



Research Article

Copyright@ Murat Basar

The Assessment of Purity Level of CO₂ Used in Horizontal Incubators and The Effect of Additional In-Line Filters on Embryo Development

Yagmur Ergun¹, Melek Bozkizil², Aydin Arici^{1,3} and Murat Basar^{1,2,3*}

¹Department of Obstetrics, Gynecology and Reproductive Sciences, Yale School of Medicine, USA

²Department of Histology and Embryology, Clinical Embryology Master Program, Biruni University, Turkey

³Yale Fertility Center, USA

***Corresponding author:** Murat Basar, PhD, Biruni University, Graduate Education Institute, Department of Histology and Embryology Address: 10. Yıl Caddesi Protokol Yolu No: 45 34010 Topkapı / İstanbul.

To Cite This Article: Yagmur E, Melek B, Aydin A, Murat B. The Assessment of Purity Level of CO₂ Used in Horizontal Incubators and The Effect of Additional In-Line Filters on Embryo Development. *Am J Biomed Sci & Res.* 2022 17(2) AJBSR.MS.ID.002323, DOI: [10.34297/AJBSR.2022.17.002323](https://doi.org/10.34297/AJBSR.2022.17.002323)

Received: 📅 September 24, 2022; **Published:** 📅 September 28, 2022

Abstract

Objectives: In this study, we aimed to investigate whether carbon dioxide (CO₂) used in horizontal incubators during in vitro fertilization (IVF) is pure enough and whether the addition of In-Line filters affects the stress on blastocysts and blastocyst utilization rate.

Materials and Methods: To examine the effect of the purity of CO₂ gas on embryo quality, two pronuclei (PN) embryos of patients younger than 35 years old and undergoing intracytoplasmic sperm injection (ICSI) were divided into two groups. Embryos in Group 1 were cultured in the incubator, which has an extra CO₂ filter, whilst those in Group 2 (control group) were cultured without any additional filtering. On day 3 (D3), Day 5 (D5), embryo qualities and blastocyst utilization rates between groups were examined. Three different filters with different brands were examined regarding their effects on embryo quality within groups. For animal studies, 10-week-old female BALB/C mice (n=10) were stimulated with Follicle Stimulating Hormone (FSH) followed by Human Chorionic Gonadotropin (hCG) and mated with fertility-proven adult male mice. Mice with vaginal plaques after 24 hours were sacrificed, and 1-cell embryos were collected. Embryos were pooled first and then distributed into Group 1 and Group 2 randomly. To assess the effect of gas purity on embryonic stress, Glucose-regulated Protein 78 (GRP78) level was examined using Western blot method.

Results: CO₂ gas content used in IVF laboratories was not as pure as indicated, and the following impurities were isolated: ethyl methyl ketone, tetrahydrofuran, Fenol, 2-heptanol, and formaldehyde. We found that D3 embryo quality significantly improved within Group 1 compared to Group 2 (p<0,001). Similarly, when the blastocyst utilization rates were assessed, a significant increase was found in all three extra-filtered incubators compared to the ones without filters (p<0,001). Additionally, mouse embryos cultured without an extra filter have higher GRP78 expression than those cultured with an extra CO₂ filter (p<0.001).

Conclusion: It has been shown that the CO₂ gasses used in the IVF laboratory are not as pure as reported. This can be eliminated by using an additional filter, decreasing stress and increasing embryo quality. In addition, it has been shown that the UPR plays a key role in regulating the response to ambient air/gas impurity in embryo development with long-term culture. Our study demonstrates that additional extra CO₂ filters significantly improve embryo quality and impact the UPR pathway in D3 and D5 compared with a control group. This suggests that filtration of CO₂ has a positive impact, perhaps by removing potentially harmful substances or affecting UPR pathways.

Keywords: UPR; Grp78; In-Line Filter; IVF; Western Blot

Introduction

Infertility is defined as difficulty conceiving after one year of regular unprotected intercourse [1]. To diagnose infertility, men and women undergo many investigations. The causes of infertility

can be grouped as follows: 40% of male factors, %40 of female factors, 10% of both, and 10% of idiopathic factors [2]. One of the most critical factors for successful In Vitro Fertilization (IVF) treatment is optimizing the laboratory conditions. The level of CO₂



in the incubators, which is stabilized to 37°C, needs to be monitored regularly to allow the successful development of blastocysts. The most important factor is balancing the pH of the culture media, which can be impacted by inappropriate CO₂ exposure. The pH level is linked to environmental CO₂, and to maintain the balance of the culture conditions in IVF laboratories, CO₂ measurements need to be regularly evaluated [3].

Optimizing and sustaining various culture conditions that impact the viability of embryos and pregnancy rates are crucial. Several factors such as air quality, temperature, and light can affect embryo quality [3,4]. Volatile Organic Compounds (VOCs) are highly toxic for in vitro embryo development [5]. During embryonic development, VOCs directly bind to DNA strands and impact normal development [6,7]. Due to this reason, High-Efficiency Particulate Air (HEPA) filters are used in IVF laboratories to maintain air quality and stability. To remove the toxic particles in the air, positive air pressure is also used alongside conventional filtering systems [8].

It has been shown that Unfolded Protein Response (UPR) plays a crucial role in stress response mechanisms [9]. According to a study published by Basar et al., UPR is induced by increasing ER stress during in vitro embryo development and has a detrimental effect on blastocyst formation, suggesting that activation of UPR signaling may be the cause of low blastocyst development rates [10]. Similarly, an endoplasmic reticulum chaperon protein Grp78 also occurs in early embryo development [11].

In this study, our objective was to assess the purity level of CO₂ gas used in benchtop incubators in IVF laboratories and whether the usage of additional CO₂ filters affects embryo development.

Material and Methods

Animals

To investigate the UPR role in embryo development, ten weeks-old 25-30 grams BALB/C female (n=10) and 10-weeks-old 30-35 grams BALB/C male mice (n=4) were used for the experiments. All animal care and experimental procedures were conducted per Biruni University and Akdeniz University animal research requirements, using protocols approved by the Institutional Animal Care and Use Committee (Protocol # 2019/35-07).

Superovulation of Female Mice and Embryo Collection

10IU of human chorionic gonadotropin (hCG; Sigma, St. Louis, MO) was injected 48h hours after the PMSG (Sigma, St. Louis, MO) injection, and female mice were mated with adult male mice. A plug check was performed to verify the mating. The mice were sacrificed by cervical dislocation. The embryos were collected by puncturing the oviduct under a stereomicroscope and put in a 37°C incubator with and without additional CO₂ filters in culture media (G-TL, Vitrolife, USA).

Retrospective Analysis of Human Material

This study was approved by the Ethics Committee of the Biruni University Medical Faculty (No. 2020-12-9 and Date. 21.01.2020), and written informed consent was signed by each patient. Patients seeking infertility treatment at Anadolu Medical Center IVF Unit were included in the study. Inclusion criteria included being under 35 years old, having a diagnosis of tubal factor infertility, and generating more than 15 oocytes. After ICSI was performed, the two pronuclei (PN) embryos were divided into two groups: Group 1 was incubated with extra CO₂ filters, and Group 2 was incubated without extra CO₂ filters. Embryos were assessed on days 3 and 5 using Alpha Istanbul Consensus on Embryo assessment [12].

The Groups of Incubators and Embryo Culture

K-System G210 model incubator with BLUE; ORIGIO® Gas Line Filter was used. 2 additional in-line filters as CODA® XTRA INLINE® FILTERS – BLUE; ORIGIO® Gas Line Filter and REPROCARE GLF30 VOC Holder Filter were also attached to the incubators separately. During the experiments, all other conditions were kept the same so that the only variable remained the filtering system. The development stages of the embryos were monitored, and the blastomere size and regularity, granulation of cytoplasm, and fragmentation ratio were investigated during this assessment. Equal blastomere size and shape and absence of fragmentation were used as two parameters to define embryo quality.

Western Blot

Embryos from each group were collected in 5 mL PBS and 5 mL lysis buffer and incubated on ice for 30 minutes. Samples were mixed with 10 mL of Laemmli buffer, boiled for 5 minutes, cooled on ice, and then centrifuged at 2,000 rpm at 4°C for 5 minutes. Levels of Grp78 protein were determined using a rabbit monoclonal anti-BiP antibody (Cell Signaling Technology) in a standard Western blot protocol as described elsewhere [13].

Statistical Analysis

All experiments were repeated at least three times. Statistical data analysis used Student's t-test and one-way analysis of variance. p<0.05 was considered statistically significant.

Results

The Purity Levels of Gasses in the Incubators

When Nova 436 Series Gas Analyzer assessed the CO₂ gas content, the following impurities were isolated: ethyl methyl ketone, tetrahydrofuran, Fenol, 2-heptanol, and formaldehyde (concentrations at 0.000271%, 0.001316%, 0.001638%, 0.000012%, 0.001162%, respectively). As shown in Figure 1, it has been observed that the purity levels of substances indicated in the CO₂ gas were unsuitable when the extra filter was not used. The purity increased significantly after using the extra CO₂ filter.

Quality of Embryos Cultured in Filtered and Unfiltered Incubators

When the embryo quality was examined on the third day, a statistically significant increase was observed in Group 1 compared to Group 2 (47%, 48%, 51%, and 52%; for Group 1, CODA, ORIGIO, and REPROCARE, respectively; $p < 0,001$; $n = 30/\text{group}$) (Figure 2).

The rate of blastocyst utilization rate as a result of culture was examined in all three extra CO₂ filter groups, and a significant increase was observed compared to Group 1 (26%, 36%, 38%, and 42%; for Group 1, CODA, ORIGIO, and REPROCARE respectively; $p < 0,001$; $n = 30/\text{group}$) (Figure 3). When the embryo quality on the third and fifth day was compared between the extra-filtered groups, no statistically significant difference was found ($p > 0.05$) (Table 1).

Table 1: The comparisons of D3 and D5 embryos between Group 1 and Group 2. The quality of embryos on the third and fifth days was significantly higher in Group1 ($*p < 0.001$).

	GROUP 2	CODA	ORIGIO	REPROCARE	p
D3	47%	48%	51%	52%	<0.001
D5	26%	36%	38%	42%	<0.001

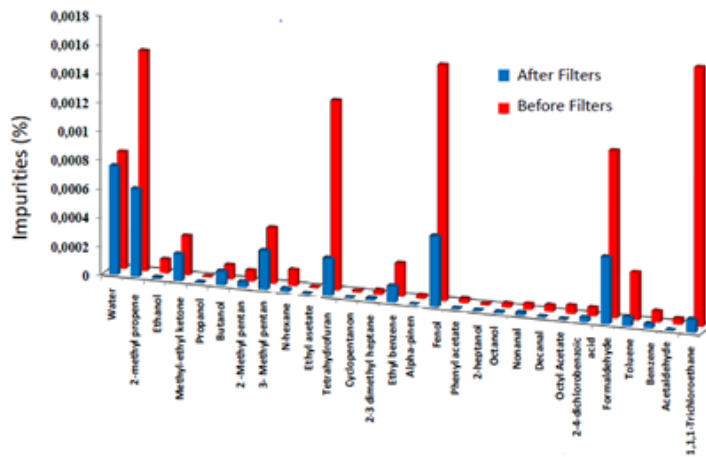


Figure 1: Percentage of impurities relative to the total amount of gas. It has been shown that the impurity percentages of the examined gasses are reduced when the extra filter is used in the incubator. (Red column: pre-filter; blue filter: post-filter).

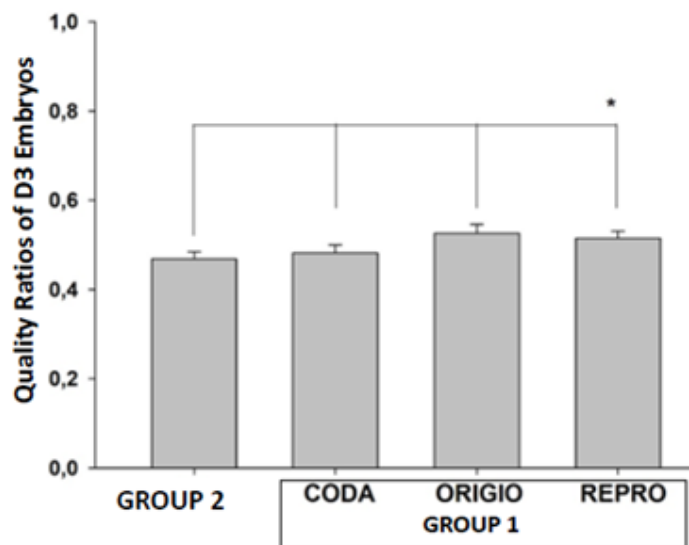


Figure 2: The ratios of Day 3 embryo qualities according to the groups with no extra filter (Group 2) and those with extra-filter (Group 1). It has been shown that the quality of embryos cultured in incubators with filters used for Group 1, CODA, ORIGIO and REPRO, is significantly better than Group 2 ($*p < 0.001$).

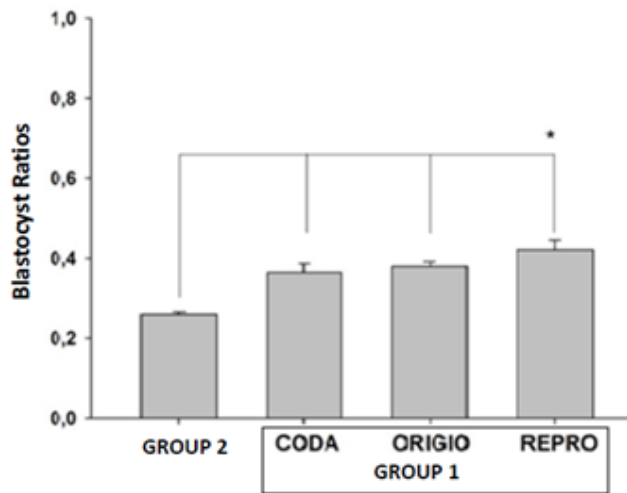


Figure 3: Blastocyst utilization rates between Group 1 and Group 2. It was shown that the quality of embryos cultured in incubators with filters used for Group 1, CODA, ORIGIO and REPRO was significantly better than Group 2 (*p<0.001).

Effect of Grp78 Protein Expression Level on Blastocyst Formation

To investigate whether the difference in embryo quality and blastocyst retrieval on the third day may depend on the UPR

signaling pathway, the GRP78 protein level was examined by Western Blot method. It was found to be higher in Group 2 (37%) compared to Group 1 (61%) (p<0.001) (Figure 4).

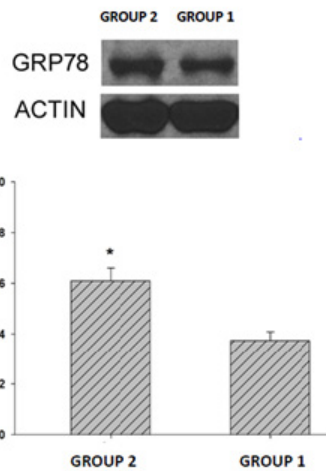


Figure 4: Comparison of Grp78 protein expression levels. It was shown that the Grp78 protein levels of mouse embryos cultured in incubators with filters were significantly lower than Group 2 (*p<0.001).

Discussion

IVF includes complex treatment processes tailored to the individual and can result in unique conditions and different outcomes. One of the most important factors known to reduce pregnancy is the air’s cleanliness in the laboratory environment. The presence of substances such as VOCs in the environment is extremely harmful to in vitro embryo development. Volatile organic compounds, which are constantly produced by laboratory materials and cleaning agents, react with indoor ozone to produce

submicron-sized particles and harmful by-products that directly affect pregnancy rates [14,15]. Air quality control within the laboratory is one of the most critical determinants of assisted reproductive success. It has been shown to increase parameters such as live birth rates significantly [16,17]. Various studies have shown that gametes and human embryos are sensitive to contaminants found in clinical and laboratory settings [7,17-19]. Many studies have investigated the effect of improved air quality on embryo quality [20,21].

In this study, the purity of CO₂ gas used in horizontal incubators with and without filters was investigated to evaluate whether this impacts embryo development. Heitmann et al. observed in their study that when they increased the air quality of the laboratory, the embryo implantation rate increased from 24.3% to 32%, and the live birth rate increased from 31.8% to 39.3% [22]. In another study by Bento and Esteves in 2013, pregnancy outcomes were compared after ICSI between 1999 and 2010 after the implementation of Brazilian national cleanroom standards [23]. They showed that the rate of spontaneous abortion decreased from 28.7% to 20% between the two study periods, while live birth rates increased from 25.8% to 35.6%. However, due to the wide time span of the study period, it suggests that other advances in laboratory and assisted reproductive techniques may also impact embryo development.

Contrary to other studies, in this study, since filtered and unfiltered comparisons were evaluated simultaneously, it is thought that more promising results were obtained to increase embryo quality. Khoudja et al. observed an increase in embryo quality and birth rate, as well as a decrease in spontaneous abortions, by using a new filter system for air purification in addition to the HEPA filters they used in the laboratory [5] and stated that there would be an increase in embryo quality with the reduction of volatile organic compounds. They also verified that the main reason for the change in embryo quality was the effect of the excess formaldehyde in the CO₂ gas. In this study, when we examined the CO₂ gas used in the laboratories, it was determined that the impurities of Methyl Ethyl ketone, Tetrahydrofuran, Phenol, 2-heptanol, and Formaldehyde substances were not suitable (Figure 1). When the extra filters were used, it was observed that these impurity rates were greatly reduced, but the amount of purity remaining after any extra filter was not sufficient due to the high amount of some species in the CO₂ gas (Figure 1).

Lin et al. investigated the effects of Endoplasmic Reticulum (ER) stress and Unfolded Protein Response (UPR) on mammalian oocyte maturation and preimplantation embryo development. ER stress generally plays a negative role in oocyte maturation and/or early embryo development [23,24]. They showed that understanding the mechanistic relationships between ER stress and in vitro development during in vitro human embryo production may help the development of assisted reproductive technology. In a cohort study conducted between 1993 and 1997, Boone et al. investigated the effect of volatile organic compounds on pregnancy rate and observed an increase in oocyte and embryo quality by reducing volatile organic compounds [20]. However, they concluded that there is still a need to investigate what the effect of air quality is directly related to. Swain made a comparative analysis of embryo culture incubators in study [25]. It has been shown that the choice of the incubator is one of the most critical decisions for the IVF laboratory as the devices regulate various environmental variables that can affect embryo development. In addition to functional

aspects of the incubator such as gas capacity and sensor type, temperature control and size/patient capacity should also be considered.

In this study, it has been shown that the gasses used in IVF laboratories are not as clean as thought, this impurity adversely affects embryo development, and this can be addressed and prevented by attaching an extra filter to the incubators used. In addition, it has been shown that UPR pathways play a role in the negative effect of ambient air/gas impurity on embryo development in long-term culture (5 or 6 days). As a result of this study, adding extra filters to the incubators used in IVF laboratories will reduce the stress on the embryos and provide better embryo development and, accordingly, higher pregnancy results.

References

1. World Health Organization (WHO) (2018) International Classification of Diseases, 11th Revision (ICD-11) Geneva: WHO 2018.
2. Tanoomand A, Hajibemani A, Abouhamzeh B (2019) Investigation of the association of idiopathic male infertility with polymorphisms in the methionine synthase (MTR) gene. *Clin Exp Reprod Med* 46(3): 107-111.
3. Tarahomi M, Melker A, Wely M, Hamer G, Repping S, et al. (2018) pH stability of human preimplantation embryo culture media: effects of culture and batches. *Reproductive biomedicine online* 37(4): 409-414.
4. Chansel Debordeaux L, Dagorne V, Mercier M, Soula V, Chauzit E (2018) Method validation of a critical parameter in human embryos culture: culture media pH. *Ann Biol Clin (Paris)* 76(3): 251-258.
5. Khoudja RY, Xu Y, Li T, Zhou C (2012) Better IVF outcomes following improvements in laboratory air quality. *Journal of assisted reproduction and genetics* 30(1): 69-76.
6. Ritz B, Wilhelm M (2008) Ambient air pollution and adverse birth outcomes: methodologic issues in an emerging. *Field Basic Clin Pharmacol Toxicol* 102(2): 182-190.
7. Morbeck DE (2015) Air quality in the assisted reproduction laboratory: a mini-review. *J Assist Reprod Genet* 32(7): 1019-1024.
8. Dickey RP, Potts A, Wortham JWE, Welch A (2010) Effect of IVF laboratory air quality on pregnancy success. *Fertility and Sterility* 94(4): 151.
9. Wang T, Babayev E, Jiang Z, Li G, Zhang M, et al. (2018) Mitochondrial unfolded protein response gene Clpp is required to maintain ovarian follicular reserve during aging, for oocyte competence, and development of pre-implantation embryos. *Aging Cell* 17(4): e12784.
10. Basar M, Bozkurt I, Guzeloglu-Kayisli O, Sozen B, Tekmen I, et al. (2014) Unfolded protein response prevents blastocyst formation during preimplantation embryo development in vitro. *Fertil Steril* 102(6): 1777-1784.
11. Wang M, Wey S, Zhang Y, Ye R, Lee AS (2009) Role of the unfolded protein response regulator GRP78/BiP in development, cancer, and neurological disorders. *Antioxid Redox Signal* 11(9): 2307-2316.
12. Alpha Scientists in Reproductive Medicine and ESHRE Special Interest Group of Embryology (2011) The Istanbul consensus workshop on embryo assessment: proceedings of an expert meeting. *Hum Reprod* 26(6): 1270-1283.
13. Seval Y, Cakmak H, Kayisli UA, Arici A (2006) Estrogen-mediated regulation of p38 mitogen-activated protein kinase in human endometrium. *J Clin Endocrinol Metab* 91(6): 2349-2357.
14. Cecchi M, Esposito B, Gilligan A, Garrisi GJ, Schimmel T (1997) Removal of volatile organic compounds from incubators used for gamete and embryo culture. *Fertility and Sterility* 68(1): 165.

15. Munch EM, Sparks AE, Duran HE, Van Voorhis BJ (2015) Lack of carbon air filtration impacts early embryo development. *Journal of Assisted Reproduction and Genetics*. 32(7): 1009-1017.
16. Esteves SC, Bento FC (2016) Air quality control in the ART laboratory is a major determinant of IVF success. *Asian Journal of Andrology* 18(4): 596-599.
17. Esteves SC, Bento FC (2016) Implementation of cleanroom technology in reproductive laboratories: the question is not why but how. *Reproductive Biomedicine Online*. 32(1): 9-11.
18. Palter S, DiPaola K, Sparks AE, Degelos S, Koulianos GT (2016) Multi-center study: innovative control of ambient air quality in multiple IVF laboratories is associated with statistically significant improvements in clinical outcomes - analysis of 5319 cycles. *Fertil Steril* 106: e27-e28.
19. Munch EM, Sparks AE, Van Voorhis BJ, Duran HE (2014) Poor laboratory air quality and its impact on early embryo development. *Fertil Steril* 101(2): e14.
20. Boone WR, Crane MM, Johnson JE, Locke AJ, Price TM (1999) Control of air quality in an assisted reproductive technology laboratory. *Fertility and Steril* 71(1): 150-154.
21. Johnson JE, Boone WR, Bernard RS (2016) The effects of Volatile Compounds (V.C.) on the outcome of in vitro mouse embryo culture. *Fertil Steril (Suppl 1)*: S98-S99.
22. Heitmann RJ, Hill MJ, James AN, Schimmel T, Segars JH (2015) Live births achieved via IVF are increased by improvements in air quality and laboratory environment. *Reproductive Biomedicine Online* 31(3): 364-371.
23. Esteves SC, Bento FC (2013) Implementation of air quality control in reproductive laboratories in full compliance with the Brazilian Cells and Germinative Tissue Directive. *Reproductive Biomedicine Online* 26(1): 9-21.
24. Lin T, Lee JE, Kang JW, Shin HY, Lee JB (2019) Endoplasmic Reticulum (E.R.) Stress and Unfolded Protein Response (UPR) in Mammalian Oocyte Maturation and Preimplantation Embryo Development. *International Journal of Molecular Sciences*. 20(2): 409.
25. Swain JE (2014) Decisions for the IVF laboratory: comparative analysis of embryo culture incubators. *Reproductive biomedicine online* 28(5): 535-547.