



Mini Review

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Two Sorts of DNA in Eukaryotic Cells: Linear Chromosomal DNA and Circular Extrachromosomal DNA

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Abstract

In addition to Chromosomal DNA (chr-DNA), eukaryotic cells contain Extrachromosomal DNA (ec-DNA), e.g., Mitochondrial DNA (mt-DNA), and a particular class named Extrachromosomal Circular DNA (ecc-DNA). The ecc-DNA structures harbor functional sequences known to be present in the chr-DNA. Ecc-DNA has many sizes, from thousands of base pairs to mega base pairs. Their proportion of the total cellular DNA can be up to 20%, both in cells from healthy people and people with diseases, for example, cancer. A research focus is an exploration of how ecc-DNA is involved in oncology. The possible influence of ecc-DNA on chromosomal sequences in the Human Genome Project gets addressed.

Keywords: ecc-DNA, Genetics, Human Genome, Diseases, Diagnostic Reservoir

Introduction

This article describes certain basics of Chromosomal DNA (chr-DNA) and Extrachromosomal Circular DNA (ecc-DNA). Research indicates genetic activities at the level of ecc-DNA in cells with potential relevance to diseases. This concerns an oncogenic potential.

Human Chromosomes are the source of sequences contained in ecc-DNA

The linear chromosomes apart from mt-DNA in eukaryotes cells contain DNA indicating the organism's blueprint. The protein-coding sequences making up the genes represent less than 2% of the total chr-DNA [1]; careful consideration of the remaining sequences is necessary [2]. The "telomere-to-telomere" project achieved a "...gapless, telomere-to-telomere assembly of a human chromosome" [3].

Gene-coding DNA sequences are not present as a continuous sequence stretch (statements range from hypothesis to a general fact); structured in discrete protein-coding subunit sequences, exons, interspersed among non-protein-coding sequences, introns [4]. Following the "intron-exon architecture of eukaryotic genes," this structure is reviewed as a "genes in pieces" model [5]. Studies are required to determine how this model can be consistently resilient. In addition to the multiple-step formation of "functional" genes with their authentic gene pieces, recombination between nonrelated exons can arise: Recombination and "exon shuffling," after transcription to mRNA generate "new proteins with novel combinations of protein domains" [6,7]. With the continuous improvements of the molecular protocols, the "number of errors when mapping ..." could be reduced [8].

Which mechanisms are responsible for selecting genuine sequences from linear chr-DNA and assembling them into



circular double-stranded DNA, for example, containing functional oncogenes?

Endogenous Chromosomal tools are both able to move positions of Sequences within the Chr-DNA and to Generate Extrachromosomal DNA (Ec-DNA)

Apart from various protein-coding sequences, the remaining chromosomal sequences belong to the “non-coding regions”-not coding for proteins. At least parts of these DNA sequences code for functional tasks: e. g., after transcription, there are diverse forms of RNA that act as regulatory factors. Furthermore, Mobile Genetic Elements (MGE) [9] such as ALU-elements, transposons (jumping genes), and retrotransposons have various functions; e. g., “... how it contributes to human disease” [10].

Certain parts of the “non-coding regions” of the human genome have received attention [11]; for example, sequences such as “enhancers” are involved in the development of cancer [12]. Even genetic ablation of a particular RNA locus could cause a “genetic disorder” that leads to organic malformations [13].

Sequences that exert both structural and functional DNA properties

Endogenous Retroviruses (ERVs) “ ... are remnants of ancient active retroviruses that infected germline cells” [14]. This process ensures that they get passed on from generation to generation. The ERV-DNA copies became an integrated part of human chromosomes, then named HERV. The genome sequences of HERV-K make up approximately 8% of the human genome [15], about four times more than the protein-coding sequences of “human genes.”

Studies indicate that the integrated HERV-K sequences are under genetic and epigenetic control [16]. There seems to be a broad consensus that HERVs cannot replicate [17,18]. But these integrated sequences (selected ones?) can be reactivated: upon exogenous/endogenous impact/s? [19] They have pathogenic potentials, such as inducing (?) / promoting (?) neurodegenerative diseases, autoimmune diseases, and multiple cancers [20-22]. If parts of HERV K’s genome are present in ecc-DNA, does their pathogenetic potential contribute to causing diseases from this level?

The control of DNA “activities”

Several epigenetic mechanisms control the activities of genomic sequences: e. g., DNA methylation, histone post-translational modifications, and microRNAs (miRNAs). DNA methylation/demethylation is adding/removing methyl groups to one or two components of DNA, specifically the deoxyribonucleotides adenine

and cytosine. DNA methylation/demethylation regulates gene expression. Epigenetic modifications don’t alter the underlying chromosomal DNA sequence. The project “Epigenetic Patterns in a Complete Human Genome” explores the full epigenome [23].

Extrachromosomal DNA (ec-DNA)

Besides chr-DNA and mitochondrial DNA, eukaryotic cells contain ec-DNA, generated by excision/copying sequences from the chromosomes; the underlying principles of selecting sequences are unknown. Sequences for functional genes also reside in Extrachromosomal (ec) elements. Extrachromosomal circular DNA (ecc-DNA) in eukaryotic cells has been known for a long time [24].

Extrachromosomal circular DNA (ecc-DNA)

The origin, genetic activities, and their control

Various ways describe the generation of ecc-DNA, such as during apoptotic cell disintegration [25] or repair; [26] originating from the chromosomes [27]. Depending on their size, they may contain protein-coding and properly non-coding sequences and sequences derived from MGE; MGE can mediate intra- or intercellular DNA trafficking [28]. The composition of the sequences may be subject to metabolic pressure. The term “ecc-DNA” describes the full spectrum of circular DNAs in eukaryotes [29]. Their sizes range from a few thousand base pairs to mega base pairs. What are the basics for selecting chr-DNA-derived sequences that become incorporated into ecc-DNA?

What kind of superordinate control regulates “genetic activities” of ecc-DNA [30]? Data hint that epigenetic mechanisms such as DNA methylation control ecc-DNA - an operational regime.

There is growing consensus that faulty DNA methylation might be an essential factor in developing diseases. Research focuses on the links between chr-DNA and ecc-DNA methylation and human diseases such as cancer and various congenital disabilities. So far, much of this research targets cancer and tumor suppressor genes. Is there is an ongoing exchange of sequences between chr-DNA and ecc-DNA?

The proportion of ecc-DNA of whole cellular DNA increases in eukaryotic cells with cell stress or aging and “contribute to intercellular heterogeneity in normal and tumor cells” [31]. Studies have indicated that cells containing ecc-DNA are prone to develop diseases – for example, ecc-DNA with several 100kbp may increase the oncogenic potential of cells [32].

An oncogene is a gene “... that is a mutated (changed) form of a gene involved in average cell growth. Oncogenes may cause the growth of cancer cells “[33] Such mutations provide characteristics

to the sequence that render it gradually/completely resistant to epigenetic control mechanisms.

These findings apply to the connection of ecc-DNA in “oncogene amplification” [34], the generation of tumors [35-39], their pathogenesis [40-42], and their acceleration [43].

Ecc-DNA is particularly abundant in multiple human cancer cells, although its frequency varies among different tumor types. Elevated levels of ecc-DNA have been considered an effective biomarker of cancer pathogenesis. Reports have demonstrated the amplification/evolution of oncogenes and therapeutic resistance genes located on ecc-DNA [44-46]. The molecular cascades from proto-oncogenes to functional oncogenes seem ordinary events that may drive intratumor genetic heterogeneity. Are “tumor suppressor genes” effective on “oncogenes” encoded in both chromosomal and extrachromosomal DNA? Specific details have revealed how ecc-DNA “remodel” genomic DNA [47]. ecc-DNA also has implications in degenerative processes in the central nervous system [48]. Their features might provide the basics for developing biomarkers [49] and applications designed for diagnostic purposes [50,51]. Diagnostics based on cell-free, circulating tumor DNA (ct-DNA) in the plasma should be standardized when evaluating samples from the serum/plasma of patients presumed or already confirmed to have cancer [52-54]. Is discrimination required to determine the origin of the selected sequence, namely from chr-DNA (genes in pieces) or ecc-DNA (composed of sequence pieces of chromosomal basis)?

Exceptional case/s?

Ecc-DNA is also detectable in healthy individuals' Peripheral Blood Mononuclear Cells (PBMCs). The investigated samples contain a sequence that is part of the 5'-non-coding region (5'-NCR) of the Hepatitis C Virus (HCV), a single-stranded RNA virus but not a retrovirus! This DNA sequence section is part of the Internal Ribosomal Entry Site (IRES) of HCV [55]. First, it shows individual patterns of methylation (methylomes)-epigenetically controlled? [56]. Second, the methylation patterns of this HCV-DNA sequence section have changed over time. A possible interpretation of this phenomenon is that the changing methylation patterns of this sequence may indicate a particular active control mechanism. The origin of this sequence section of the 5'-NCR present in the ecc-DNA may be from a chromosomal basis [57].

The possible impact of ecc-DNA on the Human Genome Project (HGP)

The goals of the HGP are to reveal the DNA sequences of human chromosomes as a basis for gene identification. The constitutive considerations about a human reference genome [58] do not mention ecc-DNA as a possible falsifying factor on the chromosomal

sequences. The point is that only the sequestration of ecc-DNA from chr-DNA may warrant pure/genuine chr-DNA sequences. The ECCs plorer protocol [59] may be an appropriate tool for this task. A database for ecc-DNA is available [60]. In consideration of new details regarding ecc-DNA, there are requests for a new human reference genome [61]. Concerning the human genome project using whole cellular DNA: How was the possible mixing of sequences of chr-DNA and ecc-DNA excluded when designing the human reference genome?

Final comment

Ecc-DNA is present in eukaryotic cells, but what induces their generation is unclear: individual genetic rearrangements due to endogenous triggers, exogenous elicitors such as “onco viruses,” e. g. EBV? So far, the ongoing research has drawn attention to ecc-DNA because of its involvement in cancer development and possible use in diagnostics. It seems appropriate to consider testing for ecc-DNA with oncogenic coding potential in individuals suspected of having a tumor.

Perspectives: Eukaryotic cells can contain high numbers of ecc/ecc DNA with coding sequences. Therefore, the definite separation of ecc/ecc DNA from chromosomes must be guaranteed before “genome sequencing” for the HGP to rule out any possible mixing with chromosomal DNA. This occurrence also applies to the re-incorporation of eccDNA into chromosomes [62]. Besides the chromosomes, the eccDNA fraction should be considered when performing diagnostics based on the patient's sequence data.

References

1. Sana J, Faltejskova P, Svoboda M, Slaby O (2012) Novel classes of non-coding RNAs and cancer. *J Transl Med* 10: 103.
2. Clamp M, Fry B, Kamal M, Xie X, Cuff J, et al. (2007) Distinguishing protein-coding and noncoding genes in the human genome. *Proc Natl Acad Sci USA* 104: 19428-19433.
3. Miga KH, Koren S, Rhie A, Vollger MR, Gershman A, et al. (2020) Telomere-to-telomere assembly of a complete human X chromosome. *Nature* 585: 79-84.
4. Rogozin IB, Sverdlov AV, Babenko VN, Koonin EV (2005) Analysis of evolution of exon-intron structure of eukaryotic genes. *Brief Bioinform* 6: 118-134.
5. Smithers B, Oates M, Gough J (2019) Why genes in pieces?-revisited. *Nucleic Acids Res* 47(10): 4970-4973.
6. Berk AJ (2016) Discovery of RNA splicing and genes in pieces. *Proc Natl Acad Sci USA* 113(4): 801-805.
7. Kolkman JA, Stemmer WP (2001) Directed evolution of proteins by exon shuffling. *Nat Biotechnol* 19(5): 423-428.
8. Knutsen A (2020) A new human reference genome represents common sequences.
9. Miller WJ, Cagy P (2004) Mobile genetic elements as natural tools for genome evolution. *Methods Mol Biol* 260: 1-20.

10. Kazazian HH, Moran JV (2017) Mobile DNA in health and disease. *N Engl J Med* 377: 361-370.
11. Vitsios D, Dhindsa RS, Middleton L, Gussow AB, Petrovski S (2021) Prioritizing non-coding regions based on human genomic constraint and sequence context with deep learning. *Nat Commun* 12(1): 1-4.
12. Cao K, Shilatifard A (2018) Enhancers in cancer: genetic and epigenetic deregulation. *Encyclopedia of Cancer*. Elsevier, New York p.559-568.
13. Allou L, Balzano S, Magg A, Quinodoz M, Royer Bertrand B, et al. (2021) Noncoding deletions identify Maenli lncRNA as a limb specific En1 regulator. *Nature* 592(7852): 93-98.
14. Johanning GL, Malouf GG, Zheng X, Esteva FJ, Weinstein JN, et al. (2017) Expression of human endogenous retrovirus-K is strongly associated with the basal-like breast cancer phenotype. *Sci Rep* 7: 41960.
15. Shin W, Lee J, Son SY, Ahn K, Kim HS, et al. (2013) Human-specific HERV-K insertion causes genomic variations in the human genome. *PLoS One* 8(4): 60605.
16. Vincendeau M, Göttesdorfer I, Schreml JMH, Wette AGN, Mayer J, et al. (2015) Modulation of human endogenous retrovirus (HERV) transcription during persistent and de novo HIV-1 infection. *Retrovirology* 12: 27.
17. Lee, Y N, Bieniasz PD (2007) Reconstitution of an infectious human endogenous retrovirus. *PLoS pathogens* 3(1): 10.
18. Hohn O, Hanke K, Bannert N (2013) HERV-K(HML-2), the best-preserved family of HERVs: endogenization, expression, and implications in health and disease. *Front Oncol*.
19. Xue B, Sechi LA, Kelvin DJ (2020) Human endogenous retrovirus K (HML-2) in health and disease. *Front Microbiol* 11: 1690.
20. Dembny P, Newman AG, Singh M, Hinz M, Szczepek M, et al. (2020) Human endogenous retrovirus HERV-K(HML-2) RNA causes neurodegeneration through Toll-like receptors. *JCI Insight* 5(7): 131093.
21. Nelson PN, Carnegie PR, Martin J, Davari Ejtehadi H, Hooley P, et al. (2003) Demystified. Human endogenous retroviruses. *Mol Pathol* 56(1): 11-18.
22. Mustelin T, Ukadike KC (2020) How retroviruses and retrotransposons in our genome may contribute to autoimmunity in rheumatological conditions. *Front Immunol* 11: 593891.
23. Gershman A, Sauria MEG, Hook PW, Hoyt SJ, Razaghi R, et al. (2021) Epigenetic Patterns in a Complete Human Genome.
24. Møller HD, Mohiyuddin M, Prada Luengo I, Sailani MR, Halling JF, et al. (2018) Circular DNA elements of chromosomal origin are common in healthy human somatic tissue. *Nat Commun* 9: 1069.
25. Wang Y, Wang M, Djekidel MN, et al. (2021) eccDNAs are apoptotic products with high innate immunostimulatory activity. *Nature* 599: 308-314.
26. Chiu RWK, Dutta A, Hensson AG, Lo YMD, Mischel P, et al. (2020) What Is extrachromosomal circular DNA and what does it do? *Clin Chem* 66(6): 754-759.
27. Cohen S, Agmon N, Sobol O, Segal D (2010) Extrachromosomal circles of satellite repeats and 5S ribosomal DNA in human cells. *Mob DNA* 1: 11.
28. Frost LS, Leplae R, Summers AO, Toussaint A (2005) Mobile genetic elements: the agents of open-source evolution. *Nat Rev Microbiol* 3: 722-732.
29. Wang M, Chen X, Yu F, Ding H, Zhang Y, et al. (2021) Extrachromosomal Circular DNAs: Origin, formation, and emerging function in Cancer. *International journal of biological sciences*, 17(4): 1010-1025.
30. Kumar S, Chinnusamy V, Mohapatra T (2018) Epigenetics of modified DNA bases: 5-methylcytosine and beyond. *Front Genet* 9: 640.
31. Kumar P, Kiran S, Saha S, Su Z, Paulsen T, et al. (2020) ATAC-seq identifies thousands of extrachromosomal circular DNA in cancer and cell lines. *Sci Adv* 6: 2489.
32. Cao X, Wang S, Ge L, Zhang W, Huang J, et al. (2021) Extrachromosomal circular DNA: category, biogenesis, recognition, and functions. *Front Vet Sci* 8: 693-641.
33. Shimizu N (2021) Gene Amplification, and the Extrachromosomal Circular DNA. *Genes* 12: 1533.
34. Verhaak RGW, Bafna V, Mischel PS (2019) Extrachromosomal oncogene amplification in tumour pathogenesis and evolution. *Nat Rev Cancer* 19(5): 283-288.
35. Liao Z, Jiang W, Ye L, Li T, Yu X, et al. (2020) Classification of extrachromosomal circular DNA with a focus on the role of extrachromosomal DNA (eccDNA) in tumor heterogeneity and progression. *Biochim Biophys Acta Rev Cancer* 1874(1): 188392.
36. Wang T, Zhang H, Zhou Y, Shi J (2021) Extrachromosomal circular DNA: a new potential role in cancer progression. *J Transl Med* 19: 257.
37. Li RY, Liang ZY (2020) Circulating tumor DNA in lung cancer: real-time monitoring of disease evolution and treatment response. *Chin Med J (Engl)* 133(20): 2476-2485.
38. Ling X, Han Y, Meng J, Zhong B, Chen J, et al. (2021) Small extrachromosomal circular DNA (eccDNA): major functions in evolution and cancer. *Mol Cancer* 20: 113.
39. Hong J, Zheng S, Jiang D (2021) The contributions of extrachromosomal DNA elements in neoplasm progression. *Am J Cancer Res* 11(6): 2417-2429.
40. Yan Y, Guo G, Huang J, Gao M, Zhu Q, et al. (2020) Current understanding of extrachromosomal circular DNA in cancer pathogenesis and therapeutic resistance. *J Hematol Oncol* 13: 124.
41. Kim H, Nguyen NP, Turner K, Wu S, Gujar AD, et al. (2020) Extrachromosomal DNA is associated with oncogene amplification and poor outcome across multiple cancers. *Nat Genet* 52: 891-897.
42. Zuo, S, Yi Y, Wang C, Li X, Zhou M, et al. (2022) Extrachromosomal Circular DNA (eccDNA): From Chaos to Function. *Frontiers in Cell and Developmental Biology* 82(1): 209-217.
43. Bailey C, Shoura MJ, Mischel PS, Swanton C (2020) Extrachromosomal DNA-relieving heredity constraints, accelerating tumour evolution. *Ann Oncol* 31(7): 884-893.
44. Robert M, Crasta K (2021) Breaking the vicious circle: Extrachromosomal circular DNA as an emerging player in tumour evolution. *Semin Cell Dev Biol* 123:140-150.
45. Hui Qiu, Zhi Ying Shao, Xin Wen, Long Zhen Zhang (2020) New insights of extrachromosomal DNA in tumorigenesis and therapeutic resistance of cancer. *Am J Cancer Res* 10(12): 4056-4065.
46. Wu S, Bafna V, Chang HY, Mischel PS (2022) Extrachromosomal DNA: An Emerging Hallmark in Human Cancer. *Annu Rev Pathol* 24(17): 367-386.
47. Koche RP, Rodriguez Fos E, Helmsauer K, Burkert M, MacArthur IC, et al. (2020) Extrachromosomal circular DNA drives oncogenic genome remodeling in neuroblastoma. *Nat Genet* 52(1): 29-34.
48. Ain Q, Schmeer C, Wengerodt D, Witte OW, Kretz A (2020) Extrachromosomal circular DNA: current knowledge and implications for CNS aging and neurodegeneration. *Int J Mol Sci* 21(7): 2477.
49. Sin STK, Jiang P, Deng J, Ji L, Cheng SH, et al. (2020) Identification and characterization of extrachromosomal circular DNA in maternal plasma. *Proc Natl Acad Sci USA* 117(3): 1658-1665.

50. Zhu J, Chen S, Zhang F, Wang L (2018) Cell-free eccDNAs: a new type of nucleic acid component for liquid biopsy? *Mol Diagn Ther* 22(5): 515-522.
51. Keller L, Belloum Y, Wikman H, Pantel K (2021) Clinical relevance of blood-based ctDNA analysis: mutation detection and beyond. *Br J Cancer* 124: 345-358.
52. Bratman SV, Yang SYC, Iafolla MAJ, Liu Z, Hansen AR, et al. (2020) Personalized circulating tumor DNA analysis as a predictive biomarker in solid tumor patients treated with pembrolizumab. *Nat Cancer* 1(9): 873-881.
53. Keller L, Belloum Y, Wikman H, Pantel K (2021) Clinical relevance of blood-based ctDNA analysis: mutation detection and beyond. *Br J Cancer* 124: 345-358.
54. Said R, Guibert N, Oxnard GR, Tsimberidou AM (2020) Circulating tumor DNA analysis in the era of precision oncology. *Oncotarget* 11: 188-211.
55. Dennin RH, Wo JE (2019) DNA sequences homologous to Hepatitis C Virus (HCV) in the extrachromosomal circular DNA in peripheral blood mononuclear cells of HCV-negative subjects. *J Zhejiang Univ Sci B* 20: 637-646.
56. Zhou W, Liang G, Molloy PL, Jones PA (2020) DNA methylation enables transposable element-driven genome expansion. *Proc Natl Acad Sci USA* 117: 19359-19366.
57. Kanduc D (2011) HCV - Written in our DNA. *Self/Nonself* 2(2): 108-113.
58. Miga KH (2021) Breaking through the unknowns of the human reference genome. *Nature* 590(7845): 217-218.
59. Mann L, Seibt KM, Weber B, Heitkam T (2021) ECCsplorer: a pipeline to detect extrachromosomal circular DNA (eccDNA) from next-generation sequencing data. *BioRxiv*.
60. (2020) The extrachromosomal circular DNA database; Homo sapiens. eccDNAdb.
61. Ballouz S, Dobin A, Gillis JA. (2019) Is it time to change the reference genome? *Genome Biol* 20: 159.
62. Mouakkad Montoya L, Murata M, Sulovari A, Suzuki R, Osia B, et al. (2021) Quantitative assessment reveals the dominance of duplicated sequences in germline-derived extrachromosomal circular DNA. *PNAS* 118(47): 2102842118.