



Research Article

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# Development And Assessment of Quality Criteria of Routine Phytosomes in The Treatment of Periodontitis

Sevil Mehraliyeva Jabrail<sup>1\*</sup>, Gasimov Eldar Kochari<sup>2</sup>, Rzayev Fuad Huseynali<sup>2</sup> and Shahla Yusubova<sup>3\*</sup>

<sup>1</sup>PhD, Associate Professor, Pharmaceutical technology and management, Azerbaijan Medical University, Baku

<sup>2</sup>Professor, Head of the Department of Cytology, Embryology and Histology, Azerbaijan Medical University, Baku

<sup>3</sup>PhD, Ass. Prof, Head of Electron Microscopy Department, Scientific Research Center, Azerbaijan Medical University, Baku

<sup>3</sup>Ph.D, Associate Professor, Therapeutic Stomatology, Azerbaijan Medical University, Baku

\*Corresponding author: Sevil Mehraliyeva Jabrail, PhD, Associate Professor, Pharmaceutical technology and management, Azerbaijan Medical University, Baku.

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## Abstract

Currently, one of the priority issues of pharmaceutical nanotechnology is to develop phytosomal gel technology, a new drug delivery system based on rutin in the treatment and prevention of dental diseases. In this regard, it was determined that the size of rutin phytosomes prepared in the Laboratory of Pharmaceutical Technology of Azerbaijan Medical University is 60-180 nm based on SEM, TEM, and other analysis methods. Nanomedicine was prepared by incorporating rutin phytosomes into gel (a mass obtained from alginic acid and polysaccharides of plantain seeds). The innovative gel developed by us based on phytosomes has several advantages, which are significantly different from traditional gel preparations. The phytosomal gel studied at the Department of Therapeutic Dentistry of AMU provided 85-90% natural cell regeneration in the damaged tissue during the treatment of periodontitis. As a result of one-time use of phytosomal gel, it was observed that its effect is long-lasting, even lasting for several years. Also, the effect of the gel restores the barrier function of the periodontal tissue and local immunity, which helps to prevent repeated inflammation of the tissue. The conducted comparative studies once again proved that the phytosomal gel of rutin has a therapeutic effect within 120 hours, while well-known drugs in the treatment and prevention of periodontitis (Metragil, Parodium, Levomekol, Miramistin, etc.) have a therapeutic effect within 7-14 days.

**Keywords:** Periodontitis, Rutin, Phytosomes, Lecithin, Electron Microscopy Analysis, Anti-inflammatory effect

## Introduction

Currently, periodontitis is one of the most common dental diseases and is a social problem, because it leads to disruption of the supporting function of teeth, disruption of chewing function, bleeding gums and, as a result, loss of teeth, and infection of periodontal pockets is negative affects the body as a whole. Almost every person suffers from periodontitis to one degree or another. And periodic bleeding of the gums when brushing your teeth is the first

symptom of a dangerous chronic disease. The cause of the development of periodontitis can be carious teeth, overhanging edges of fillings, traumatic gingival papilla, etc., but the main reason that triggers the development of periodontitis is dental plaque. WHO claims that 10-15% of the world's population is infected with this disease [8]. In modern times, drug delivery systems are widespread. Conducting research in this direction is distinguished



by its relevance. We have conducted research on the production of nanoemulsions, nanocapsules, and niosomal gel with plant and animal extracts, gold, and silver nanoparticles, and obtained appropriate results [1-7].

In addition to the above, the synthesis of various types of nanoparticles, a number of their physicochemical properties, biological activity, antiparasitic effect, application in industry, bioaccumulation in various components of a simple food chain (plants, molluscs, fish, parasites) and pathological changes caused by Transmission Electron Microscope (TEM)) studied the ultrastructural characteristics [9-16]. Phytosomes, which are one of the drug delivery systems, are distinguished by the simplicity of the preparation technology, long-term effect, and stability of the received drug form. The preparation of phytosomal gel based on rutoside is considered a priority in the treatment and prevention of dental diseases. In terms of the lack of long-term effect and side effects, the application of phytosomal gel, a new delivery system, is considered satisfactory in such diseases. Phytosomes have been selected as a new delivery system and are mainly based on plant-derived raw materials. Phytosomes, distinguished by a number of advantages, improve the absorption of biologically active substances, synergistic effect and reduce the need for their amount; increases the absorption of oil-insoluble hydrophilic compounds; increases

the bioavailability of phytoconstituents orally and topically; has the ability to easily pass through the cell membrane and enter the cell; phytosomes exhibit a good stability profile due to the formation of chemical bonds between the phosphatidylcholine molecule and phytoconstituents [19].

Phytosomes are not liposomes and structurally the two are very different (Figure 1). Unlike phytosomes, liposomes are formed by mixing a water-soluble substance with phosphatidylcholine. No chemical bonds are formed, and the phosphatidylcholine molecules surround the water-soluble substance. There may be hundreds or even thousands of phosphatidylcholine molecules surrounding the water-soluble compound.

Unlike the phytosome process, phosphatidyl-choline and the individual plant components are actually in a 1:1 or 2:1 ratio, depending on the substance. The liposomal drug complex is formed in the presence of water or a buffer solution, where the phytosomes are immobilized with a solvent of reduced dielectric constant.

Currently, world scientists have prepared phytosomes from plants such as curcumin, echinacea, ginkgo biloba, licorice, ginseng, silybin, grape seed, olive, green tea, ginger, and rose hips in the field of medicine and cosmetology. These phytosomes have anti-inflammatory, immunomodulatory, anti-aging, anti-cancer and antioxidant effects [17-30] (Figure 1).

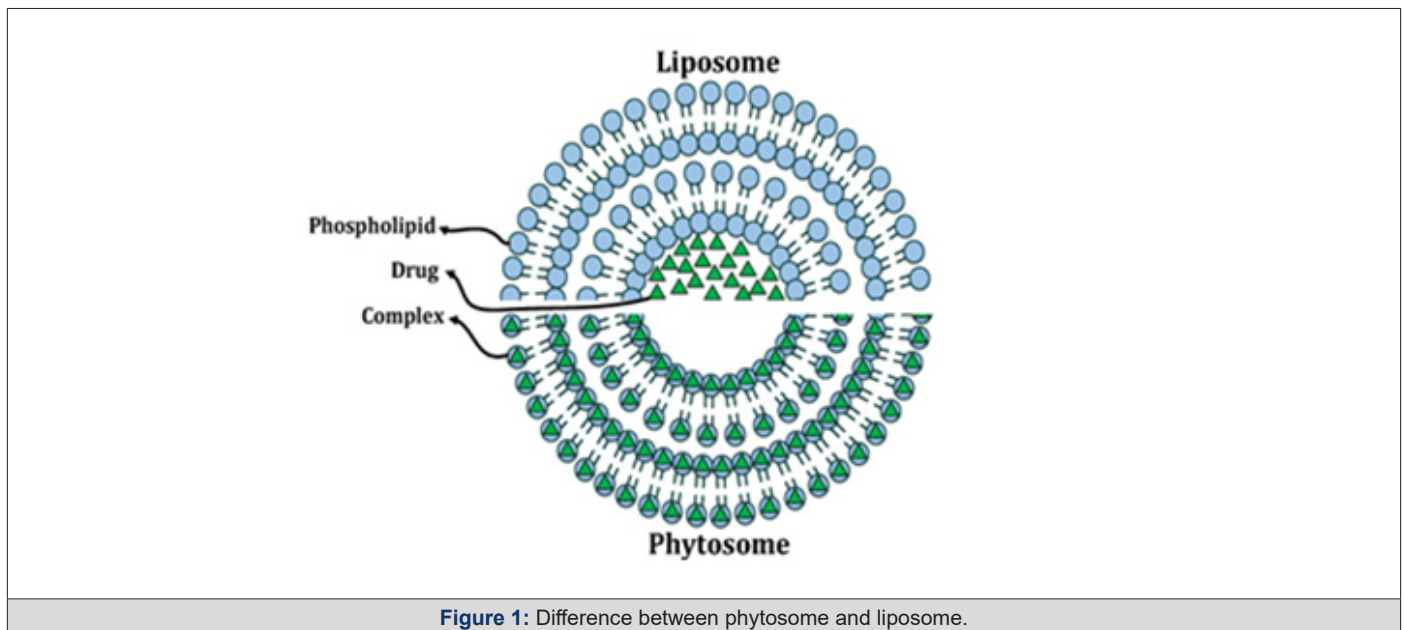


Figure 1: Difference between phytosome and liposome.

Rutoside, which we have chosen as a research object for the treatment of periodontitis, as a result of the literature review, it was found that rutin prevents the accumulation of platelets, reduces capillary permeability, improves blood circulation, and has anti-inflammatory properties. It can be used in the treatment of routine hemorrhoids, varicose veins and microangiopathies. In relatively high doses, rutin increases the absorption of iodine by the thyroid gland, reduces the levels of T3 and T4 hormones. At the same time, it was determined that rutin is more powerful than quercetin, hesperidin and naringin in terms of antioxidant activity [31]. Taking into account the mentioned effective therapeutic properties of rutin, the main goal was to conduct research on the development of phytosomal gel as a transdermal drug delivery system in the treat-

ment of periodontitis.

## Material and Methods

Materials. Rutoside, which was used as a research object, was identified in laboratory conditions from the fruits of Japanese sophora, distributed in Azerbaijan. Phosphatidylcholine (soy lecithin) was obtained from Russia (TS 10.89.19.-029-13578491-2019; 000 Rostov Pharmaceutical factory). Research were carried out in the laboratories of the "Pharmaceutical technology", "Cytology, embryology and histology" and "Therapeutic dentistry" departments of the Azerbaijan Medical University, "Catalysis" and "Geology and Geophysics" Institutes of the Ministry of Science and Education of the Republic of Azerbaijan.

### Preparation Technology

The preparation of rutin phytosomes was carried out with different ratios of rutin and phosphatidylcholine. To do this, transfer the routine powder into a 150 ml flask and add acetone to it. Then phosphatidylcholine is added to a 250 ml flask and a solution of rutoside in acetone is added. The resulting mixture is heated at 65°C for 2 hours under reflux. The solution is concentrated until 10 ml of the original volume remains. After 2 hours, the flask is separated from the refrigerator, cooled to room temperature, and filtered. After evaporating the organic solvent for 1 day, the resulting phytosomes are dried in a desiccator.

### Differential Scanning Calorimetry (DSC)

Interactions in DSC can be observed by comparing transition temperatures, appearance of new peaks, disappearance of original peaks, melting points, and changes in relative peak area. Phyto-phospholipid complexes usually show radically different characteristic peaks than physical mixtures. The studied phytosome was a physical mixture of rutin and PX (1:1). DSC studies of phytosome samples consisting of pure rutin and phosphatidylcholine-rutin mixture were conducted in a Jupiter STA 449 F3 differential scanning calorimeter manufactured by the German company NETZSCH by heating in closed metal pans in the temperature range of 100-750°C under a nitrogen gas environment at a speed of 20°C per minute.

### Transmission Electron Microscopy (TEM)

Samples were dispersed in distilled water and a drop was placed on a carbon coated copper square mesh grid (Electron Microscopy Science, USA). Nanoparticles examples were examined under the Transmission Electron Microscope JEM-1400 (JEOL, Japan) at a voltage of 80-120 kV. The morphometric analysis (Min, Max, mean SD, et al.) of the electronograms was carried out in TIF format via a computer program (TEM Imaging Platform- ITEM) developed by Olympus Soft Imaging Solutions GmbH (Germany).

### Scanning Electron Microscopy (SEM)

Rutin and our prepared phytosome sample were examined by

JSM-6010RV scanning electric microscope manufactured by JEOL, Japan.

### X-Ray Diffraction (XRD)

Research were carried out with rutin and phytosome samples in Cu-K $\alpha$  radiation at angles  $50 \leq 2\theta \leq 750$  with the D2 Phaser Automatic X-ray Diffractometer of the German "Bruker" company.

### Fourier Transform Infrared Spectroscopy (FTIR)

The analysis was performed on a Nicolet IS 10, Smart OMNI TRANS-Mission device. A comparative analysis of the standard and the sample was carried out. Studies were performed with pure rutin and phytosome separately (4000-500 cm<sup>-1</sup>). The obtained IR spectra were analyzed according to functional groups.

## Results and Discussion

### Preparation of Phytosomes

We prepared RN-P with rutin lecithin (phosphatidylcholine) in 3 different ratios (1:1-1:3). Of these, 2 formulations were pale yellow in color and lumpy, meaning they did not flow freely. Sticky masses were felt. Phytosomes had the best fluidity in the 1:1 formula. In this regard, further studies were conducted on phytosomes prepared in a 1:1 ratio. We used acetone as a solvent. Lecithin and rutin were mixed in a ratio of 1:1, acetone was added and extracted under refrigeration. The prepared phytosomes were able to maintain their stability for 2 years.

### Differential Scanning Calorimetry

Sample weight (Phytosome IV) 5.4 mg. Starting from a temperature of 30°C, it is heated up to 750°C with a heating step of 20°C per minute. The atmosphere inside the oven is nitrogen gas. 4 main peaks were observed in this derivatogram. The first peak was observed with a minimum temperature of 147.7°C and a weight loss of 5.93% of the total mass. The second peak at 227.9°C with a weight loss of 36.47% of the total sample weight, the third peak at 372.4°C with a weight loss of 36.47%, and the fourth peak at 637.1°C with a weight loss of 30.70% has been done. As a result of the analysis, it was found that the residual mass is 4.61% of the total weight of the sample (Figures 2,3).

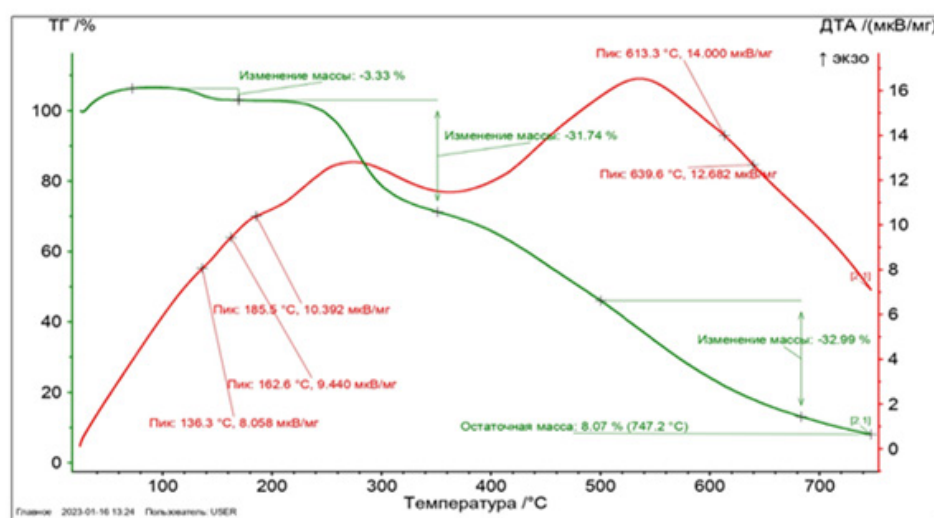


Figure 2: DSC thermogram of pure rutoside.

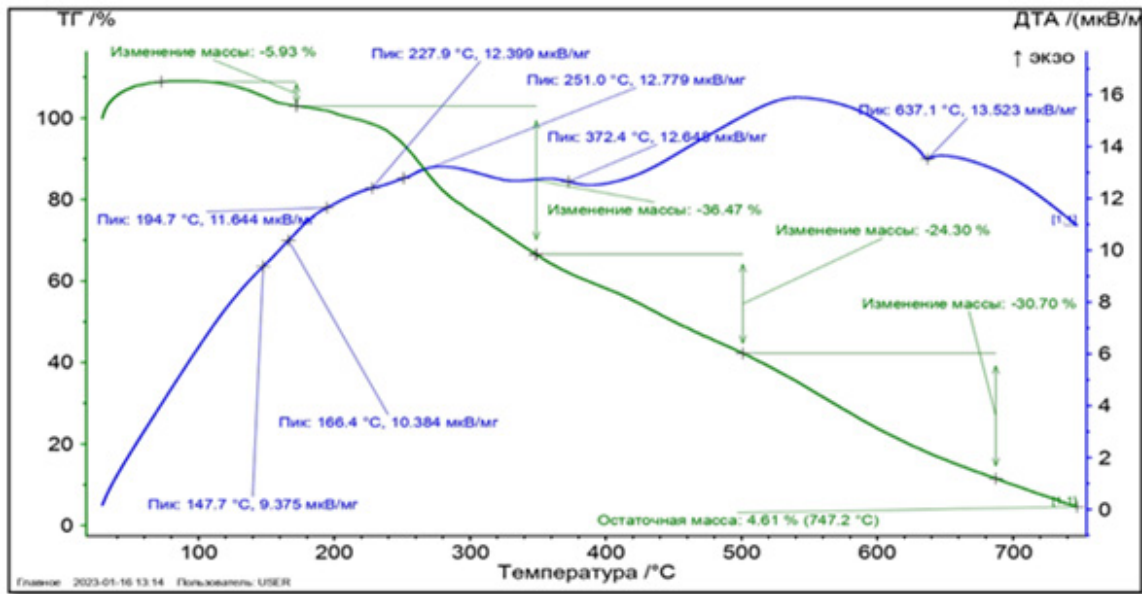


Figure 3: Disintegration temperature of rutin phytosome.

**X-Ray Diffraction (XRD)**

Currently, X-ray diffraction is an effective method for investigating the microstructure of both crystalline materials and some amorphous materials. X-ray diffraction is usually performed on either active components or active ingredient phyto-phospholipid complexes, PCs, and their physical mixtures. X-ray diffraction of the active ingredient and physical mixture shows dense crystalline peaks indicating a highly crystalline form. In contrast, phyto-phospholipid complexes with the active ingredient do not exhibit a crys-

talline peak, indicating that the components complexed with phospholipids are in molecular or amorphous form. This may explain the observation that phyto-phospholipid complexes have better lipophilicity and hydrophilicity than the active components [24]. XRD of RN shows intense crystalline peaks indicating high crystallinity of the drug (Figures 4,5). In our example, as a result of XDR analysis, it was observed that the phytosomes prepared by us have few crystal peaks. As can be seen from Figure 5, rutin phytosomes are distinguished by their high amorphousness.

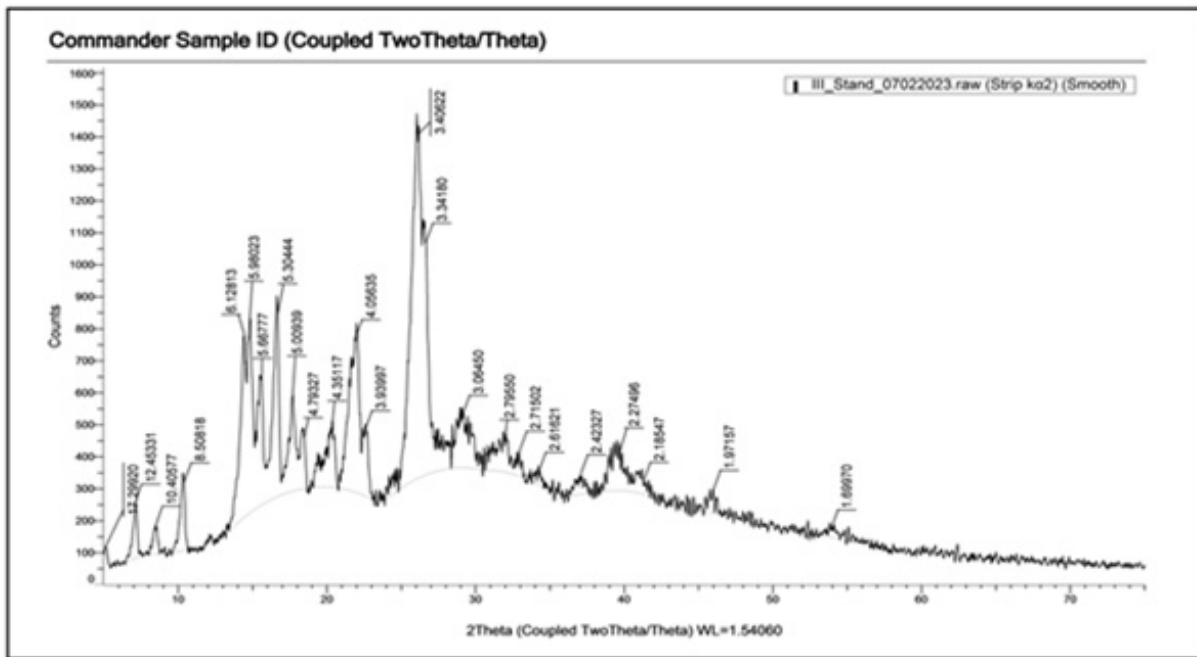


Figure 4: X-Ray spectrum of the standard rutin.

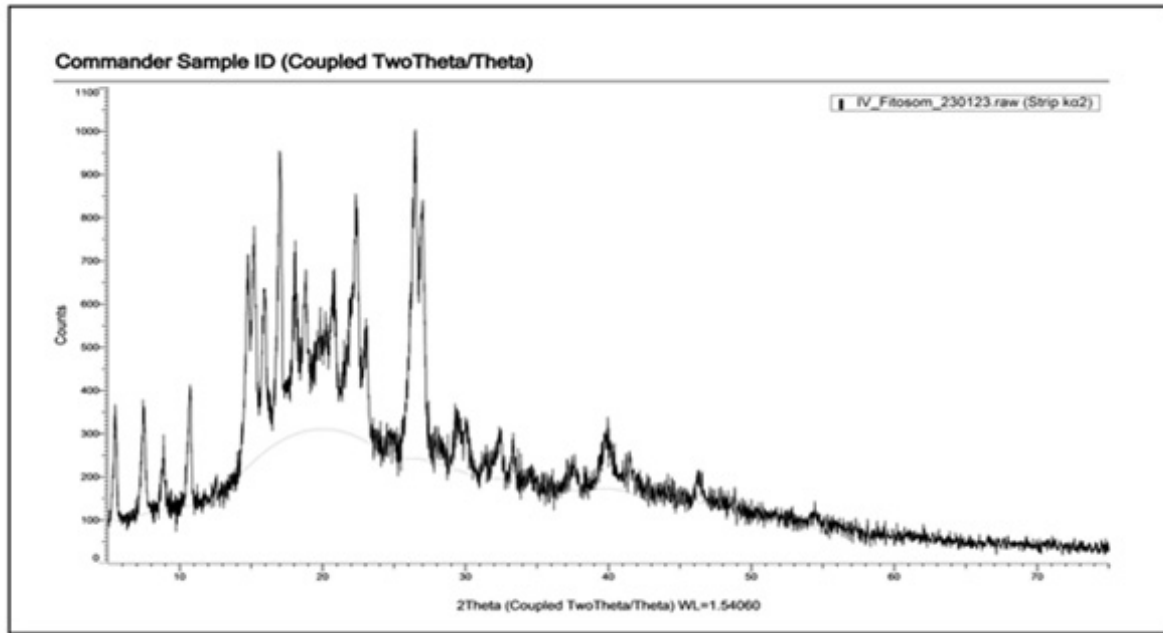


Figure 5: Phytosome X-Ray spectrum.

**Fourier Transform Infrared Spectroscopy (FT-IR)**

FT-IR is a powerful technique for structural analysis and yields a variety of functional groups that exhibit distinct characteristics in band number, position, shape, and intensity. The formation of phyto-phospholipid complexes can be verified by comparing the spectroscopy of phospholipid complexes with the spectroscopy of physical mixtures. Individual studies may show different results. The analysis was performed on a Nicolet IS 10, Smart OMNI TRANS-Mission device. A comparative analysis of the standard and

the sample was carried out. As a result of the analysis, the IR spectrum of standard rutin is 1657 cm<sup>-1</sup> (C=O), 3582.71 cm<sup>-1</sup> and 3446.58 cm<sup>-1</sup> (OH), 2992.57 cm<sup>-1</sup> (-CH<sub>2</sub>), 1598.40, 1574.29, 1557.08 cm<sup>-1</sup> (aromatic ring), 1235, 1297.27, 1204.15 cm<sup>-1</sup> (C-O-C) absorption band (Picture 1), and IR spectra of Phytosome 1482.46; 1504.85 cm<sup>-1</sup> 1598.75; 1657.49 cm<sup>-1</sup> (aromatic ring), 1738.72 cm<sup>-1</sup> (-C=O group), 2922.42-2853.47 cm<sup>-1</sup> (-CH<sub>2</sub>), 3582.72, 3353.42 cm<sup>-1</sup> (OH), 1297.79; It was formed in the absorption band of 1376.64 cm<sup>-1</sup> (C-O-C). (Figures 6,7).

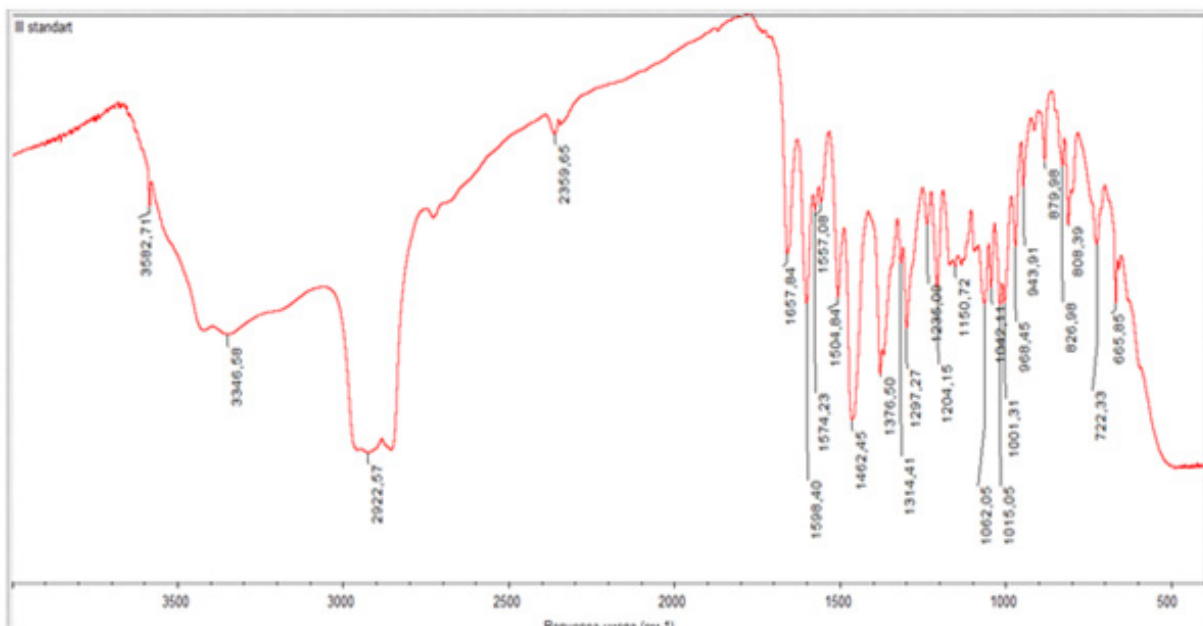


Figure 6: IR-spectrum of the standard rutin.

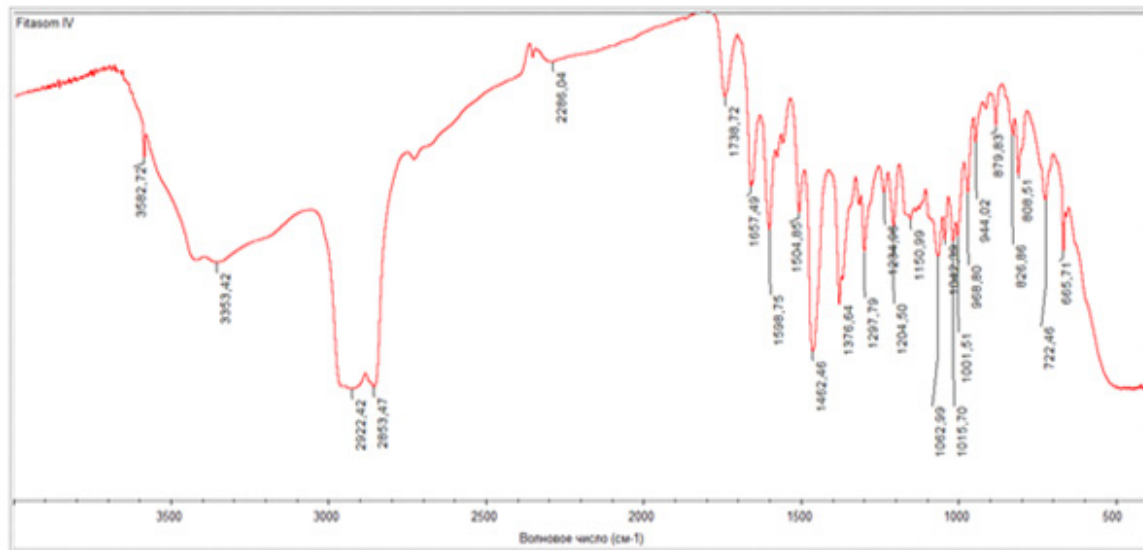


Figure 7: IR spectrum of rutin phytosome.

### SEM Analysis

While the high crystalline state of rutin was observed in image

(A) taken with rutin during the analysis, in image (B) it was observed that rutin had already turned into phytosomes (Figure 8).

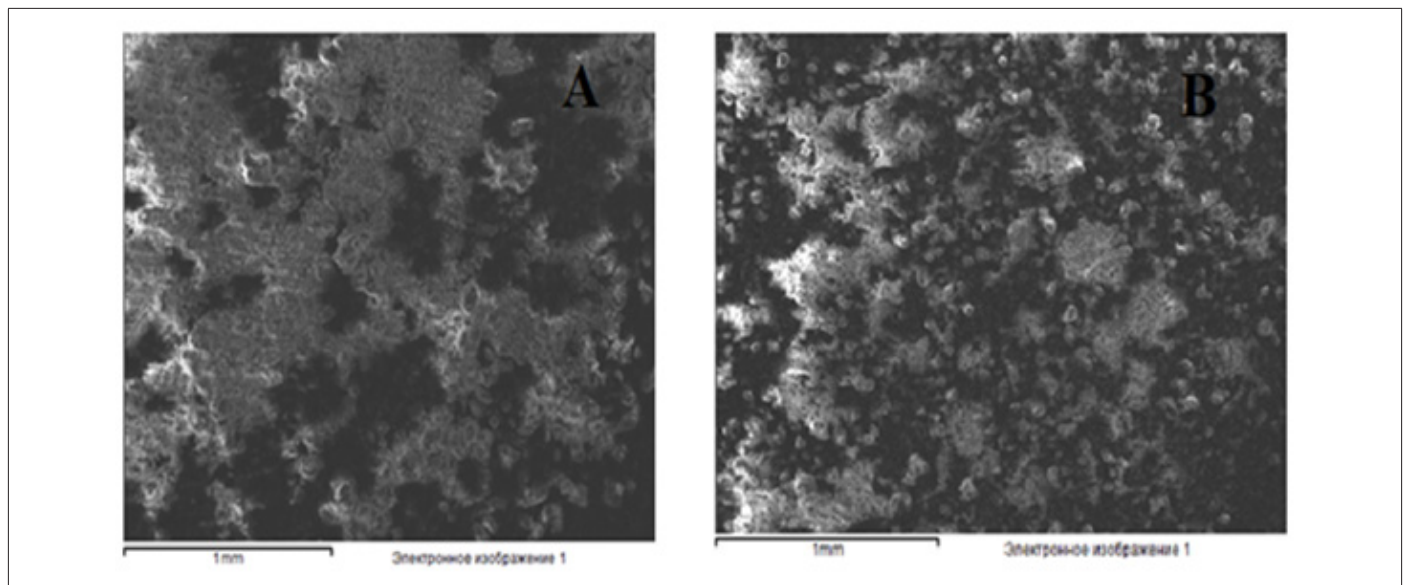


Figure 8: SEM analysis results. A- rutin, B-phytosom.

### Transmission Electron Microscopy (TEM)

TEM studies reveal the formation of a dark discrete structure that appears as uneven spheres. When dispersed in water by gentle shaking, the phytosomes self-arranged in response to surface tension. As a result of the analysis of electrograms obtained from the large magnifications (40000-60000) of TEM, images of phytosomes up to the size of 60-180 nm ( $103 \pm 1,12$  nm) were observed (Figure 9A and 9B).

### Solubilization of Phytosomes

For this purpose, the solubility of phytosomes was checked in weak alkaline environment (pH-6.7-7.4) and acidic environment (pH-1.5-1.8). As a result, it was observed that phytosomes are bet-

ter dissolved in a weak alkaline environment, which provides the basis for ensuring the local effect of the drug in the oral cavity.

### Productivity of Phytospholipid Complexes (Degree of Complexation)

The formula is as follows:

$$\text{Yield(\%)} = [(a-b)/a] \times 100\%$$

While "a" is the weight or composition of the primary active component, "b" denotes the weight or composition of the free active component, and "(a - b)" denotes the weight or composition of phospholipid complexes [22]. As a result of the calculations, the surface coverage of rutin with phosphatidylcholine was 99.83-100%.

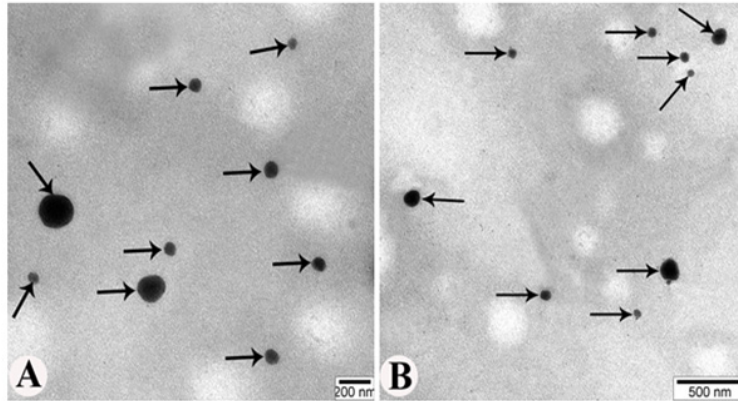


Figure 9: TEM photographs of phytosomes.

**Application of Phytosomal Gel in the Treatment of Periodontitis**

Patient F.A., 30 years old. clinic periodontal pocket. The PMA index (papillary-marginal-alveolar) was 19.4%, The Papillary Bleeding Index (PBI) was 2.5 points, and the Periodontal Index (PI) was

1.69 points. An application of phytosomal gel (experiment) in the amount of 0.5 ml was applied to the affected gum twice with an interval of 2 days. After 3 days, a decrease in gum inflammation was determined: PBI index - 0 points (no bleeding gums); PMA index - 0; PI - 0 points. The treatment period was 3-5 days.



Figure 10: Treatment with rutin phytosomal gel.

Patient N.Sh. 37 years. The indicators of the PMA index were 35.4%, the PBI index - 0.75 points, and the PI index - 1.67 points. An antimicrobial gel (metragil denta) in an amount of 0.5 ml was applied to the affected gum twice with an interval of 2 days. After 5 days, a decrease in gum inflammation was observed, a decrease

in the PMA index was noted to 10.5%, the PBI index to 0.12 points, and PI to 0.15 points. The gel was reapplied. After 7 days, a persistent decrease in gum inflammation was determined (PMA index - 2%; PBI index - 0.05 points; PI index - 0 points).

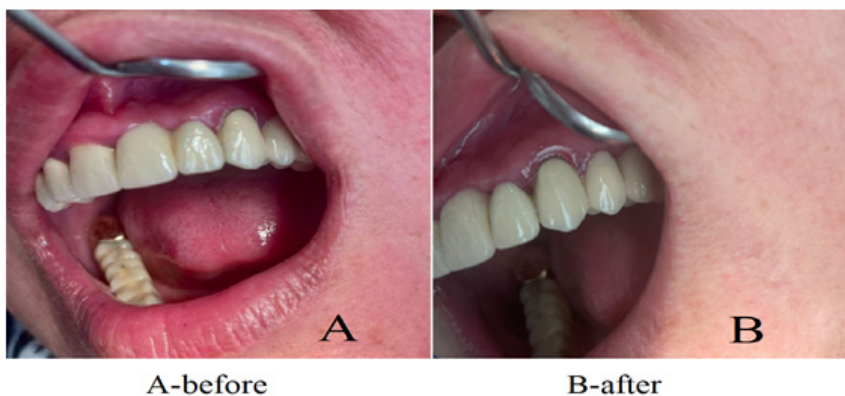


Figure 11: Treatment with Metrogil.

**Conslusions**

Phytosomes are of interest to researchers as a widespread innovative delivery system in modern times. Coating the surface of many polyphenol compounds with soy and egg lecithin ensures

their local, long-term effect. Since the absorption of rutin is difficult, the development of its transdermal form is considered one of the priority issues in pharmaceutical nanotechnology. Taking this into account, we developed a dental delivery system in our research. As

a result of the conducted experiments, it was determined that rutin phytosomes have high bioavailability in the treatment of periodontitis and have an effective anti-inflammatory effect.

For this purpose, phytosomes based on rutin and lecithin were first prepared. X-Ray Diffraction (XRD), Fourier Transform Infrared Spectroscopy (FT-IR), Differential scanning calorimetry, SEM analysis, solubility test were performed to determine the vesicular size. TEM micrograph confirmed the structure of phytosomes (60-180 nm). Gel of rutin phytosomes is prepared on the basis of alginic acid and plantain polysaccharides. A gel made with rutin phytosomes was tested for better penetration. Phytosome gel has been found to be a good carrier for topical delivery of rutin. This fact has been proven once again when applied in the initial stage of periodontitis. It is considered appropriate to conduct extensive research in this direction.

## Acknowledgments

None.

## Conflict of Interest

None.

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