



Review Article

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Mechanisms of VSMC Proliferation, Migration, and Phenotypic Transformation in Homocysteine-Induced Atherosclerosis: A Review

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Abstract

Evidence-based medical evidence has shown that homocysteine (Hcy) is an independent risk factor for atherosclerosis (As). If the pathogenesis of Hcy induction As can be elucidated, it will provide an important theoretical basis for the prevention and clinical treatment of As. Vascular smooth muscle cells (VSMC) are the main components of the vascular media, and the proliferation, migration, and phenotypic transformation of VSMC play an important role in the pathogenesis of Hcy induced As. This paper reviews the mechanisms related to the proliferation, migration, and phenotypic transformation of VSMC during Hcy induction of As.

Keywords: Homocysteine; Atherosclerosis; Vascular smooth muscle cells; Phenotypic transformation

Introduction

Atherosclerosis (As) is an important basis lesion in many cardiovascular and cerebrovascular diseases such as ischemic cardiomyopathy and cerebral stroke. Homocysteine (Hcy) is an independent risk factor for atherosclerosis (As). Hyperhomocysteinemia (HHcy) can induce As, but the pathogenesis is unclear. Vascular smooth muscle cells (VSMC) are the main components of the vascular media and the main source of foam cells in As plaques. Hcy can induce the proliferation, migration, and phenotypic transformation of VSMC, which plays an important role in the pathogenesis of As induced by Hcy. This paper reviews the mechanisms related to the proliferation, migration and phenotypic transformation of VSMC during Hcy induction of As.

The Main Pathogenesis of Atherosclerosis

Atherosclerosis (As) is a chronic compensatory arterial inflammatory response associated with abnormal lipid metabolism and changes in the composition of blood vessel walls, which is the

main cause of cardiovascular disease (CVD). It is considered to be the common pathological basis of most cardiovascular diseases such as myocardial infarction, stroke, and peripheral artery disease [1]. The formation of As lesions is a chronic inflammatory process involving complex signal networks and a variety of effector molecules [2-4]. Currently, the main theories on the pathogenesis of As include [5]: inflammatory response theory [6], oxidative stress theory [7], lipid infiltration theory [8,9], homocysteine theory, thrombus formation theory [10], and immune injury theory. It is believed that the formation of As is mainly caused by circulating factors, cholesterol, low density lipoprotein, inflammatory factors, chemokines, and a variety of cells in the vascular wall. Including the results of interactions among vascular endothelial cells, lymphocytes, monocytes/macrophages, and vascular smooth muscle cells (VSMC) [11], which are specifically manifested as damage of vascular intima, activation of chemokines and inflammatory factors, lipid infiltration, and dysfunction

of endothelial cells [12]. VSMC proliferation and phenotypic transformation [13], macrophage-like cell formation, migration, foam cell formation [14], and atheromatous plaque formation are main pathologic processes in As formation. VSMC is one of the active cells in the As plaque proliferation system [15,16], and the proliferation, migration and phenotypic transformation of VSMC and the injury of vascular endothelial cells are considered to be the main initial links in the formation of As [17,18]. Although many studies have been conducted on As, the mechanisms that cause proliferation, migration and phenotypic transformation of VSMC remain unclear.

Proliferation, Migration, and Phenotypic Transformation of VSMC Play an Important Role in the Pathogenesis of As

VSMC is a highly specific cell located in the middle layer of the arterial wall and is the main source of macrophage-like and foam cells in As plaques [8,19]. The main function of VSMC is to regulate vascular elasticity and maintain vascular tone. During embryonic vascular development, VSMC gradually differentiated from immature undifferentiated/poorly differentiated phenotype to mature highly differentiated phenotype. When the vascular wall is stimulated by internal and external environmental factors or the VSMC is stimulated by inflammatory factors, cytokines, vasoactive peptides, oxidative stress, drug damage, mechanical effects, and other pathological factors, on the one hand, the imbalance of VSMC proliferation and apoptosis can be caused, resulting in VSMC proliferation and migration. At the same time, VSMC can transform from a highly mature differentiated phenotype to a dedifferentiated/poorly differentiated phenotype, and obtain proliferation and phagocytosis capabilities, and eventually form macrophage-like cells. This pathological process is called the phenotypic transformation of VSMC [20]. The proliferation, migration, and cell phenotypic transformation of VSMC may be key events in the pathological process of various vascular diseases such as As, and play an important role in the occurrence and development of As [1,5,13]. According to the maturity of VSMC differentiation, VSMC can be divided into two phenotypes: highly differentiated systolic phenotype (differentiated type) and poorly differentiated synthetic phenotype (dedifferentiated or undifferentiated type) [21,22]. The systolic phenotype of VSMC is highly differentiated, spindle-shaped, rich in intracellular muscle fibers, and mainly manifested as systolic function, poor proliferation and migration ability. The main molecular markers were α -smooth muscle actin (α -SMA) [23], smooth muscle 22 α (Smooth Muscle 22 α , SM22 α) [24,25], calponin, Smooth muscle myosin heavy chain (SM-MHC), smooth muscle cell actin [13,26-28]. However, the synthetic phenotype of VSMC has a low differentiation degree, strong proliferation and migration ability, reduced intracellular muscle fibers, reduced or disappeared systolic function, and increased contents of intracellular rough endoplasmic reticulum, Golgi apparatus and ribosome, and stronger ability to secrete and synthesize cytokines and extracellular matrix than the systolic phenotype. In addition,

VSMCS undergoing phenotypic transformation can also acquire macrophage-like markers and properties, including LGALS3/Mac2, CD11b, F4/80, and CD68 [13], and have phagocytic function. The ability of synthetic phenotypic VSMCS to absorb lipids was also increased, The unique molecular markers of osteopontin (OPN) and thrombospondin-1 were Osteopontin (OPN) and Thrombospondin-1. TSP1), epidermal growth factor; epiregulin, etc. [29-31]. The phenotypic transformation of VSMC from systolic phenotype to synthetic phenotype has long been considered important for As. The phenotypic transformation of VSMC during the occurrence of As is not only the main feature of VSMC in As, but also the premise for VSMC to form macrophage-like cells and myogenic foam cells and play an important role in plaque formation and development [13,32]. Under normal conditions, VSMC exists in the media of arterial wall with a contractile phenotype, which is conducive to maintaining the contraction of smooth muscle cells and vascular tone to maintain vascular wall homeostasis. However, when VSMC is subjected to different pathologic and physiological stimuli, changes occur in the surrounding environment of the cells, including growth factors, extracellular matrix, mechanical forces, oxidative stress, intercellular interactions, and neuroregulation, resulting in phenotypic changes of VSMC, from a highly differentiated contractile phenotype to an undifferentiated or poorly differentiated synthetic phenotype. They began to proliferate, migrate and synthesize excessive extracellular matrix to form macrophage-like cells, resulting in increased lipid phagocytosis of VSMC, which could take in large amounts of oxidized low-density lipoprotein, and migrate from the arterial media to the inner artery to transform into foam cells [13,15,19]. Foam cells are deposited under the intima of the damaged blood vessels, causing local vascular lumen stenosis [33,34] and changes in hemodynamics, eventually leading to the formation of atherosclerotic plaques. Literature has shown that, in plaque formation of As, 70% of plaque components are composed of VSMC and its derivatives [35,36], and 40% of foam cells, which constitute an important part of lesions, are from VSMC, which is called smooth muscle-derived foam cells [37,38]. It is suggested that VSMC is an important component in the formation of As. At the same time, the transformation from systolic phenotype to synthetic phenotype of VSMC can promote endometrial hyperplasia and the formation of As lesions and abnormal phenotype transformation of VSMC is therefore considered to be one of the main markers of the progression of As lesions [39], As well as an important step in the development of AS restenosis, vascular remodelling and other pathophysiological processes [40,41]. It is certain that the transformation of VSMC into foam cells not only damages the vasoconstrictive function, but also promotes its own proliferation, migration and secretion of pro-inflammatory mediators [42], and the expression of different phenotypic markers is regulated by various factors. Such as platelet-derived growth factor (PDGF), transforming growth factor β , TGF- β), interleukin, endothelin, Angiotensin and microRNAs, etc. [13,15]. In carotid artery ligation animal models, blocking the phenotypic transformation of VSMC can inhibit the formation of

intima damage [43,44], and promoting this process can accelerate the development of As in mice [45]. Meanwhile, the phenotypic transformation of VSMC also plays an important role in the stability of As and plaque, and inhibiting the phenotypic transformation of VSMC may be beneficial for advanced As [15]. Although there have been some reports on the proliferation, migration and phenotypic transformation of VSMC, the specific molecular mechanism has not been clarified and needs further study.

Hcy Can Induce the Occurrence of As

Homocysteine (Hcy), also known as homocysteine, is a kind of sulfur-containing amino acid, which is the intermediate product of methionine metabolism. Normally, plasma Hcy levels are very low; When the plasma Hcy concentration is higher than 15 $\mu\text{mol/L}$, it is called hyperhomocysteinemia (HHcy). Evidence of evidence-based medicine shows that HHcy is an independent risk factor for As [46,47], and Hcy induces As through a variety of pathway interactions and correlations. Every 5 $\mu\text{mol/L}$ increase in plasma Hcy is equivalent to a 0.5 mmol/L increase in cholesterol, and the vascular risk increases by about 1/3 [48]. Hcy is closely associated with the risk of coronary heart disease, stroke, and peripheral vascular diseases, and is no less harmful than hyperlipidemia. Known as the "cholesterol of the 21st century", HCY has been identified as a potentially important risk factor for cardiovascular diseases [49-51]. The Chinese Guidelines for the Prevention and Treatment of Hypertension (2018 Revision) clearly proposed serum Hcy concentration as a selective item for laboratory tests in the diagnostic assessment of hypertension. HHcy ($\geq 15 \mu\text{mol/L}$), together with old age, smoking, impaired glucose tolerance, dyslipidemia, abdominal obesity and other factors, are considered to be important cardiovascular risk factors affecting the cardiovascular prognosis of patients with hypertension [52], indicating that reducing blood Hcy is an important strategy for collaborative prevention and treatment of cardiovascular diseases. As an inflammatory stimulant, Hcy promotes the occurrence and development of AS through various mechanisms such as affecting the function of endothelial cells and VSMC, participating in oxidative stress and inflammatory response, and altering gene expression activity [53,54].

Hcy Promotes the Proliferation, Migration, and Phenotypic Transformation of VSMC

At present, the mechanism of Hcy causing As through vascular endothelial cell injury has been relatively clear [55,56]. It includes the following aspects: (1) direct endothelial injury through oxidation and inflammation [57,58]; (2) Induced oxidative stress mitochondrial dysfunction and endoplasmic reticulum stress [59-61]; (3) the protective effect of NO was weakened [62]; (4) Interference with DNA and protein methylation [63,64]; (5) Promote adhesion and penetration [65]. The promotion effect of Hcy on the proliferation of VSMC is considered to be one of the important pathological basis of As [66], however, the mechanism

of Hcy causing the proliferation, migration and expression transformation of VSMC is not very clear.

Studies have shown that, on the one hand, Hcy can act on related growth factors and genes, activate VSMC, and promote the proliferation and migration of VSMC. PDGF is an important mitogenic factor, which can stimulate specific cell mitosis and promote cell proliferation. Hcy can up-regulate PDGF levels through DNA demethylation of human and mice vascular endothelial cells, affect the cross-linking between vascular endothelial cells and VSMC, and lead to activation of VSMC [46]. Hcy affects the methylation of As-related genes and mediates the overall methylation state of VSMC proliferation [67,68]. Hcy can be hypermethylated through promoter regions of p53, PTEN, MFN2, etc. Demethylation of PDGF promoter region affects epigenetic regulation of p53, PTEN, MFN2, PDGF, etc., thus promoting proliferation of VSMC [69-71]. Hcy can cause dysregulation of the expression and activity of metalloproteinase 2/9 (MMP-2/9) and tissue inhibitor of metalloproteinase 2 (TIMP-2) in VSMC, affect the dynamic balance of extracellular matrix, degrade extracellular matrix such as basement membrane, destroy the physiological barrier of VSMC migration, and induce the proliferation and migration of VSMC in rats [72]. Hcy (50-1000 $\mu\text{mol/L}$) increased the production and activation of MMP-2 and the expression of TIMP-2 in rat VSMC in a dose-dependent manner, while the expression of MMP-2 was up-regulated and the activity was down-regulated when incubated with 5000 $\mu\text{mol/L}$ Hcy [73]. It is possible that Hcy promotes the expression of the protein Concave protein-1, which inhibits the activity of endothelial nitric oxide synthase (eNOS) and the production of NO and activates the expression of PI3K and p-Akt. The proliferation and migration of thoracic aorta smooth muscle cells cultured in vitro in SD rats were induced, leading to As [74,75]. Meanwhile, compared with the control group, the proliferation and migration ability of VSMC in aorta of rats in the Hcy stimulation group were significantly enhanced, the mRNA and protein expression levels of MMP-2, MMP-9, and p-P70S6K were significantly increased, and the expression levels of p21 and p27 were significantly decreased [76]. These results suggest that Hcy can induce the proliferation and migration of VSMC, and play an important role in the pathogenesis of As induced by Hcy.

On the other hand, Hcy affects collagen synthesis and metabolism in VSMC. In Cbs^{-/-} mice aortic VSMC, the effect of Hcy on collagen secretion was observed. It was found that Cbs^{-/-} mice with severe HHcy had significantly thickened vascular intima, a higher percentage of lumen stenosis, and significantly increased the deposition of elastin and collagen in the newborn intima and the secretion of collagen in VSMC. These results suggest that Hcy stimulates increased neointima formation, elastin and collagen deposition, contributing to the development of vascular remodelling [77].

Thirdly, Hcy can promote lipid deposition in blood vessel walls. Hcy can induce lipid deposition in the arterial wall, an

increase of foam cells, plaque calcification, and low-density lipoprotein oxidation. Hcy can promote oxidative stress reaction in VSMC and generate reactive oxygen species (ROS). Through oxidative modification of low-density lipoprotein, oxidized low-density lipoprotein (ox-LDL) increases in oxidized low-density lipoprotein (Oxidized low-density lipoprotein), thereby promoting foam cell generation [49]. At the same time, ox-LDL itself is highly cytotoxic to vascular endothelial cells, macrophages and VSMC. ox-LDL can directly damage vascular endothelial cells, activate monocytes in blood to swim to the wall of the bleeding tube and transform into macrophages, and activate VSMC to phagocytic ox-LDL, and eventually form foam cells. Aggravate the occurrence and development of As [37-39,78]. Our research group also found that Hcy induced proliferation, migration and phenotypic transformation of VSMC by activating PI3K/Akt and mTOR signalling pathways. miRNA-145 can inhibit the activity of PI3K/Akt and mTOR pathways, and reduce the proliferation, migration and phenotypic transformation of VSMC induced by Hcy [79]. Lycium barbarum polysaccharide inhibits the PI3K/Akt signalling pathway by up-regulating the expression of miRNA-145 and alleviates the proliferation and phenotypic transformation of Hcy-induced VSMC [80]. By inhibiting the expression of CTRP9, Hcy negatively regulates the occurrence of endoplasmic reticulum stress, thereby inducing VSMC migration. Meanwhile, the up-regulation of DNMT1 plays an important role in this process, suggesting that CTRP9 may be regulated by methylation [81].

Conclusion

These studies suggest that Hcy can affect the extracellular matrix, destroy the vascular basement membrane, and activate growth factors and related genes in VSMC, ultimately promoting the proliferation and migration of VSMC. Hcy induced proliferation and migration of VSMC is one of the important mechanisms of Hcy induced As [82]. VSMC over proliferate and migrate through basement membrane to endovascular subcutaneous, transforming into foam cells by phagocytosis of lipids, and eventually developing into fibrous plaques [83]. These results indicate that Hcy is closely related to the damage of As VSMC. Hcy can induce the proliferation, migration and phenotypic transformation of VSMC, and ultimately promote the formation of foam cells in As plaques. However, the above studies also have some shortcomings, such as the relatively independent and lack of internal correlation between the mechanisms, and it is not clear whether there is interaction between them. Furthermore, whether there are other unknown molecular mechanisms in the process of Hcy promoting the proliferation, migration and phenotypic transformation of VSMC, so As to promote the occurrence and development of As also needs further study.

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Author Contributions

ZMH designed the idea of the article. WXY, MT, MX, ZY and XLB collected relevant literature and analyzed it. ZMH and WXY wrote the manuscript. All authors have read and approved the manuscript.

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Conflict of Interest

All authors declare that they have no conflict of interest.

Consent to Participate

Not applicable.

Consent to Publish

Not applicable.

References

1. Björkbacka H (2021) Atherosclerosis: cell biology and lipoproteins. *Curr Opin Lipidol* 32(1): 74-75.
2. Tan L, Xu Q, Shi RZ (2021) Bioinformatics analysis reveals the landscape of immune cell infiltration and immune-related pathways participating in the progression of carotid atherosclerotic plaques. *Artif Cells Nanomed Biotechnol* 49(1): 96-107.
3. Takahashi M (2021) NLRP3 inflammasome as a key driver of vascular disease. *Cardiovasc Res* 118(2): 372-385.
4. Wolf D, Ley K (2019) Immunity and inflammation in atherosclerosis. *Circ Res* 124(2): 315-327.
5. Weber C, Noels H (2011) Atherosclerosis: current pathogenesis and therapeutic options. *Nat Med* 17(11): 1410-1422.
6. Taleb S (2016) Inflammation in atherosclerosis. *Arch Cardiovasc Dis* 109(12): 708-715.
7. Shao BZ, Han BZ, Zeng YX, Su DF, Liu C, et al. (2016) The roles of macrophage autophagy in atherosclerosis. *Acta Pharmacol Sin* 37(2): 150-156.
8. Chistiakov DA, Melnichenko AA, Myasoedova VA, Grechko AV, Orekhov AN, et al. (2017) Mechanisms of foam cell formation in atherosclerosis. *J Mol Med (Berl)* 95(11): 1153-1165.
9. Wang DD, Yang Y, Lei YN, Tzvetkov NT, Liu X, et al. (2019) Targeting foam cell formation in atherosclerosis: therapeutic potential of natural products. *Pharmacol Rev* 71(4): 596-670.
10. Maguire EM, Pearce SWA, Xiao Q (2019) Foam cell formation: a new target for fighting atherosclerosis and cardiovascular disease. *Vascul Pharmacol* 112: 54-71.
11. Schaftenaar F, Frodermann V, Kuiper J (2016) Atherosclerosis: the interplay between lipids and immune cells. *Curr Opin Lipidol* 27(3): 209-215.

12. Sitia S, Tomasoni L, Atzeni F, Ambrosio G, Cordiano C, et al. (2010) From endothelial dysfunction to atherosclerosis. *Autoimmun Rev* 9(12): 830-834.
13. Bennett MR, Sinha S, Owens GK (2016) Vascular smooth muscle cells in atherosclerosis. *Circ Res* 118(4): 692-702.
14. Doodnauth SA, Grinstein S, Maxson ME (2019) Constitutive and stimulated macropinocytosis in macrophages: roles in immunity and in the pathogenesis of atherosclerosis. *Philos Trans R Soc Lond B Biol Sci* 374(1765): 20180147-20180158.
15. Basatemur GL, Jørgensen HF, Clarke MCH (2019) Vascular smooth muscle cells in atherosclerosis. *Nat Rev Cardiol* 16(12): 727-744.
16. Zhu JM, Liu B, Wang ZY, Wang D, Ni H, et al. (2019) Exosomes from nicotine-stimulated macrophages accelerate atherosclerosis through miR-21-3p/PTEN-mediated VSMC migration and proliferation. *Theranostics* 9(23): 6901-6919.
17. Wolf MP, Hunziker P (2020) Atherosclerosis: Insights into vascular pathobiology and outlook to novel treatments. *J Cardiovasc Transl Res* 13(5): 744-757.
18. Qi YX, Han Y, Jiang ZL (2018) Mechanobiology and vascular remodeling: from membrane to nucleus. *Adv Exp Med Biol* 1097: 69-82.
19. Yu XH, Fu YC, Zhang DW, Yin K, Tang CK, et al. (2013) Foam cells in atherosclerosis. *Clin Chim Acta* 23: 245-252.
20. Liu K, Fang CC, Shen YW, Liu Z, Zhang M, et al. (2017) Hypoxia-inducible factor 1a induces phenotype switch of human aortic vascular smooth muscle cell through PI3K/AKT/AEG-1 signalling. *Oncotarget* 8(20): 33343-33352.
21. Li Q, Wen JK, Zheng B (2003) Recent advances in phenotypic regulation of vascular smooth muscle cells. *Progress in Physiological Sciences* 34(1): 27-31.
22. Lacolley P, Regnault V, Nicoletti A, Li Z, Michel JB, et al. (2012) The vascular smooth muscle cell in arterial pathology: a cell that can take on multiple roles. *Cardiovasc Res* 95(2): 194-204.
23. Shen F, Yang P, Tao XJ et al. (2019) Regulation of SM22 α expression by 27nt-miRNA in vascular smooth muscle cells and its effects on cell viability, migration and phenotypic changes. *Chinese Journal of Pathophysiology*, 2019, 35(2): 200-205.
24. Zhao LL, Zhang F, Chen P, Xie XL, Dou YQ, et al. (2017) Insulin-independent GLUT4 translocation in proliferative vascular smooth muscle cells involves SM22 α . *J Mol Med* 95(2): 181-192.
25. Feil S, Hofmann F, Feil R (2004) SM22 α modulates vascular smooth muscle cell phenotype during atherogenesis. *Circ Res* 94(7): 863-865.
26. Ha JM, Yun SJ, Kim YW, Jin SY, Lee HS, et al. (2015) Platelet-derived growth factor regulates vascular smooth muscle phenotype via mammalian target of rapamycin complex1. *Biochem Biophys Res Commun* 464(1): 57-62.
27. Gomez D, Owens GK (2012) Smooth muscle cell phenotypic switching in atherosclerosis. *Cardiovasc Res* 95(2): 156-164.
28. Patel JJ, Bolrne LE, Davies BK, Arnett TR, MacRae VE, et al. (2019) Differing calcification processes in cultured smooth muscle cells and osteoblasts. *Exp Cell Res* 380(1): 100-113.
29. Monzl RM, Saleii A, Jlliana M (2014) Mechanisms of vascular calcification and associated diseases. *Curr Pharm Des* 20(37): 5801-5810.
30. Chang SY, Song SJ, Lee JS, Yoon J, Park J, et al. (2014) Phenotypic modulation of primary vascular smooth muscle cells by short-term culture on micropatterned substrate. *PLoS One* 9(2): e88089- e88090.
31. Ehrenborg E, Paloschi V, Goncalves I (2020) Repression of MAP1LC3A during atherosclerosis progression plays an important role in the regulation of vascular smooth muscle cell phenotype. *Atherosclerosis* 315(1): e22-e33.
32. Su JY, Tang CS, H QD (1994) Pathophysiological basis and pathogenesis of cardiovascular diseases. First edition: 40-41.
33. Wang D, Tan ZZ, Fu SL (2021) Research progress of circrnas in atherosclerosis. *Chinese Journal of Pathophysiology* 29(1): 1-9.
34. Tan ZZ, Chen JB, Wang D (2021) Research progress of microns in foam cell formation. *Chinese Journal of Pathophysiology* 29(1): 30-36.
35. Shankman LS, Gomez D, Cherepanova OA (2015) KLF4-dependent phenotypic modulation of smooth muscle cells has a key role in atherosclerotic plaque pathogenesis. *Nat Med* 21(6): 628-637.
36. Allahverdian S, Chehroudi AC, McManus BM, Abraham T, Francis GA, et al. (2014) Contribution of intimal smooth muscle cells to cholesterol accumulation and macrophage-like cells in human atherosclerosis. *Circulation* 129(15): 1551-1559.
37. Susanne F, Birgit F, Robert L, Essmann F, Schulze-Osthoff K, et al. (2014) Trans differentiation of vascular smooth muscle cells to macrophage-like cells during atherogenesis. *Circ Res* 115(7): 662-667.
38. Chaabane C, Coen M, Bochaton-Piallat ML (2014) Smooth muscle cell phenotypic switch: implications for foam cell formation. *Curr Opin Lipidol* 25(5): 374-379.
39. Dubland JA, Francis GA (2016) So much cholesterol: the unrecognized importance of smooth muscle cells in atherosclerotic foam cell formation. *Curr Opin Lipidol* 27(2): 155- 161.
40. Fisher EA, Miano JM (2014) Don't judge books by their covers: vascular smooth muscle cells in arterial pathologies. *Circulation* 129(15): 1545-1547.
41. Frisantiene A, Philippova M, Erne P (2018) Smooth muscle cell-driven vascular diseases and molecular mechanisms of VSMC plasticity. *Cell Signal* 52: 48-64.
42. Zhang MJ, Zhou Y, Chen L, Wang YQ, Wang X, et al. (2016) An overview of potential molecular mechanisms involved in VSMCs phenotypic modulation. *Histochem Cell Biol* 145(2): 119-130.
43. Lu QB, Wan MY, Wang PY, Zhang CX, Xu DY, et al. (2018) Chicoric acid prevents PDGF-BB-induced VSMCs de- differentiation, proliferation and migration by suppressing ROS/NFkappaB/mTOR/P70S6K signalling cascade. *Redox Biol* 14: 656-668.
44. Li P, Zhu N, Wang ND, Chen M, You X, et al. (2013) MicroRNA-663 regulates human vascular smooth muscle cell phenotypic switch and vascular neointimal formation. *Circ Res* 113(10): 1117-1127.
45. Zhang M, Liu LM, Zhi F, Niu P, Yang M, et al. (2016) Inactivation of semi carbazide-sensitive amine oxidase induces the phenotypic switch of smooth muscle cells and aggravates the development of atherosclerotic lesions. *Atherosclerosis* 249: 76-82.
46. Zhang DH, Chen YQ, Xie XN, Liu J, Wang Q, et al. (2012) Homocysteine activates vascular smooth muscle cells by DNA demethylation of platelet-derived growth factor in endothelial cells. *J Mol Cell Cardiol* 53(4): 487-496.
47. Xiao YJ, Su XF, Huang W, Zhang J, Peng C, et al. (2015) Role of S-adenosylhomocysteine in cardiovascular disease and its potential epigenetic mechanism. *Int J Biochem Cell Biol* 67: 158-166.
48. Held C, Sumner G, Sheridan P, McQueen M, Smith S, et al. (2008) Correlations between plasma homocysteine and folate concentrations and carotid atherosclerosis in high-risk individuals: baseline data from the Homocysteine and Atherosclerosis Reduction Trial (HART). *Vasc Med* 13(4): 245-253.
49. Nehler MR, Taylor Jr LM, Porter JM (1997) Homocysteinemia as a risk factor for atherosclerosis: a review. *Cardiovasc Surg* 5(6): 559-567.

50. Tribouilloy CM, Peltier M, Iannetta Peltier MC (2000) Plasma homocysteine and severity of thoracic aortic atherosclerosis. *Chest* 118(6): 1685-1689.
51. Sen S, Reddy PL, Grewal RP, Busby M, Chang P, et al. (2010) Hyperhomocysteinemia is associated with aortic atheroma progression in stroke/TIA patients. *Front Neurol* 26(1): 131-138.
52. Revision Committee of Chinese Guidelines on Hypertension Prevention and Treatment. Guidelines for Hypertension Prevention and Treatment in China (2018 Revision). Prevention and treatment of cardiovascular and cerebrovascular diseases, 2019, 19(1): 1-44.
53. Horvath B, Szapary L, Debrenci L, Feher G, Kenyeres P, et al. (2009) Effect of sclerovit on endothelial dysfunction, hemorheological parameters, platelet aggregation, plasma concentration of homocysteine and progression of atherosclerosis in patients with vascular diseases. *Clin Hemorheol Microcirc* 42(1): 19-28.
54. Potter K, Lenzo N, Eikelboom JW, Arnolda LF, Beer C, et al. (2009) Effect of long-term homocysteine reduction with B vitamins on arterial wall inflammation assessed by fluorodeoxyglucose positron emission tomography: a randomised double-blind, placebocontrolled trial. *Cerebrovasc Dis* 27(3): 259-265.
55. Fu Y, Wang X, Kong W (2018) Hyperhomocysteinemia and vascular injury: advances in mechanisms and drug targets. *Br J Pharmacol* 175(8): 1173-1189.
56. Esse R, Barroso M, Tavares de Almeida I (2019) The contribution of homocysteine metabolism disruption to endothelial dysfunction: state-of-the-art. *Int J Mol Sci* 20(4): 867-891.
57. Hu HM, Wang CY, Jin Y, Meng Q, Liu Q, et al. (2019) Catalpol inhibits homocysteine-induced oxidation and inflammation via inhibiting Nox4/NF- κ B and GRP78/PERK pathways in human aorta endothelial cells. *Inflammation* 42(1): 64-80.
58. Leng YP, Ma YS, Li XG, et al. I-homocysteine-induced cathepsin V mediates the vascular endothelial inflammation in hyperhomocysteinemia. *Br J Pharmacol*. 2018, 175(8): 1157- 1172.
59. Zhang ZM, Wei CY, Zhou YF, et al. Homocysteine induces apoptosis of human umbilical vein endothelial cells via mitochondrial dysfunction and endoplasmic reticulum stress. *Oxid Med Cell Longev*, 2017, 2017: 5736506-5736519.
60. Zhu L, Jia F, Wei J, Yu Y, Yu T, et al. (2017) Salidroside protects against homocysteine-induced injury in human umbilical vein endothelial cells via the regulation of endoplasmic reticulum stress. *Cardiovasc Ther* 35(1): 33-39.
61. Kaplan P, Tatarkova Z, Sivonova MK, Racay P, Lehotsky J (2020) Homocysteine and mitochondria in cardiovascular and cerebrovascular systems. *Int J Mol Sci* 21(20): 7698-7717.
62. Chen J, Huang Y, Hu X, Bian X, Nian S (2021) Gastrodin prevents homocysteine-induced human umbilical vein endothelial cells injury via PI3K/Akt/eNOS and Nrf2/ARE pathway. *J Cell Mol Med* 25(1): 345-357.
63. Che SS, Xu H, Guo W (2019) Role of DNA methylation of silk/threonine protein kinase in homocysteine-induced apoptosis of vascular endothelial cells. *Journal of practical medicine* 35(5): 709-714.
64. Zhang HP, Wang YH, Ma SH, Zhang H, Yang AN, et al. (2018) Homocysteine inhibits endothelial progenitor cells proliferation via DNMT1-mediated hypomethylation of Cyclin A. *Exp Cell Res* 362(1): 217-226.
65. Chantal S, Md RA, Katherine CS, Schoots IG, Brkovic A, et al. (2008) Priming effect of homocysteine on inducible vascular cell adhesion molecule-1 expression in endothelial cells. *Biomed Pharmacother* 62(6): 395-400.
66. Liu XH, Shen J, Zhan R, Wang X, Wang X, et al. (2009) Proteomic analysis of homocysteine induced proliferation of cultured neonatal rat vascular smooth muscle cells. *Biochim Biophys Acta* 1794(2): 177-184.
67. Zhang Y, Mei J, Li J, Zhang Y, Zhou Q, et al. (2021) DNA methylation in atherosclerosis: a new perspective. *Evid Based Complement Alternat Med* 23: 6623657.
68. Ma SC, Cao CJ, Zhang HP, Jiao Y, Zhang H, et al. (2017) Aberrant promoter methylation of multiple genes in VSMC proliferation induced by Hcy. *Mol Med Rep* 16(5): 7775-7783.
69. Han XB, Zhang HP, Cao CJ (2014) Aberrant DNA methylation of the PDGF gene in homo-cysteine-mediated VSMC proliferation and its underlying mechanism. *Mol Med Rep* 10(2): 947-954.
70. Xu L, Hao HY, Hao YJ (2019) Aberrant MFN2 transcription facilitates homocysteine-induced VSMCs proliferation via the increased binding of c-Myc to DNMT1 in atherosclerosis. *J Cell Mol Med* 23(7): 4611-4626.
71. Ma SC, Zhang HP, Jiao Y, Wang YH, Zhang H, et al. (2018) Homocysteine-induced proliferation of vascular smooth muscle cells occurs via PTEN hypermethylation and is mitigated by resveratrol. *Mol Med Rep* 17(4): 5312-5319.
72. Meng LP, Liu LB, Zhou CZ, Pan S, Zhai X, et al. (2016) Polyphenols and polypeptides in chinese rice wine inhibit homocysteine-induced proliferation and migration of vascular smooth muscle cells. *J Cardiovasc Pharmacol* 67(6): 482-490.
73. Shi YF, Chi JF, Tang WL, Xu FK, Liu LB, et al. (2013) Effects of rosuvastatin on the production and activation of matrix metalloproteinase-2 and migration of cultured rat vascular smooth muscle cells induced by homocysteine. *J Zhejiang Univ Sci B(English)* 14(8): 696-704.
74. Bao XM, Deng HC (2015) Effect of homocysteine on proliferation and migration of vascular smooth muscle cells in rats and its possible mechanism. *Shandong Medical Journal* 55(44): 25-27.
75. Ji X, Wang X, Yue XD (2017) Effects of eNOS, CAV1 and PI3K/Akt signalling pathways on homocysteine promoting migration and proliferation of vascular smooth muscle cells in rats. *Chinese General Practice* 20(12): 1469-1473.
76. Dong YX, Li J (2016) Tanshinone I_A inhibits homocysteine-induced proliferation and migration of rat aortic smooth muscle cells and its mechanism. *China Pharmacy* 27(22): 3072-3076.
77. Tan HM, Shi CW, Jiang XH (2014) Hyperhomocysteinemia promotes vascular remodelling in vein graft in mice. *Front Biosci (Landmark Ed)* 19: 958-966.
78. Chernyavskiy I, Veeranki S, Sen U (2016) Atherogenesis: hyperhomocysteinemia interactions with LDL, macrophage function, paraoxonase 1, and exercise. *Ann N Y Acad Sci* 1363(1): 138-154.
79. Zhang MH, Li F, Wang XY, Gong J, Xian Y, et al. (2020) MiR-145 alleviates Hcy-induced VSMC proliferation, migration, and phenotypic switch through repression of the PI3K/Akt/ mTOR pathway. *Histochem Cell Biol* 153(5): 357-366.
80. Zhang MH, Li F, Pokharel S, Ma T, Wang X, et al. (2020) Lycium barbarum polysaccharide protects against Homocysteine-induced Vascular smooth muscle cell proliferation and phenotypic transformation via PI3K/Akt pathway. *J Mol Histol* 51(6): 629-637.
81. Wang XY, Ma X, Zeng Y (2023) Reduced C1q/Tumor Necrosis Factor-Related Protein9 Expression Promotes Hcy-Induced VSMCs Migration via Negative Regulating Endoplasmic Reticulum Stress. *Am J Biomed Sci & Res* 19(1):33-38.
82. Jiang YD, Zhang MH (2015) Hyperhomocysteinemia and cardiovascular disease. *science press (Beijing)* 95-101.
83. Weinert S, Poitz DM, Auffermann-Gretzinger S, Eger L, Herold J, et al. (2013) The lysosomal transfer of LDL/cholesterol from macrophages into vascular smooth muscle cells induces their phenotypic alteration. *Cardiovasc Res* 97(3): 544-552.