications. *J Hepatobiliary Pancreat Surg* 12(3): 181-189.
 67. Gao L, Utsumi T, Tashiro K, Liu B, Zhang D, et al. (2013) Reticulon 4B (Nogo-B) facilitates hepatocyte proliferation and liver regeneration in mice. *Hepatology* 57(5): 1992-2003.
 68. Pellicoro A, Ramachandran P, Iredale JP, Fallowfield JA (2014) Liver fibrosis and repair: immune regulation of wound healing in a solid organ. *Nat Rev Immunol* 14(3): 181-194.
 69. Jin X, Zhang Z, Beer-Stolz D, Zimmers TA, Koniaris LG (2007) Interleukin-6 inhibits oxidative injury and necrosis after extreme liver resection. *Hepatology* 46(3): 802-812.
 70. Quétier I, Brezillon N, Duriez M, Massinet H, Giang E, et al. (2013) Hepatitis B virus HBx protein impairs liver regeneration through enhanced expression of IL-6 in transgenic mice. *J Hepatol* 59(2): 285-291.
 71. Tachibana S, Zhang X, Ito K, Ota Y, Cameron AM, et al. (2014) Interleukin-6 is required for cell cycle arrest and activation of DNA repair enzymes after partial hepatectomy in mice. *Cell Biosci* 4(1): 6.
 72. Michalopoulos GK (2007) Liver regeneration. *J Cell Physiol* 213(2): 286-300.
 73. Swanton C (2004) Cell-cycle targeted therapies. *Lancet Oncol* 5(1): 27-36.
 74. Gentilini A, Marra F, Gentilini P, Pinzani M (2000) Phosphatidylinositol-3 kinase and extracellular signal-regulated kinase mediate the chemotactic and mitogenic effects of insulin-like growth factor-I in human hepatic stellate cells. *J Hepatol* 32(2): 227-234.
 75. Green DR (2000) Apoptotic pathways: paper wraps stone blunts scissors. *Cell* 102(1): 1-4.
 76. Kang LI, Mars WM, Michalopoulos GK (2012) Signals and cells involved in regulating liver regeneration. *Cells* 1(4): 1261-1292.
 77. Nakamura T, Nishizawa T, Hagiya M, Seki T, Shimonishi M, et al. (1989) Molecular cloning and expression of human hepatocyte growth factor. *Nature* 342(6248): 440-443.
 78. Fafalios A, Ma J, Tan X, Stoops J, Luo J, et al. (2011) A hepatocyte growth factor receptor (Met)-insulin receptor hybrid governs hepatic glucose metabolism. *Nat Med* 17(12): 1577-1584.
 79. Wang B, Gao C, Ponder KP (2005) C/EBP β contributes to hepatocyte growth factor-induced replication of rodent hepatocytes. *J Hepatol* 43(2): 294-302.
 80. Ross J, Gherardi E, Mallorqui-Fernandez N, Bocci M, Sobkowicz A, et al. (2012) Protein engineered variants of hepatocyte growth factor/scatter factor promote proliferation of primary human hepatocytes and in rodent liver. *Gastroenterology* 142(4): 897-906.
 81. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, et al. (2018) Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality

- worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* 68(6): 394-424.
82. Affo S, Yu LX, Schwabe RF (2017) The Role of Cancer-Associated Fibroblasts and Fibrosis in Liver Cancer. *Annu Rev Pathol* 12: 153-186.
83. Heymann F, Trautwein C, Tacke F (2009) Monocytes and macrophages as cellular targets in liver fibrosis. *Inflamm Allergy Drug Targets* 8(4): 307-318.
84. Dooley S, ten Dijke P (2012) TGF- β in progression of liver disease. *Cell Tissue Res* 347(1): 245-256.
85. Issa R, Williams E, Trim N, Kendall T, Arthur MJ, et al. (2001) Apoptosis of hepatic stellate cells: involvement in resolution of biliary fibrosis and regulation by soluble growth factors. *Gut* 48(4): 548-557.
86. Giannelli G, Rani B, Dituri F, Cao Y, Palasciano G. (2014) Moving towards personalised therapy in patients with hepatocellular carcinoma: the role of the microenvironment. *Gut* 63(10): 1668-1676.